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Problem formulation for the ERA of insecticidal RNAi-based genetically modified plants and RNAi-based pesticides: Effects on non-target arthropods

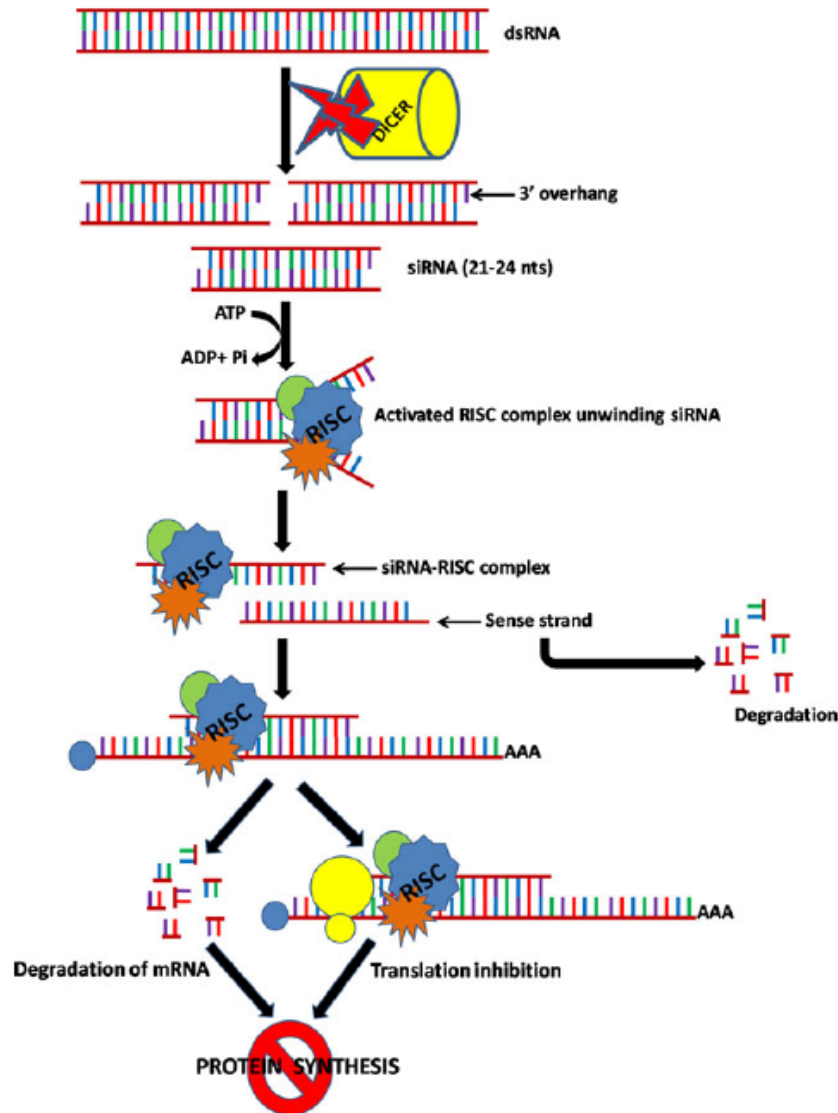
Jörg Romeis

Agroscope, Zurich, Switzerland





RNAi – how does it work ?



Endogenous **sequence-specific gene silencing** mechanism elicited by small RNA molecules

- dsRNA is ingested
- Processed by DICER into siRNA (21-25 bp)
- Incorporated into RISC
- siRNA-RISC complex **targets similar sequence** in mRNA (degradation or inhibition of translation)
- **Protein synthesis is blocked**

Problem formulation

What do we not want to see harmed? What must be protected?

➔ *Protection goals*

Can we envision a way in which they could be harmed?

➔ *Pathway to harm*

How can we assess whether they are likely to be harmed?

