

ZOONOSES MONITORING

Iceland

TRENDS AND SOURCES OF ZOONOSES AND ZOONOTIC AGENTS IN FOODSTUFFS, ANIMALS AND FEEDINGSTUFFS

including information on foodborne outbreaks, antimicrobial resistance in zoonotic and indicator bacteria and some pathogenic microbiological agents

IN 2016

PREFACE

This report is submitted to the European Commission in accordance with Article 9 of Council Directive 2003/99/ EC*. The information has also been forwarded to the European Food Safety Authority (EFSA).

The report contains information on trends and sources of zoonoses and zoonotic agents in Iceland during the year 2016.

The information covers the occurrence of these diseases and agents in animals, foodstuffs and in some cases also in feedingstuffs. In addition the report includes data on antimicrobial resistance in some zoonotic agents and indicator bacteria as well as information on epidemiological investigations of foodborne outbreaks. Complementary data on susceptible animal populations in the country is also given. The information given covers both zoonoses that are important for the public health in the whole European Union as well as zoonoses, which are relevant on the basis of the national epidemiological situation.

The report describes the monitoring systems in place and the prevention and control strategies applied in the country. For some zoonoses this monitoring is based on legal requirements laid down by the European Union legislation, while for the other zoonoses national approaches are applied.

The report presents the results of the examinations carried out in the reporting year. A national evaluation of the epidemiological situation, with special reference to trends and sources of zoonotic infections, is given. Whenever possible, the relevance of findings in foodstuffs and animals to zoonoses cases in humans is evaluated.

The information covered by this report is used in the annual European Union Summary Reports on zoonoses and antimicrobial resistance that are published each year by EFSA.

^{*} Directive 2003/ 99/ EC of the European Parliament and of the Council of 12 December 2003 on the monitoring of zoonoses and zoonotic agents, amending Decision 90/ 424/ EEC and repealing Council Directive 92/ 117/ EEC, OJ L 325, 17.11.2003, p. 31

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1 ANIMAL POPULATIONS

The relevance of the findings on zoonoses and zoonotic agents has to be related to the size and nature of the animal population in the country

1.1 Populations

1.1.1 Information on susceptible animal population

Sources of information

Information on cattle, pigs, horses, sheep and goat population are collected from the livestock database BUSTOFN. Data from poultry production is collected both from this database and from registrations at MAST on authorized premises. MAST is responsible for the database. Information regarding slaughtered animals is based on data from the slaughterhouses. The data is stored in databases hosted at the Farmers Association but under surveillance of MAST.

Dates the figures relate to and the content of the figures

The number livestock population in the report are collected from an autumn report that all livestock owners have to turn in before the end of November each year. The database is based on information which farmers have to turn in according to law. MAST reviews the autumn reports before official numbers are released in May the following year. The information for 2016 is reported in autumn that same year. The number of slaughtered animals indicates all slaughtered animals in slaughterhouses from 1st of January 2015 to 31st of December 2015.

Definitions used for different types of animals, herds, flocks and holdings as well as the types covered by the information

The number of poultry flocks is given by the number of houses. The number of holdings and herds are the same for cattle, pigs, horses, sheep and goats.

National evaluation of the numbers of susceptible population and trends in these figures

When comparing animal population from 2015 and 2016 the population is relatively stable. There has been an increase in the number of slaughtered cattle, lambs and poultry but a decrease of slaughtered pigs.

Geographical distribution and size distribution of the herds, flocks and holdings

All existing animal groups in Iceland are relatively evenly spread around the agricultural lowland areas. There are no herds or holdings in the highlands, which cover over 80 % of the island. In the summer, from June to September, the flocks of sheep and herds of horses are grazing in the highlands.

2 DISEASE STATUS

2.1 TUBERCULOSIS, MYCOBACTERIAL DISEASES

2.1.1 General evaluation of the national situation

2.1.1.1 Mycobacterium tuberculosis complex (MTC) - general evaluation

History of the disease and/or infection in the country

Mycobacterium bovis is almost unknown in Iceland. The only clinical case in cattle was detected in 1922. Sporadic monitoring from 1923 until 1971, all negative, and there are no positive findings in slaughterhouses. Passive monitoring at slaughterhouse level today.

2.1.2 Mycobacterium in animals

2.1.2.1 Mycobacterium tuberculosis complex (MTC) in animal - Cattle (bovine animals) - animal sample

Status as officially free of bovine tuberculosis during the reporting year

The entire country free

The entire country is free.

2.2 BRUCELLOSIS

2.2.1 Brucella in animals

2.2.1.1 B. abortus in animal - Cattle (bovine animals) - animal sample

Status as officially free of bovine brucellosis during the reporting year

The entire country free

The entire country is free of bovine brucellosis.

Monitoring system

Sampling strategy

Randomly selection of 16 herds, from each herd 5 blood samples are taken. In 2016, 82 samples were taken from 16 herds.

Frequency of the sampling

Samples are usually taken every year, but some years like 2015, no samples were taken.

Type of specimen taken

Serum

Diagnostic/analytical methods used

Brucella abortus (agg) antibody examination by agglutination. Neg: less than 30iu/ml

Control program/mechanisms

The control program/strategies in place

Annual serological surveillance of dairy cow herds, and annually are taken ca 80 samples.

Measures in case of the positive findings or single cases

Isolation of the farm and attempts to eradication of the agent.

Notification system in place

Brucellosis in animals is notifiable.

Results of the investigation

No findings.

National evaluation of the recent situation, the trends and sources of infection

Neither Brucella abortus nor other Brucella spp. has ever been detected in Iceland.

2.2.1.2 B. melitensis in animal - Goats - animal sample

Status as officially free of caprine brucellosis during the reporting year

The entire country free

The entire country is free of caprine brucellosis.

2.2.1.3 B. melitensis in animal - Sheep - animal sample

Status as officially free of ovine brucellosis during the reporting year

The entire country free

The entire country is free of ovine brucellosis

Monitoring system

Sampling strategy

Samples are randomly taken from 5 ewes on around 20 randomly selected farms.

Frequency of the sampling

Annually.

Type of specimen taken

Serum.

Diagnostic/analytical methods used

Brucella melitensis antibody examination by agglutination

Control program/mechanisms

The control program/strategies in place

Annual serological surveillance and anually are taken ca 100 samples.

Measures in case of the positive findings or single cases

Isolation of the farm and attempts to eradication of the agent.

Notification system in place

Brucella melitensis in animals is a notifiable disease.

Results of the investigation

No findings.

3 INFORMATION ON SPECIFIC ZOONOSES AND ZOONOTIC AGENTS

Zoonoses are diseases or infections, which are naturally transmissible directly or indirectly between animals and humans. Foodstuffs serve often as vehicles of zoonotic infections. Zoonotic agents cover viruses, bacteria, fungi, parasites or other biological entities that are likely to cause zoonoses.

3.1 SALMONELLOSIS

3.1.1 General evaluation of the national situation

3.1.1.1 Salmonella - general evaluation

History of the disease and/or infection in the country

Salmonella is present in the pig production. After 2000, prevalence of Salmonella has been below 1% in broiler flocks and has even been 0% for three consecutive years (2005-2007). In 2009 however, an increase in broiler flocks was seen. Salmonella in breeding flocks (Gallus gallus) has only been once confirmed in the last decade, in 2013 (S. Agona). The same applies to flocks of laying hens, salmonella has only been confirmed once, in 2010 (S. Rissen). Salmonella infections in sheep holdings have not been investigated but clinical outbreaks have sporadically occurred. A study in 2010 on Salmonella in bovine animals revealed very low prevalence (less than 0,5%) with no samples positive. As for sheep, sporadic clinical cases and outbreaks of Salmonella have occurred in horses. Spread of Salmonella into the pig production has likely occurred by raw feed materials. PFGE investigations indicate that Salmonella Worthington has spread into poultry production with compound feed.

National evaluation of the recent situation, the trends and sources of infection

Salmonella is present in broiler flocks, although at low levels, and S. Enteritidis and S. Typhimurium have not been detected in poultry production. The prevalence of salmonella in swab samples from carcasses remained low.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

Because of the control programmes for reducing Salmonella in the pig and poultry production, domestic food products of animal origin are considered to represent a small risk to the consumer in regard to Salmonella. This assumption is supported by the experience in 2009 and 2010 where the prevalence of Salmonella in pig and poultry meat production rose significantly but where the prevalence of human cases of salmonellosis did not alter in the same direction.

3.1.2 Salmonella in foodstuffs

3.1.2.1 Salmonella in food - Meat from bovine animals - food sample

Monitoring system

Sampling strategy

At slaughterhouse and cutting plant

Sampling according to 2073/2005 by FBO is verified under the regular offical control of the FBO.

At meat processing plant

Sampling according to 2073/2005 by FBO is verified under the regular offical control of the FBO.

At retail

No official sampling.

Frequency of the sampling

At slaughterhouse and cutting plant

The frequency of sampling by FBO is proportional to the slaughterhouse's throughput. Slaughtering of beef and horses. More than 10.000 carcasses / year; 5 samples every second week. 2000- 10000 carcasses; /year 5 samples per month. 500-2000 carcasses / month 5 samples 4 times a year. Less than 500 carcasses no sampling if GHP can be demonstrated. Slaugtering of lamb / sheep. More than 100.000 carcasses / slaughtering periode of 6- 8 weeks; 5 samples 5 times during the periode. 50.000- 100.000 carcasses / slaughtering periode of 6- 8 weeks; 5 samples 4 times during the periode. 10.000- 50.000 carcasses / slaughtering periode of 6- 8 weeks; 5 samples 4 times during the periode. 10.000- 50.000 carcasses / slaughtering periode of 6- 8 weeks; 5 samples 4 times during the periode. 10.000- 50.000 carcasses / slaughtering periode of 6- 8 weeks; 5 samples 4 times during the periode. 10.000- 50.000 carcasses / slaughtering periode of 6- 8 weeks; 5 samples 4 times during the periode. 10.000- 50.000 carcasses / slaughtering periode of 6- 8 weeks; 5 samples 4 times during the periode. 10.000- 50.000 carcasses / slaughtering periode of 6- 8 weeks; 5 samples 4 times during the periode. 10.000- 50.000 carcasses / slaughtering periode of 6- 8 weeks; 5 samples 4 times during the periode. 10.000- 50.000 carcasses / slaughtering periode of 6- 8 weeks; 5 samples 4 times during the periode. 10.000- 50.000 carcasses / slaughtering periode of 6- 8 weeks; 5 samples 4 times during the periode. Less than 10.000 no sampling if GHP can be demonstrated.

At meat processing plant

The frequency of sampling of minced meat and meat preparation by the FBO is according to the production capacity per week. More than 3 tons / week; 5 samples per week. 1 - 3 tons per week; 5 samples every second week. 500 kg - 1 ton; 5 samples 4 times per year. Frequency can be reduced when results have been negative for 30 weeks. Processing plants producing less the 500 kg / week does not need to take samples if they can demonstrate good hygiene practices (GHP).

At retail

The local health authorities (LCAs) are responsible for sampling at retail. No official sampling 2016.

Type of specimen taken

At slaughterhouse and cutting plant

Surface of carcass

At meat processing plant

Each sample is 25 g of minced meat or meat preparations.

Methods of sampling (description of sampling techniques)

At slaughterhouse and cutting plant

Swab samples from carcasses before chilling in four designated areas with a sponge. Each area 10×10 cm; total 400 cm2. Swabs of 5 carcasses can be pooled to one sample.

At meat processing plant

Each sample is 25 g of minced meat or meat preparations. Samples can be pooled into one sample (225 g)

At slaughterhouse and cutting plant

Sample is considered to be positive when Salmonella spp. is detected.

At meat processing plant

Sample is considered to be positive when Salmonella spp. is detected.

Diagnostic/analytical methods used

At slaughterhouse and cutting plant

NMKL No 71:1999, 5th edition

At meat processing plant

NMKL No 71:1999, 5th edition

Preventive measures in place

HACCP procedures are verified under official control of FBO.

Measures in case of the positive findings or single cases

Meat is withdrawn from the market.

Notification system in place

Laboratories as well as the FBO analysing Salmonella notify any detection of Salmonella to MAST. Salmonella is a notifiable disease, according to national legislation on animal diseases No. 25/1993 and according to the national Food Law No. 93/1995. MAST receives all positive results from the respective laboratories regarding Salmonella testing (including serotyping and antimicrobial resistance) on samples from FBO own checks in slaughterhouses, cutting plants and processing plants.

Results of the investigation

The negative results from the FBO own checks are not collected. Salmonella in food is notifiable. Salmonella in bovine meat production has not been notified by FBO to MAST.

National evaluation of the recent situation, the trends and sources of infection

Bovine meat is not believed to be a important source of human salmonellosis.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

Salmonella in bovine meat seems not to be a source for human infections.

3.1.2.2 Salmonella in food - Meat from broilers (Gallus gallus) - food sample

Monitoring system

Sampling strategy

At meat processing plant

No official sampling. Sampling according to 2073/2005 by FBO is verified under the regular offical control.

At retail

The local competent authorities (LCA) are responsible for sampling at retail. No sampling in 2016

Frequency of the sampling

At meat processing plant

The frequency of sampling of minced meat and meat preparation by the FBO is according to the production capacity per week. More than 3 tons / week; 5 samples per week. 1 - 3 tons per week; 5 samples every second week. 500 kg - 1 ton; 5 samples 4 times per year. Frequency can be reduced when results have been negative for 30 weeks. Processing plants producing less the 500 kg / week does not need to take samples if they can demonstrate good hygiene practices.

Type of specimen taken

At meat processing plant

Minced meat and meat preparation

Methods of sampling (description of sampling techniques)

At meat processing plant

Each sample is 25 g of minced meat or meat preparations.

Definition of positive finding

At meat processing plant

Sample is considered to be positive when salmonella spp. is detected.

Diagnostic/analytical methods used

At meat processing plant

NMKL 71, 5th ed., 1999

Preventive measures in place

All broiler flocks are sampled before slaughter and salmonella positive flocks are destroyed since it is prohibited to distribute meat from positive flocks. HACCP activities are verified during official control. According to national legislation there is a warning note on each package containing raw poultry meat. This note recommends the consumer to prevent cross contamination when handling raw poultry meat.

Control program/mechanisms

The control program/strategies in place

As described before

Measures in case of the positive findings or single cases

When Salmonella is detected in a slaughter batch or in a product, the batch is withdrawn from the market.

Notification system in place

The FBO and the laboratory have to report a detection of salmonella spp. immediately to MAST.

National evaluation of the recent situation, the trends and sources of infection

There are no changes in prevalence of salmonella in slaughter batches.

3.1.2.3 Salmonella in food - Meat from broilers (Gallus gallus) - food sample - neck skin

Monitoring system

Sampling strategy

At slaughterhouse and cutting plant

The FBOs are sampling each slaughter batch.

Frequency of the sampling

At slaughterhouse and cutting plant

Each broiler slaughter batch is sampled

Type of specimen taken

At slaughterhouse and cutting plant

Neck skin

Methods of sampling (description of sampling techniques)

From each slaughter batch, 50 neck skin samples, each sample consisting of 2-3 g, are pooled into one sample. At the laboratory, a subsample of 25g is taken for further culturing.