➔ *Development of risk hypotheses and a plan to test them*



Protection goals

Millennium Ecosystem Assessment

Published online: August 11, 2015

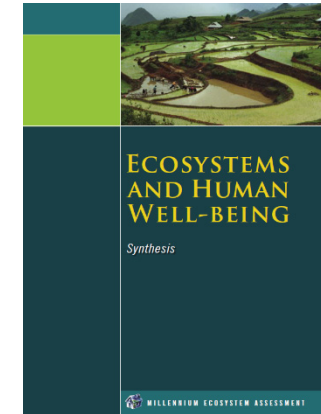
Science & Society

EMBO
reports

Optimising environmental risk assessments

Accounting for ecosystem services helps to translate broad policy protection goals into specific operational ones for environmental risk assessments

Yann Devos¹, Jörg Romeis², Robert Luttik³, Angelo Maggiore⁴, Joe N Pery⁵, Reinhilde Schoonjans⁴, Franz Streissl⁶, José V Tarazona⁶ & Theo CM Brock⁷



Regulating services

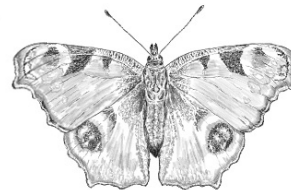
- **Biological control** of arthropod pests
- Pollination