Definition of positive finding

At slaughterhouse and cutting plant

A sample positive for Salmonella spp.

Diagnostic/analytical methods used

At slaughterhouse and cutting plant

Bacteriological method: NMKL No 71:1999, 5th edition

3.1.2.4 Salmonella in food - Meat from pig - food sample

Monitoring system

Sampling strategy

At meat processing plant

Sampling according to 2073/2005 by FBO is verified under the regular offical control of the FBO.

At retail

The local health authorities (LCAs) are responsible for sampling at retail. No sampling in 2016

Frequency of the sampling

At meat processing plant

The frequency of sampling of minced meat and meat preparation by the FBO is according to the production capacity per week. More than 3 tons / week; 5 samples per week. 1 - 3 tons per week; 5 samples every second week. 500 kg - 1 ton; 5 samples 4 times per year. Frequency can be reduced when results have been negative for 30 weeks. Processing plants producing less the 500 kg / week does not need to take samples if they can demonstrate good hygiene practices.

Type of specimen taken

At meat processing plant

Minced meat and meat preparation

Definition of positive finding

At meat processing plant

Sample is considered to be positive when Salmonella spp. is detected.

At retail

Sample is considered to be positive when Salmonella spp. is detected.

Preventive measures in place

Sanitary slaughtering is performed when slaughtering from herds that are categorised as level 3 herds. For further information about categorizing into levels see chapter Salmonella spp. in pigs (meat juice index). HACCP activities are verified under official control of FBO.

Control program/mechanisms

The control program/strategies in place

The mandatory national control programme for Salmonella in pigs is approved by MAST. The aim of the programme is to prevent Salmonella in pork and thereby mitigate the risk of human salmonellosis. The surveillance programme is constructed to monitor Salmonella infections at farm level by measuring antibodies against Salmonella in meat juice test and to survey salmonella contamination on pig carcasses at slaughtering.

Notification system in place

Salmonella is a notifiable disease, according to national legislation on animal diseases No. 25/1993 and according to the national Food Law No. 93/1995. MAST receives all results from the respective laboratories regarding Salmonella testing (including serotyping and antimicrobial resistance) on samples from pig production, slaughtering and from FBO own checks in slaughterhouses, cutting plants and processing plants.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

Salmonella on pig carcasses seems not to be a major source for human infections.

3.1.2.5 Salmonella in food - Meat from pig - food sample - carcase swabs

Monitoring system

Sampling strategy

At slaughterhouse and cutting plant

The information needed is whether carcasses are contaminated with Salmonella or not. Therefore are all slaughter batches from all holdings tested for Salmonella. The sampling is a part of permanent control programme and is objective, official and performed by the MAST. Samples are taken from carcasses where the number of samples depend on the size of the slaughter batch.

Frequency of the sampling

At slaughterhouse and cutting plant

Every herd is sampled at each slaughter. From every herd ten randomly chosen carcasses from the slaughter batch are swabbed if 40 or fewer pigs are slaughtered, twenty carcasses are swabbed if 41 - 120 pigs are slaughtered and thirty carcasses are swabbed if more than 120 pigs are slaughtered. By this number of samples it can be declared by 95% confidence level that at least one positive sample will be detected in the batch if the prevalence in the batch is 10-15% or higher.

Type of specimen taken

At slaughterhouse and cutting plant

Surface of carcasses

Methods of sampling (description of sampling techniques)

At slaughterhouse and cutting plant

At slaughtering, the pig carcasses are split into two parts along the vertebrate. After health inspection both carcass parts are swabbed in three designated areas. One cotton swab is used for both carcass parts. Two of the three areas on each part cover approx. 10 x 10 cm. The third area covers approx. 100 cm2. The three different areas on each side of the carcass cover the following sites: Area I: Anus region, inside of the thigh and the pelvis region - Area II: The cut surface of the breast and abdomen, underbelly - Area III: Inside the cranial part of thorax and the adjacent transected bones/ribs. Swabs from 10 carcasses are pooled together in one sample.

Definition of positive finding

At slaughterhouse and cutting plant

The swabs tests are RapidChek. A positive test according to the protocol for the test and before confirmation by culturing, is regarded as positive for Salmonella. Confirmation by culturing is not needed, but at least one swab from each slaughter batch is serotyped and tested for antimicrobial resistance (every other year, 2013/652/EC). If a positive RapidChek test is negative by bacteriological culturing, the result has no influence on actions already taken regarding next slaughter from the respective herd.

Diagnostic/analytical methods used

At slaughterhouse and cutting plant

RapidChek Select Salmonella. Bacteriological method: NMKL 187:2007 Bacteriological method: NMKL 71:1999 5th edition

Measures in case of the positive findings or single cases

Finding one positive sample from a level 1 herd initiates reactions. In case of a positive sample all carcasses from the respective herd must be swabbed at next slaughter and quarantined whilst waiting for the results from the swab test. All positive carcasses must be heat treated before entering the market. All carcasses from level 2 and 3 herds are swabbed (and swabs from 5 carcasses are pooled together in one sample) and quarantined at the slaughterhouse and cannot enter the market unless swabs are negative. Carcasses positive for Salmonella must be heat treated. When salmonella is detected in a product, the product is withdrawn from the market.

Results of the investigation

2016: Swabs samples (RapidChek) taken at the slaughterhouses were 0,8% positive for Salmonella or 24 of 2863. Only 18 were positive in culturing; 4xSalmonella Brandenburg, 1xSalmonella Typhimurium, 6xmonophasic Salmonella Typhimurium, 1xSalmonella Infantis, 1xSalmonella Kedougou, 1xSalmonella London, 2xSalmonella Worthington and 2xSalmonella spp. were found. Results from the FBOs own checks according to 2073/2005 are not available.

National evaluation of the recent situation, the trends and sources of infection

The prevalence for Salmonella positive swab samples is relatively stable, 0,3-1,4% for the last 10 years, with the exception of 2009 at the prevalence at 11,2%.

3.1.2.6 Salmonella in food - Meat from turkey - food sample

Monitoring system

Sampling strategy

At meat processing plant

No official sampling. Sampling according to 2073/2005 by FBO is verified under the regular offical control.

At retail

The local competent authorities (LCA) are responsible for sampling at retail.

Frequency of the sampling

At meat processing plant

The frequency of sampling of minced meat and meat preparation by the FBO is according to the production capacity per week. More than 3 tons / week; 5 samples per week. 1 - 3 tons per week; 5 samples every second week. 500 kg - 1 ton; 5 samples 4 times per year. Frequency can be reduced when results have been negative for 30 weeks. Processing plants producing less the 500 kg / week does not need to take samples if they can demonstrate good hygiene practices.

Type of specimen taken

At meat processing plant

Minced meat and meat preparation

Methods of sampling (description of sampling techniques)

At meat processing plant

Each sample is 25 g of minced meat or meat preparations.

Definition of positive finding

At meat processing plant

Sample is considered to be positive when salmonella spp. is detected.

Diagnostic/analytical methods used

At meat processing plant

NMKL No 71:1999, 5th edition

Preventive measures in place

All flocks are sampled before slaughter and salmonella positive flocks are destroyed since it is prohibited to slaughter salmonella positive flocks. HACCP activities are verified during official control.

Measures in case of the positive findings or single cases

When salmonella is detected in a slaughter batch or in a product, the batch is withdrawn from the market.

Notification system in place

The FBO and the laboratory have to report a detection of salmonella spp. immediately to MAST.

Results of the investigation

Results from the FBOs own checks according to 2073/2005 are not available.

3.1.2.7 Salmonella in food - Meat from turkey - food sample - neck skin

Monitoring system

Sampling strategy

At slaughterhouse and cutting plant

The FBOs are sampling each slaughter batch. Sampling at cutting plant according to 2073/2005 by FBO is verified under the regular offical control.

Frequency of the sampling

At slaughterhouse and cutting plant

Each turkey slaughter batch is sampled

Type of specimen taken

At slaughterhouse and cutting plant

Neck skin

Methods of sampling (description of sampling techniques)

At slaughterhouse and cutting plant

From each slaughter batch, 50 neck skin samples, each sample consisting of 2-3g, are pooled into one sample. At the laboratory, a subsample of 25g is taken for further culturing.

Definition of positive finding

At slaughterhouse and cutting plant

A sample positive for Salmonella spp.

Diagnostic/analytical methods used

At slaughterhouse and cutting plant

Bacteriological method: NMKL No 71:1999, 5th edition

3.1.2.8 Salmonella in food - Eggs - food sample

Monitoring system

Sampling strategy

No official sampling

3.1.3 Salmonella in animals

3.1.3.1 Salmonella in animal - Cattle (bovine animals) - animal sample - milk

Monitoring system

Sampling strategy

Bulk milk from 73 farms. Random. Salmonella Dublin antibody examination in milk by ELISA

Methods of sampling (description of sampling techniques)

Animals at farm

Bulk milk taken at farms and collected in the dairies. Salmonella Dublin antibody examination in milk by ELISA

3.1.3.2 Salmonella in animal - Gallus gallus (fowl) - broilers - animal sample

Monitoring system Iceland - 2016

Sampling strategy

Broiler flocks

The monitoring is applied to the whole country. The sampling strategy is according to national requirements that were implemented in 2001 and 2002 and according to the national control programme from 2012 in accordance with reg. (EC) no. 2130/2003 and reg.(EC) no. 200/2012. Every broiler flock is sampled by the food business operator (FBO). On 10% of the farms, one flock per year is sampled by MAST in accordance with the national control programme. At slaughter samples are are also taken from each slaughter batch after evisceration but before chilling.

Frequency of the sampling

Broiler flocks: Before slaughter at farm

Within 3 weeks prior to slaughter

Type of specimen taken

Broiler flocks: Before slaughter at farm

Boot/sock swabs. According to the national control program it is also allowed to take one boot/sock sample and one dust sample but it is only practiced during the official sampling.

Methods of sampling (description of sampling techniques)

Broiler flocks: Before slaughter at farm

From each broiler flock two pairs of boot/socks swabs are collected and pooled into one sample. In case one boot/sock sample and one dust sample are taken, the two samples are pooled into one sample. The official sample consists of one boot/sock sample and one dust sample, and the samples are cultured separately.

Case definition

Broiler flocks: Before slaughter at farm

A flock is considered positive when Salmonella spp. is found in a sample. According to the national control programme, no confirming samples have to be taken. But as a part of the FBOs own controls, usually a confirmatory sample is taken. When salmonella is not found in two subsequent samples and not in a third sample taken by MAST, the flock is considered to be negative for Salmonella.

Diagnostic/analytical methods used

Broiler flocks: Before slaughter at farm

Bacteriological method: NMKL 187:2007

Vaccination policy

Broiler flocks

Vaccination of poultry flocks against Salmonella is not practiced in Iceland and not allowed according to the national control program.

Other preventive measures than vaccination in place

Broiler flocks

Preventive measures include specific requirements regarding biosecurity in accordance with national legislation.

Control program/mechanisms

The control program/strategies in place

Broiler flocks

A national control programme was published by MAST on 1.11.2008 and reviewed in february 2012 and december 2013 for the control of salmonella in poultry.

Measures in case of the positive findings or single cases

Broiler flocks: Before slaughter at farm

Birds from a flock that has been tested positive may not be moved from the farm except to the slaughterhouse. It is not allowed to distribute meat from positive flocks to the market, thus positive flocks are not slaughtered but culled on the farm.

Broiler flocks: At slaughter (flock based approach)

All raw meat from a positive flock at slaughter that has already been distributed is withdrawn from the market. All meat has to be heat treated before further distribution.

Notification system in place

Salmonella is a notifiable disease according to Icelandic legal act no. 25/1993. The laboratory sends an immediate notification to MAST when there is a positive salmonella finding.

Results of the investigation

Prevalence is only 0,4% in broiler flocks in 2016 or 3 of 713. All 3 are cases are likely due to recontamination of flocks in previously contaminated houses.

National evaluation of the recent situation, the trends and sources of infection

Since 1997 salmonella prevalence in broiler flocks at the farm level has been below 1%. After 2008 however, the prevalence of salmonella in broiler flocks rose and reached a peak in 2010. Reinfection of broiler flocks in previously contaminated houses is a major source of infection, as well as infection throught contaminated feed.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

In Iceland it is not allowed to distribute meat from Salmonella positive poultry flocks. Therefore, the main sources of contaminated poultry meat are flocks with a negative pre-slaughter sample but a positive sample at the time of slaughter. Cross contamination between negative and positive flocks at the slaughterhouse is also a source. Salmonella prevalence in slaughter flocks has been far below 1% since sampling started in 1998 and until 2010 (except for 2001 when prevalence was 1,8%). After 2010 prevalence in slaughter flocks rose above 1% but has been under 1% ever since or 0,1% in 2016. Neither S. Enterititis nor S. Typhimurium have been found in broilers during this time and there have been no reports or indications of human infections from infected broiler products in the past years.

3.1.3.3 Salmonella in animal - Pigs - animal sample - meat juice

Monitoring system

Sampling strategy

Breeding herds

There are neither breeding herds nor multiplying herds as such in Iceland which produce purebred pigs (f.exs. Landrace , Yorkshire) or hybrid pigs (e.g. LY, HD) for sale. Breeding or improvement of the stock is done by import of frozen semen from Norway. Therefore all pig herds produce their own purebreds (sows and boars) and hybrid pigs (sows and boars) mostly by insemination. All pig holdings/producers produce pigs for slaughtering, but some holdings can have separate herds with purebred/hybrid production, sow herds with piglet production or herds with fattening pigs.

Frequency of the sampling

Fattening herds at slaughterhouse (herd based approach)

Depends on herd size and classification into levels (see fattening herds)

Type of specimen taken

Fattening herds at slaughterhouse (herd based approach)

Meat juice

Methods of sampling (description of sampling techniques)

Fattening herds at slaughterhouse (herd based approach)

Meat samples are collected from carcasses after cooling. The meat samples are collected in special meat juice containers. Number of samples and frequency of sampling depend on herd size, ex. 75 samples/year from herds that slaughter between 2000 and 5000 pigs/year. All herds are sorted into three levels, depending on the results. Level 1 for those who are salmonella free or very low prevalence and level 2 and 3 as the prevalence is higher. Prevalence is calculated approximately every two weeks.

Case definition

Fattening herds at slaughterhouse (herd based approach)

An individual sample is considered seropositive if OD% is more than 15 (ELISA kit).

Diagnostic/analytical methods used

Fattening herds at slaughterhouse (herd based approach)

Serological method: PIGTYPE Salmonella Ab (qiagen)

Vaccination policy

Breeding herds

No salmonella vaccination occur

Fattening herds

No salmonella vaccination occur

Other preventive measures than vaccination in place

Breeding herds

In national regulation regarding pigs there are provisions on bio security demanding special facilities for entering the herd, isolation of and delivering pigs.

Multiplying herds

See breeding pigs.

Fattening herds

See breeding pigs. Separate transport of pigs to slaughterhouse and separate slaughtering of pigs in level three herds. In national regulation regarding pigs there are provisions on bio security demanding special facilities for entering the herd, isolation of and delivering pigs.