Cultural services

- Protected butterflies

Supporting services

- Nutrient cycling, decomposition





Pathway to harm



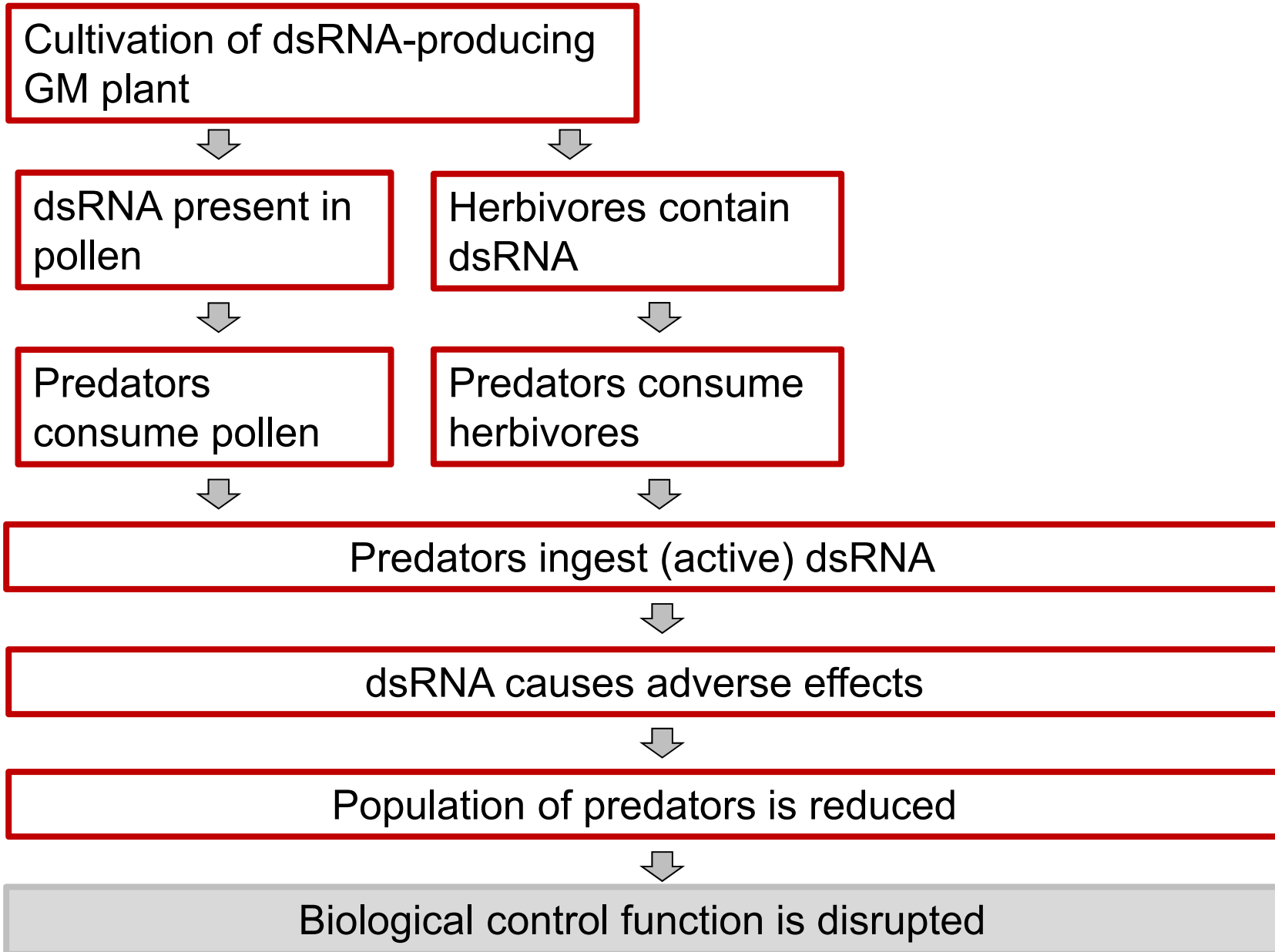
Cultivation of dsRNA-producing GM plant

Application of dsRNA-containing spray product

Biological control function is disrupted

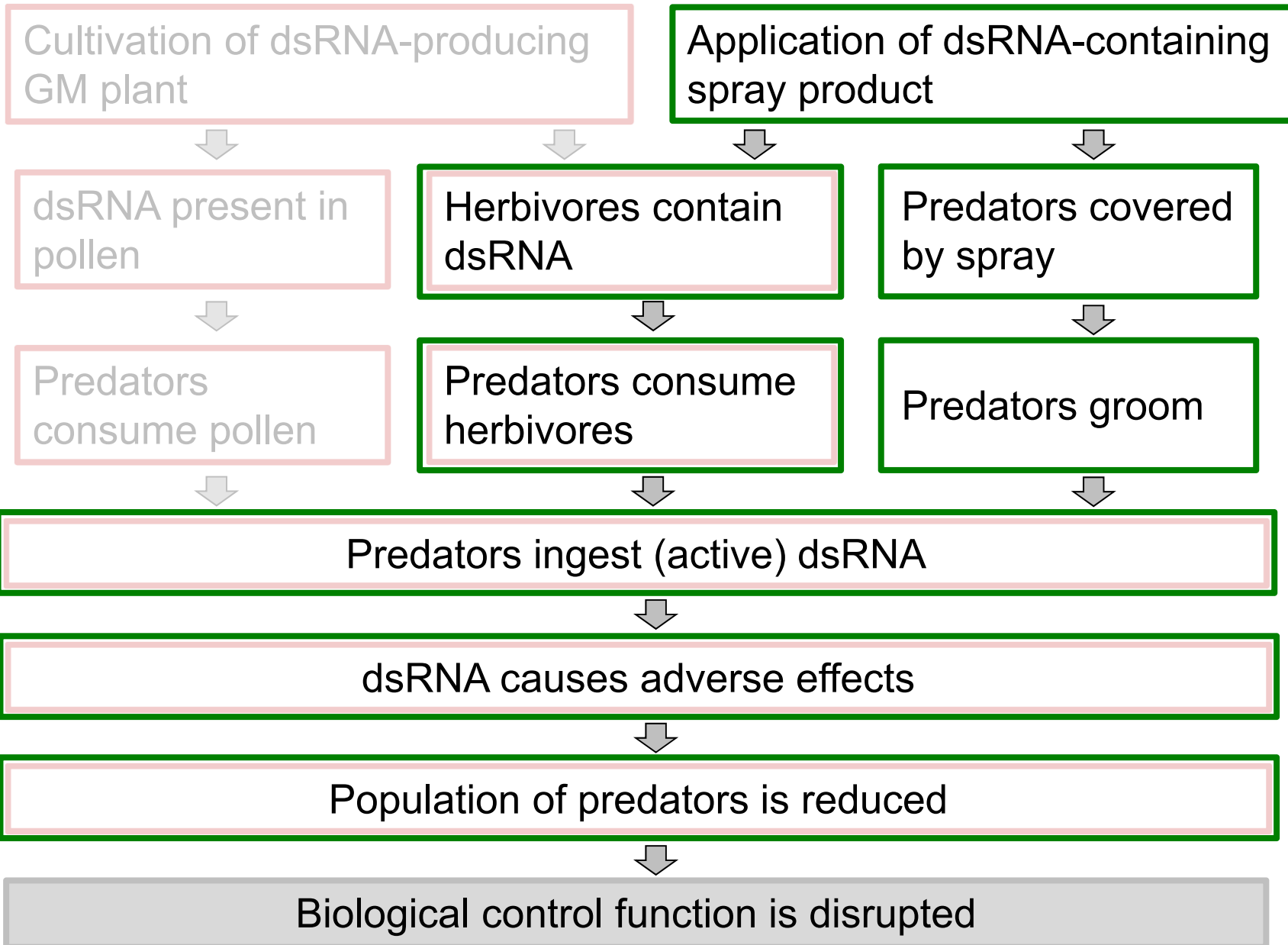


Pathway to harm





Pathway to harm





Testable risk hypotheses



Cultivation of dsRNA-producing GM plant

Application of dsRNA-containing spray product

Focus Exposure

- dsRNA molecule is not produced in plant pollen
- Predators do not consume plant pollen
 - Predators not covered by spray
- ● Herbivores do not contain (active) dsRNA