Control program/mechanisms

The control program/strategies in place

Fattening herds

The surveillance programme is constructed to monitor Salmonella infections at farm level by measuring antibodies against Salmonella in meat juice test, to survey salmonella contamination on pig carcasses at slaughtering and to monitor Salmonella serotypes in swab samples. The aim of the programme is to prevent salmonella in pork and thereby mitigate the risk of human salmonellosis. In order to control Salmonella infections at farm, the national regulation on pig health and housing no. 1276/2014 stipulates preventive measures to be implemented.

Measures in case of the positive findings or single cases

Measures are taken (described in Salmonella spp in pig meat and products therof) when herds are moved between levels or categories i.e. when a herd is downgraded from level 1 to level 2 or 3. Sanitary slaughtering is performed when herds are categorised as level 3 herds.

Notification system in place

Salmonella is a notifiable disease, according to national legislation on animal diseases No. 25/1993. MAST receives all results from the respective laboratories regarding Salmonella testing (including serotyping and antimicrobial resistance) on samples from pig production and slaughtering.

Results of the investigation

Salmonella is endemic in the pig production.

National evaluation of the recent situation, the trends and sources of infection

Prevalence has been increasing. Despite of the increase in Salmonella in the pig production salmonellosis in humans in the same period did not alter equally.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

Salmonella in pig production seems not to be a major source for human infections.

3.1.3.4 Salmonella in animal - Gallus gallus (fowl) - breeding flocks, unspecified - animal sample

Monitoring system

Sampling strategy

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

The monitoring of salmonella in breeding flocks of Gallus gallus is applied to the whole country. The sampling strategy is according to national requirements that were implemented in 2001 and 2002 and according to the national control programme from 2012 in accordance with reg. (EC) no. 2130/2003 and reg. (EU) No 200/2010. Every breeding flock consisting of 250 animals or more is sampled. During primary production samples are both taken by the food business operator (FBO) and by MAST in accordance with the national control programme. The adult breeding flocks are sampled at farm level. When a breeder flock is slaughtered, samples are taken from each slaughter batch at the time of slaughter, after evisceration but before chilling. In Iceland there are only parent flocks.

Frequency of the sampling

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

Every flock is sampled.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

At the age of 4 weeks and again 2 weeks before moving to the laying phase.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period

Every 2 weeks. If eggs from a breeding flocks are not incubated monthly or more often, then samples have to be taken each time before eggs are sent for incubation, and the result must be available before eggs are shipped to the hatchery.

Type of specimen taken

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

Socks/ boot swabs

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period

Socks / boots wabs, or boot swabs/dust sample, or fecal sample

Methods of sampling (description of sampling techniques)

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

From each breeding flock (paternal and maternal line separately) one sample is taken during hatching. The pooled sample consists of 10 g of broken eggshells taken from a total of 25 separate hatcher baskets, which in total is at least 250g of broken eggshells.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

From each breeding flock two pairs of boot swabs are collected and pooled into one sample, both at the age of 4 weeks and two weeks before moving to the laying phase.

Breeding flocks: Production period

Industry samples: On their choice, there are either taken 5 pairs of bootswabs (2 pooled samples) or one pair of bootswab and one dustsample, the dust sample is cultured separately. In case it is not possible to take bootswab samples, 2 pooled samples of in total 300 individual fecal samples are taken. The official samples are taken in the same way as the industry samples: two pooled samples with either 5 pairs of bootswabs or one pair of bootswabs and one dust sample, or 300 fecal samples. The samples are taken at the beginning and the end of the laying phase and once in beetween.

Case definition

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

A sample positive for salmonella spp. In cases where MAST has suspicion of a false positive result, MAST can confirm a first positive finding, and if it is not possible to confirm, the flock will be calculated as negative.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

A sample positive for salmonella spp. In cases where MAST has suspicion of a false positive result, MAST can confirm a first positive finding, and if it is not possible to confirm, the flock will be calculated as negative.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period

A sample positive for salmonella spp., always confirmed by MAST. If it is not possible to confirm a first positive finding in two consecutive samples taken by MAST, the flock will be calculated as negative.

Diagnostic/analytical methods used

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

Bacteriological method: NMKL 187:2007

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

Bacteriological method: NMKL 187:2007

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period

Bacteriological method: NMKL 187:2007

Vaccination policy

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

Vaccination of poultry flocks against salmonella is not allowed according to the national control program.

Other preventive measures than vaccination in place

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

Preventive measures include specific requirements regarding biosecurity in accordance with national legislation.

Control program/mechanisms

The control program/strategies in place

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

A national control programme was published by MAST on 1.11.2008 and reviewed in february 2012 and december 2013 for the control of salmonella in poultry.

Measures in case of the positive findings or single cases

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

Birds from a flock that has been tested positive may not be moved from the farm except to the slaughterhouse. It is not allowed to distribute meat from positive flocks to the market, thus positive flocks are not slaughtered but culled on the farm.

Notification system in place

Salmonella is a notifiable disease according to Icelandic legal act nr. 25/1993. The laboratory sends an immediate notification to MAST when there is a positive Salmonella finding.

Results of the investigation

No Salmonella spp. was found in breeding flocks of Gallus gallus.

National evaluation of the recent situation, the trends and sources of infection

No Salmonella spp. was found in breeding flocks of Gallus gallus.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

No salmonella spp. was found in breeding flocks of Gallus gallus and they are not considered as a source of infection in boilers, other animals, foodstuffs or humans.

3.1.3.5 Salmonella in Turkeys - breeding flocks and meat production flocks

Monitoring system

Sampling strategy

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

The monitoring of Salmonella in turkey breeding flocks is applied to the whole country. The sampling strategy is according to national requirements that were implemented in 2001 and 2002 and according to the national control programme from 2012 in accordance with reg. (EC) no. 2130/2003 and reg. (EU) No 1190/2012. Every breeding flock consisting of 250 animals or more is sampled. During primary production samples are both taken by the food business operator (FBO) and by MAST in accordance with the national control programme. The adult breeding flocks are sampled at farm level. When a breeder flock is slaughtered samples are taken from each slaughter batch at the time of slaughter, after evisceration but before chilling. In Iceland there are only parent flocks.

Meat production flocks

The monitoring is applied to the whole country. The sampling strategy is according to national requirements that were implemented in 2001 and 2002 and according to the national control programme from 2012 in accordance with reg. (EC) no. 2130/2003 and reg.(EC) no. 1190/2012. Every flock is sampled by the food business operator (FBO). Official samples are taken from 10% of flocks with more than 500 animals. At slaughter samples are are also taken from each slaughter batch after evisceration but before chilling. The sampling strategy is according to national requirements that were implemented in 2001 and 2002 and according to the national control programme. Official samples are taken by the competent authority from one flock on one farm once a year (from 10% of all farms with over 500 fattening turkeys).

Frequency of the sampling

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

Every flock is sampled.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

At the age of 4 weeks and two weeks before moving to the laying house.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period

Every 3 weeks.

Meat production flocks: Before slaughter at farm

3 weeks prior to slaughter.

Meat production flocks: At slaughter (flock based approach)

Every batch is sampled.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

Hatched eggs

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

Socks/ boot swabs

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period

Socks / boots wabs, or boot swabs/dust sample

Meat production flocks: Before slaughter at farm

Boot/sock swabs. According to the national control program it is also allowed to take one boot/sock sample and one dust sample but it is only practiced during the official sampling.

Meat production flocks: At slaughter (flock based approach)

Neck skin

Methods of sampling (description of sampling techniques)

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

From each breeding flock (paternal line and maternal line separately) one sample is taken during hatching. The pooled sample consists of 10 g of broken eggshells taken from a total of 25 separate hatcher baskets, which in total is at least 250g of broken eggshells.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

From each breeding flock two pairs of boot/sock swabs are collected and pooled into one sample, both at the age of 4 weeks and two weeks before moving to the laying phase.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period

Industry samples: On their choice, there are either taken 5 pairs of boot/sock swabs (2 pooled samples) or one pair of boot/sock swab and one dust sample, the dust sample is cultured separately. The official samples are taken in the same way as the industry samples: two pooled samples with either 5 pairs of boot/sock swabs or one pair of boot/sock swabs and one dust sample. The samples are taken once a year from 10% of all flocks.

Meat production flocks: Before slaughter at farm

From each flock two pairs of boot/sock swabs are collected and pooled into one sample. In case one boot/sock sample and one dust sample are taken, the two samples are pooled into one sample. The official sample consists of one boot/sock sample and one dust sample, and the samples are cultured separately.

Meat production flocks: At slaughter (flock based approach)

From each slaughter flock one pooled sample consisting of 50 neck skin samples is taken, with each neck skin sample weighting between 3 - 5 g.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

A sample positive for Salmonella spp., always confirmed by MAST. If it is not possible to confirm a first positive finding in two consecutive samples taken by MAST, the flock will be calculated as negative.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

A sample positive for Salmonella spp. In cases where MAST has suspicion of a false positive result, MAST can confirm a first positive finding, and if it is not possible to confirm, the flock will be calculated as negative.

Meat production flocks: Rearing period

A flock is considered positive when Salmonella spp. is found in a sample. According to the national control progam, no confirming samples have to be taken. But as a part of the FBOs own controls, usually a confirmatory sample is taken. When Salmonella is not found in two subsequent samples and not in a third sample taken by MAST, the flock is considered to be negative for salmonella.

Meat production flocks: Before slaughter at farm

A sample positive for Salmonella spp.

Meat production flocks: At slaughter (flock based approach)

Bacteriological method: NMKL 187:2007

Diagnostic/analytical methods used

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

Bacteriological method: NMKL 187:2007

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

Bacteriological method: NMKL 187:2007

Meat production flocks: Rearing period

Bacteriological method: NMKL 187:2007

Meat production flocks: Before slaughter at farm

Bacteriological method: NMKL No 71:1999

Meat production flocks: At slaughter (flock based approach)

Vaccination of poultry flocks against salmonella is not allowed according to the national control program.

Vaccination policy

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

Iceland - 2016

Vaccination of poultry flocks against salmonella is not allowed according to the national control program.

Meat production flocks

Preventive measures include specific requirements regarding biosecurity in accordance with national legislation.

Other preventive measures than vaccination in place

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

Preventive measures include specific requirements regarding biosecurity in accordance with national legislation.

Meat production flocks

A national control programme was published by MAST on 1.11.2008 and reviewed in february 2012 and december 2013 for the control of Salmonella in poultry.

Control program/mechanisms

The control program/strategies in place

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

A national control programme was published by MAST on 1.11.2008 and reviewed in february 2012 and december 2013 for the control of Salmonella in poultry.

Measures in case of the positive findings or single cases

Meat Production flocks

Salmonella is a notifiable disease according to Icelandic legal act no. 25/1993. The laboratory sends an immediate notification to MASTwhen there is a positive Salmonella finding.

Notification system in place

In 2016, Salmonella spp. was not found in turkey parent flocks nor flocks of fattening turkeys.

Results of the investigation

Salmonella had never been detected in turkey breeder flocks. However, in the recent years, S. Worthington in flocks of fattening turkeys has been connected with contaminated feed.

National evaluation of the recent situation, the trends and sources of infection

No Salmonella spp. has been found in turkey breeding flocks and they are not considered as a source of infection in flocks of fattening turkeys, other animals, foodstuffs or humans. In Iceland it is not allowed to slaughter Salmonella positive poultry flocks. Therefore, the main sources of contaminated poultry meat are flocks with a negative pre-slaughter sample but a positive sample at the time of slaughter, which has not been the case for turkey flocks. In recent years there have been no indications of any human infections from infected icelandic turkey meat.

3.2 CAMPYLOBACTERIOSIS

3.2.1 General evaluation of the national situation

3.2.1.1 Thermophilic Campylobacter spp., unspecified - general evaluation

History of the disease and/or infection in the country

Following the marketing of chilled broiler meat (only been frozen) the yearly consumption increased from 6 kg per inhabitant in 1996 to 11 kg per inhabitant in 1999. Along with the increased consumption incidences of campylobacteriosis in humans began to increase. Icelandic veterinary and human health authorities came to the conclusion that the major increase of human campylobacteriosis in Iceland in 1999 was linked to the consumption and handling of chilled broiler meat. In cooperation with the poultry industry in Iceland the authorities implemented a national surveillance programme for Campylobacter in poultry. It began as a voluntary programme in May 2000, but by late 2001 and early 2002 it was implemented into national legislation. The objective of the programme is to prevent the distribution of chilled poultry carcasses or poultry meat that is known to be Campylobacter positive. Therefore all meat from positive poultry flocks are either frozen or heat treated before distribution. However campylobacter positive carcasses or chilled meat can have been distributed when a flock turned positive after the pre-slaughter sample had been taken. Only carcasses from poultry flocks that are negative prior to slaughter can be distributed as chilled meat. With regard to the surveillance programme the producers have to produce Campylobacter negative broilers if they want to sell and distribute chilled broiler meat. This is why the Icelandic surveillance programme is the driving force for all efforts to prevent broiler flocks from getting contaminated with Campylobacter; see chapter on Campylobacter in Gallus gallus.

National evaluation of the recent situation, the trends and sources of infection

In the recent years the prevalence in broiler slaughter batches has been been very low. It is assumed that the implementation of the surveillance programme has decreased the human incidence of campylobacteriosis. These results have been achieved despite the fact that annual consumption of broiler meat has increased from 6 kg pr. inhabitant in 1996 to 24 kg pr. inhabitant in 2009 and that probably over 90% of all broiler meat on the market is sold chilled. No unfrozen or unheattreated poultry meat is imported, but the import of frozen broiler meat is increasing. Epidemiological studies in the recent years showed that other vehicles such as drinking water rather than broiler meat seem to contribute to human campylobacteriosis.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

Consumption and handling of poultry and poultry products is probably not the only important source of human campylobacteriosis in Iceland any longer. Other sources might be a more important source of human campylobacteriosis in Iceland than previously thought.

Recent actions taken to control the zoonoses

The implementation of the Icelandic surveillance programme for Campylobacter in poultry was a direct response from the food and veterinary authority and the poultry industry to the major increase in human campylobacteriosis in 1999. The programme is almost the same as it was when implemented in 2000 and only minor changes have been made.

3.2.2 Campylobacter in foodstuffs

3.2.2.1 Thermophilic Campylobacter spp., unspecified in food - Meat from broilers (Gallus gallus) - food sample

Monitoring system

Sampling strategy

At slaughterhouse and cutting plant

See text form on thermophilic Campylobacter in Gallus gallus

At meat processing plant

No official sampling

At retail

There is no regular annual sampling performed. But sporadic surveys are performed. The local competent authorities (LCA) are responsible for sampling at retail. No sampling in 2016

Preventive measures in place

Meat from positive broiler flocks is frozen for at least 14 days or heat treated before it is placed on the marked. Meat from other countries has to be frozen for at least 30 days prior to distribution.

Control program/mechanisms

The control program/strategies in place

See thermophilic Campylobacter in Gallus gallus

3.2.3 Campylobacter in animals

3.2.3.1 Thermophilic Campylobacter spp., unspecified in animal - Gallus gallus (fowl) - animal sample

Monitoring system

Sampling strategy

According to the Icelandic Campylobacter National Surveillance Programme every poultry flock is sampled at the farm 2 to 5 days prior to slaughter, if it is intended to distribute meat from the flock unfrozen and unheattreated (fresh). Samples are taken by the food business operator (FBO). Samples are also taken from each poultry flock at slaughter from april 1 to october 31 every year, if the meat from the flock is distributed unfrozen and unheattreated (fresh).