Focus Hazard/ effect

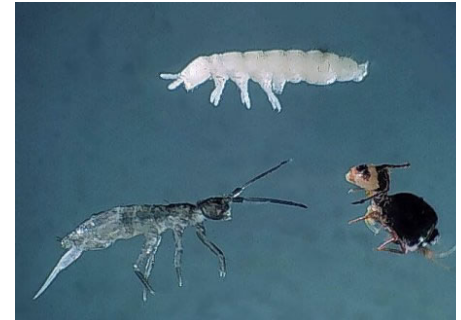
- ● Predators are not affected by consumed dsRNA
- ● Effects do not result in population declines
- ● Population declines do not lead to a disruption of the biological control function





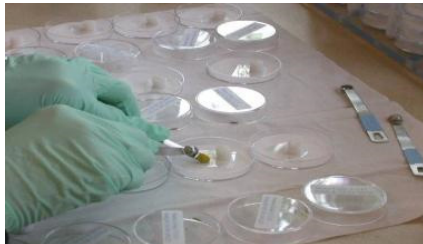
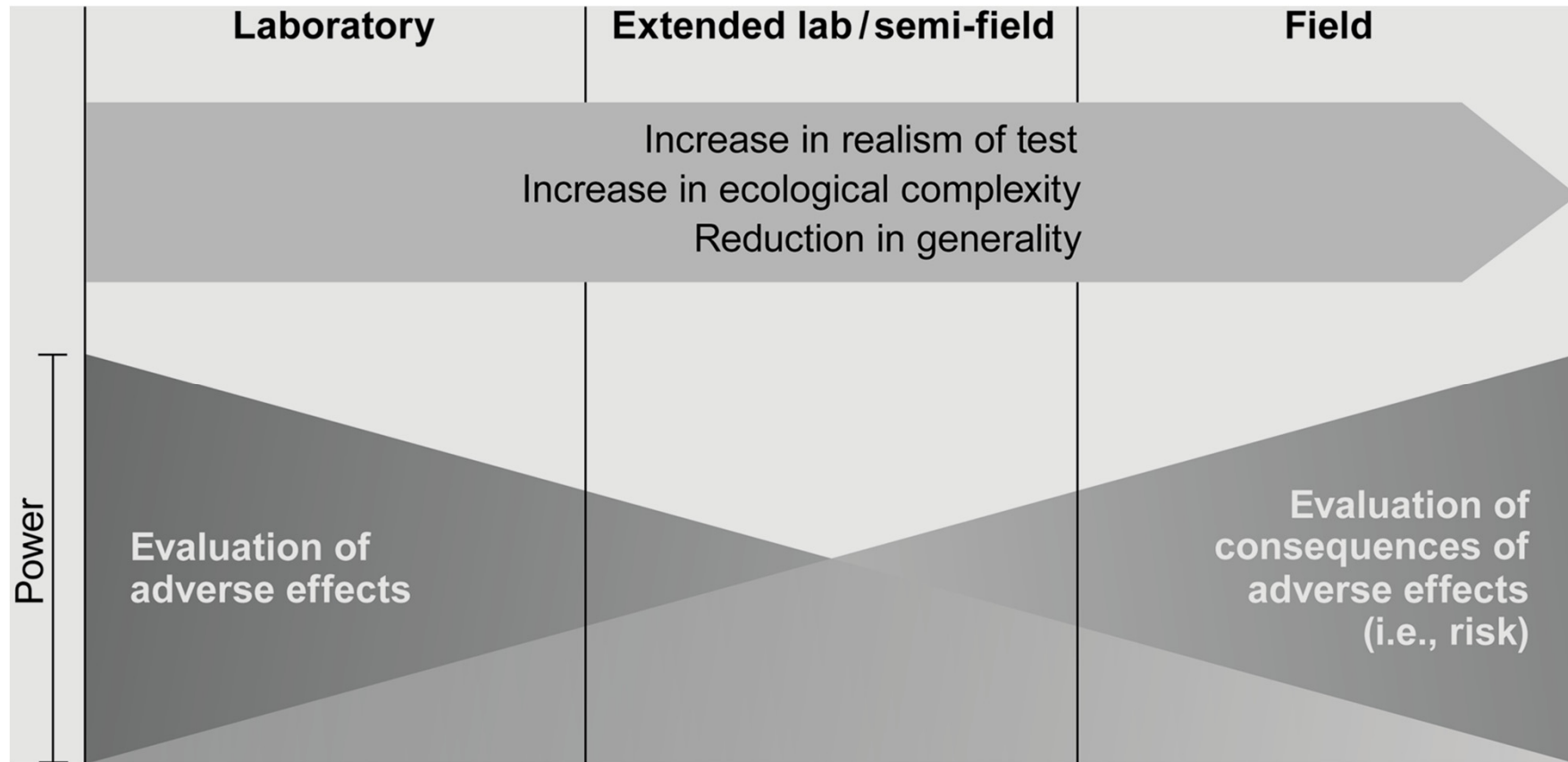
Testable risk hypothesis