Frequency of the sampling

Before slaughter at farm

Every flock from which the meat is intended to be distributed unfrozen and unheattreated is sampled by the FBO, 2-5 days before slaughter.

At slaughter

Annually, from March 1 to October 31, every batch is sampled by the FBO, if the meat from the flock is intended to be distributed unheattreated and unfrozen. If at slaughter no pre-slaughter sample is available, but it is intended to distribute meat from the batch unheattreated and unfrozen, samples are taken at slaughter and it is not allowed to distribute meat from the batch until results from slaughter samples are available and negative.

Type of specimen taken

Before slaughter at farm

Faeces

At slaughter

Caecal

Methods of sampling (description of sampling techniques)

Before slaughter at farm

10 individual fresh faecal droppings are taken by the FBO 2 to 5 days before slaughter. They are sent as one pooled sample to the laboratory.

At slaughter

Caecum from 10 broiler carcasses are taken at the slaughter line by the FBO, pooled into one sample.

Case definition

Before slaughter at farm

A flock is considered positive when Campylobacter spp. is detected.

At slaughter

A slaughter batch is considered positive when Campylobacter spp. is detected.

Diagnostic/analytical methods used

Before slaughter at farm

Bacteriological method: NMKL 119-3, 2007

At slaughter

Bacteriological method: NMKL 119-3, 2007, EN/ISO 10272 part 1 and 2

Vaccination policy

There is no vaccination against Campylobacter in Iceland.

Other preventive measures than vaccination in place

Generally the freezing policy (meat from contaminated flocks has to be frozen) is the driving force for all preventive measures at farm level. As only flocks that test negative for Campylobacter prior to slaughter can be processed as fresh chilled products the farmers aim to produce negative flocks, as the fresh chilled products have the highest market value. The most important interventions done at the farm level is maintaining a very high biosecurity all year round. Preferrably flocks are not thinned. Fly nets are used on air inlets for houses on high density farms during the summer period.

Control program/mechanisms

The control program/strategies in place

The Icelandic Campylobacter surveillance programme is build on the freezing policy which means that all poultry flocks positive in the preslaugher sample are submitted to freezing or are heat treated before distribution. Only carcasses from poultry flocks that are negative prior to slaughter can be processed as chilled meat from the slaughterhouses. The objective of the programme is to prevent distribution of poultry carcasses or poultry meat that is known to be Campylobacter positive as chilled products.

Measures in case of the positive findings or single cases

Carcasses from flocks that test positive for thermophilic Campylobacter sp. based upon the pre-slaughter sampling are either subjected to heattreatment or freezing.

Notification system in place

All results from samples taken from broiler flocks are as a part of the surveillance programme reported directly to MAST from the laboratories, both negative and positive results.

Results of the investigation

The prevalence of Campylobacter sp. in broiler flocks (pre slaughter samples) has been very low in the recent years.

National evaluation of the recent situation, the trends and sources of infection

The trend in the last decade has been that the Campylobacter prevalence in broiler flocks is decreasing.

3.3 LISTERIOSIS

3.3.1 General evaluation of the national situation

3.3.1.1 Listeria - general evaluation

History of the disease and/or infection in the country

Sporadic cases in humans, incidence from 0-1/100.000 inhabitants annually.

Recent actions taken to control the zoonoses

The requirements of the Regulation (EC) No 2073/2005 apply, i.e. verification of FBO's HACCP procedures, monitoring of the production process, shelf-life studies when deemed appropriate, withdrawal from the market when unsatisfactory results and taking measures to prevent the recurrence of the contamination, such as reviewing the production routines and shelf life of the product. Dietary advice is given to pregnant women.

3.3.2 Listeria in animals

3.3.2.1 Listeria in animal - All animals - animal sample

Monitoring system

Sampling strategy

There is no active surveillance in animals regarding L. monocytogenes.

Notification system in place

Listeriosis in animals is a notifiable disease.

National evaluation of the recent situation, the trends and sources of infection

Based on the notification system, little changes are seen in the prevalence of listeriosis in animals.

3.4 TRICHINELLOSIS

3.4.1 General evaluation of the national situation

3.4.1.1 Trichinella - general evaluation

History of the disease and/or infection in the country

Samples are taken from all slaughtered pigs and horses according to 2015/1375/EC. When particularly looked for Trichinella has neither been found in livestock nor in domestic wild animals (foxes, minks) in Iceland. Three cases of Trichinella sp. in stray polar bears are known from the years 1963, 2008 and 2010. In 1963 Trichinella sp. was found in a stray polar bear which came ashore in the north-western part of the country, in 2008 Trichinella native was found in another polar bear that came ashore in the northern part of the country and in 2010 Trichinella sp. was found in the third animal which was detected in the north-eastern part of Iceland. When exporting horse meat some purchasers demand Trichinella testing at slaughter and prior to export. As a result of these demands approx. 1.600 samples from horses had been investigated up until the year 2008, which were negative with regard to Trichinella sp. Trichinella has never been diagnosed in humans in Iceland.

National evaluation of the recent situation, the trends and sources of infection

The risk of Trichinella spp. beeing introduced into Icelandic wildlife does exist because of occasional stray polar bears coming ashore. The probability of the risk is however very low because polar bears are euthanized and their carcasses removed as soon as they are detected.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

The probability of contracting trichinellosis from food producing animals of Icelandic origin is close to zero.

Recent actions taken to control the zoonoses

3.4.2 Trichinella in animals

3.4.2.1 Trichinella in animal - Solipeds, domestic - horses - animal sample - organ/tissue

Monitoring system

Sampling strategy

Samples were taken from all slaughtered horses intended to be put on the marked.

Frequency of the sampling

Samples were taken from all horses.

Type of specimen taken

Tongue or masseter muscle.

Methods of sampling (description of sampling techniques)

Methods used were in accordance with 2015/1375/EC. For analyses, 5 g per animal is included in a pooled sample of maximum 100 g.

Case definition

An animal with a positive test result in the official examination.

Diagnostic/analytical methods used

According to 2015/1375/EC, Car 1, Chap 1 Magnetic stirrer method for pooled sample digestion for the detection of Tricinella in meat

Control program/mechanisms

The control program/strategies in place

Sampling was according to 2015/1375/EC and is monitored by official vets.

Measures in case of the positive findings or single cases

In case of a positive sample the horse carcasses will be retested individually. Pursuant to the Law on Animal Health No 25/1993, the Minister can issue national regulation on any necessary measures to be taken in order to find the source of the infection, restrict movement of animals, hinder the spread of and eradicate the disease.

Notification system in place

According to the national Law on Animal Health, no. 25/1993, trichinellosis is a list B disease that must be notified.

Results of the investigation including the origin of the positive animals

No cases of Trichinellosis were reported among slaughtered horses.

National evaluation of the recent situation, the trends and sources of infection

No cases have been reported in Iceland, neither in humans nor in animals.

3.4.2.2 Trichinella in animal - Pigs - animal sample - organ/tissue

Number of officially recognised Trichinella-free holdings

All holdings are considered trichinella - free.

Categories of holdings officially recognised Trichinella-free

No categorization of holdings.

Monitoring system

Sampling strategy

General

Samples were taken from all slaughtered pigs intended to be placed on the market.

Frequency of the sampling

General

Samples were taken from all pigs slaughtered.

Type of specimen taken

General

Muscle clip from the pillar of diaphragma.

Methods of sampling (description of sampling techniques)

General

Samples are taken according to 2015/1375/EC by a trained employee of the slaughterhouse under the supervision of the official vet. 1g of muscle clip from the pillar of diaphragma (max 100 pigs per anlyse).