"The dsRNA does not affect valued non-target arthropods at the concentration present in the field"





Tiered risk assessment





Ingested dsRNA could cause ...

- off-target effects
 - Silencing of any gene in a non-target organism
- general immune stimulation
- saturation of the RNAi machinery as consequence of the ingestion of high doses of dsRNA

- Conduct feeding studies with dsRNA, similar to insecticidal Cry proteins
- Consider mortality and sublethal endpoints



Selection of test species for GMP



EFSA (2010) “Considering that not each of these species can be tested, a representative subset of NTO species [...] shall be selected, on a case-by-case basis.”

- **representative of valued taxa and functional groups that are most likely to be exposed**
- **species most likely to be sensitive to the test compound** (*considering mode of action, known toxicity*)
- **Amenability and availability** to testing



Selection of test species for GMP



NTOs most likely to be sensitive:

- Arthropod orders differ in their sensitivity to dietary RNAi. Coleoptera are more sensitive than other insect orders (*Baum and Roberts, 2014, Adv. Insect Physiol. 47, 249-295*)
- **Pylogenetic relationship** of NTO to target is important (*Bachmann et al., 2013, Transgenic Res. 22, 1207-1222*)
- **Bioinformatic analyses** help predict NTO effects: analyse sequence complementarity between pool of siRNA and genome or (target) gene in NTO (*Roberts et al., 2015, Front. Plant Sci. 6, 958*)



Selection of test species for PPP



- Data requirements are provided in Commission Regulation (EU) No 283/2013 and 284/2013 for the approval of *active substances* and the authorisation of *plant protection products*
- Commission Regulation (EU) 546/2011 and Regulation (EC) 1107/2009 of the European Parliament and of the Council define uniform principles for evaluation and authorisation of plant protection products incl. defined set of test parameters and endpoints!
- SANCO, Draft Guidance Document on Terrestrial Ecotoxicology, 2002
- ESCORT 2, SETAC, Guidance Document on Regulatory Testing and Risk Assessment Procedures for Plant Protection products with Non-target Arthropods, 2001



Selection of test species for PPP



Fixed set of NTO test organisms

(birds, mammals, earthworms, fish, aquatic invertebrates)

Terrestrial arthropods

- Honey bees (pollinator; Hymenoptera)
- **Beneficial arthropods other than bees**
 - *Aphidius rhopalosiphi* (parasitoid; Hymenoptera)
 - *Typhlodromus pyri* (predatory mite; Acarina)





Selection of test species for PPP



EFSA Journal 2015;13(2):3996

SCIENTIFIC OPINION

Scientific Opinion addressing the state of the science on risk assessment of plant protection products for non-target arthropods¹

EFSA Panel on Plant Protection Products and their Residues (PPR)^{2,3}

- **Address oral exposure** of NTAs to PPP
- Difficult to relate currently available **endpoints** from tier 1 assessments (glass-plate tests) to realistic exposure
- **Standardised tests** for addressing oral exposure are missing
- Test systems should
 - cover **exposure routes** particular to the active substance
 - allow detection of effects resulting from **specific/ novel modes of action**



Conclusions

- Problem formulation helps to focus the risk assessment
- Regulatory framework established for GMPs works also for plants producing dsRNA
- Case-by-case approach allows to select most appropriate test species
- Test systems for oral exposure are available
- To detect non-target effects lethal and sublethal endpoints should be recorded
- The established NTO test list for PPP is not sufficient to test for non-target effects of dsRNA spray products



Thank you for your attention



Drawing by Simone Haller