Case definition

General

An animal where Trichinella spp. larvae has been detected.

Diagnostic/analytical methods used

General

Artificial digestion method of pooled samples.

Preventive measures in place

Controlled housing condition in pig farms. Regular official control of farms and slaughterhouses.

Control program/mechanisms

The control program/strategies in place

Sampling of all slaughtered pigs.

Measures in case of the positive findings or single cases

In case of a positive sample the pig carcasses will be retested according to 2015/1375/EC. Pursuant to the Law on Animal Health No 25/1993, the Minister can issue national regulation on any necessary measures to be taken in order to find the source of the infection, restrict movement of animals, hinder the spread of and eradicate the disease.

Notification system in place

No positive result on trichinella from the samples taken.

Notification system in place

According to the national Law on Animal Health, no. 25/1993, trichinellosis is a list B disease that must be notified.

Results of the investigation including description of the positive cases and the verification of the Trichinella species

Breeding sows and boars

No cases have been reported in Iceland, neither in humans or live stock nor in domestic wild animals (foxes, minks) in Iceland.

3.5 ECHINOCOCCOSIS

3.5.1 General evaluation of the national situation

3.5.1.1 Echinococcus - general evaluation

History of the disease and/or infection in the country

Echinococcus multilocularis has never been diagnosed. Echinococcus granulosus used to be quite frequent but has now apparently been eradicated, the last incidence detected in animals was in 1979 and in humans in 1988.

National evaluation of the recent situation, the trends and sources of infection

Emphasis is put on keeping the country free from Echinococcus. Annual treatment of dogs with anthelmintics is required and all carcasses at slaughterhouses are checked visually. The risk of introduction of Echinococcus multilocularis is considered moderate or low as the main definitive host, the red fox (Vulpes vulpes) is not present in the country and not wolves or coyotes neither. The same applies for many common intermediate hosts such as vole, lemming and muskrat. The only fox in Iceland is the arctic fox (Vulpes lagopus), which is seldom seen in the vicinity of residential areas.

4 ANTIMICROBIAL RESISTANCE INFORMATION ON SPECIFIC ZOONOSES AND ZOONOTIC AGENTS

4.1 SALMONELLOSIS

4.1.1 Salmonella in foodstuffs

4.1.1.1 Antimicrobial resistance in Salmonella Meat from pig

Description of sampling designs

The isolates detected through the national control programme (swab samples from carcasses) for Salmonella in pigs were NOT tested for AMR in 2016 (2013/652/EU). See chapter on Salmonella in pigs.

Stratification procedures per animal populations and food categories

Since all samples were included in the AMR monitoring, no stratification procedures had to be implemented.

Randomisation procedures per animal populations and food categories

Since all isolates were included in the AMR testing, no randomisation procedures had to be implemented

4.1.1.2 Antimicrobial resistance in Salmonella Meat from poultry, unspecified

Stratification procedures per animal populations and food categories

Since all samples were included in the AMR monitoring, no stratification procedures had to be implemented.

Randomisation procedures per animal populations and food categories

Since all isolates were included in the AMR testing, no randomisation procedures had to be implemented

Sampling strategy used in monitoring

Frequency of the sampling

Salmonella found in neck skin samples taken according to the national control programme is included in the resistance monitoring. One positive sample from each poultry flock is serotyped and tested for antimicrobial sensitivity.

Type of specimen taken

Neck skin samples. Isolates detected within the national control program

Methods of sampling (description of sampling techniques)

Isolates detected within the national control program

Procedures for the selection of isolates for antimicrobial testing

MAST selects the isolates to ensure that only one isolate from each serovar from each batch is tested for AMR.

Methods used for collecting data

The laboratory performing the AMR testing sends all results to MAST.

Laboratory methodology used for identification of the microbial isolates

NMKL No 71:1999. For serotyping, the presumtive Salmonella colonies are confirmed with MALDI TOF, then serotyped.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Ampicillin, Cefotaxime, Ceftazidime, Chloramphenicol, Ciprofloxacin, Colistin, Gentamicin, Meropenem, Nalidixic acid, Sulfamethoxazole, Tetracycline, Trimethoprim, Azithromycin, Tigecycline. According to decision 2013/652/EU.

Cut-off values used in testing

The values used are ECOFF values given in the 2013/652/EU decision.

4.1.2 Salmonella in animals

4.1.2.1 Antimicrobial resistance in Salmonella Pigs

Sampling strategy used in monitoring

Frequency of the sampling

Salmonella isolates found in pig herds within the national control programme are included in the resistance monitoring (every other year, according to Desicion 2013/652/EU). No samples were taken from pig herds on farms in 2016.

4.1.2.2 Antimicrobial resistance in Salmonella Poultry, unspecified

Sampling strategy used in monitoring

Frequency of the sampling

Salmonella found in poultry in samples taken according to the national control programme is included in the resistance monitoring.

Type of specimen taken

Sock samples, fecal samples, dust samples from all types of poultry production, according to the national control programme.

Methods of sampling (description of sampling techniques)

Isolates detected within the national control programme.

Procedures for the selection of isolates for antimicrobial testing

Mast selects the isolates, in order to ensure, that only one isolate from each flock is tested.

Methods used for collecting data

The laboratory performing the AMR testing sends all results to MAST.

Laboratory methodology used for identification of the microbial isolates

NMKL No 187. For serotyping, the presumtive Salmonella colonies are confirmed with MALDI TOF, then serotyped.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Ampicillin, Cefotaxime, Ceftazidime, Chloramphenicol, Ciprofloxacin, Colistin, Gentamicin, Meropenem, Nalidixic acid, Sulfamethoxazole, Tetracycline, Trimethoprim, Azithromycin, Tigecycline. According to decision 2013/652/EC

Cut-off values used in testing

The values used are ECOFF values given in the 2013/652/EU decision.

4.2 ESCHERICHIA COLI, NON-PATHOGENIC

4.2.1 Escherichia coli, non-pathogenic in animals

4.2.1.1 Antimicrobial resistance in E.coli, non-pathogenic, unspecified Pigs

Description of sampling designs

Ceacal samples at slaughter. For ESBL/AmpC producing E. coli, 150 samples are taken, evenly distributed over the year. For commensal indikator E. coli, sample from each epidemiological unit.

Stratification procedures per animal populations and food categories

Since all samples were included in the AMR monitoring, no stratification procedures had to be implemented.

Randomisation procedures per animal populations and food categories

Since all isolates were included in the AMR testing, no randomisation procedures had to be implemented

Sampling strategy used in monitoring

Type of specimen taken

Ceacal sample at slaughter

Procedures for the selection of isolates for antimicrobial testing

All positive samples tested for AMR

Methods used for collecting data

The laboratory performing the AMR testing sends all results to MAST.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Panel of antimicrobial substances (panel 1) according to decision 2013/652/EC; Ampicillin, Cefotaxime, Ceftazidime, Chloramphenicol, Ciprofloxacin, Colistin, Gentamicin, Meropenem, Nalidixic acid, Sulfamethoxazole, Tetracycline, Trimethoprim, Azithromycin, Tigecycline. For positive ESBL/AmpC producing E. coli panel 2 is included; Cefoxitin, Cefepime, Cefotaxime+clavulanic acid, Ceftazidime+clavulanic acid, Meropenem, Temocillin, Imipenem, Ertapenum, Cefotaxime, Ceftazidime.

Cut-off values used in testing

The values used are ECOFF values given in the 2013/652/EU decision.

5 FOODBORNE OUTBREAKS

Foodborne outbreaks are incidences of two or more human cases of the same disease or infection where the cases are linked or are probably linked to the same food source. Situation, in which the observed human cases exceed the expected number of cases and where a same food source is suspected, is also indicative of a foodborne outbreak.

5.1 Outbreaks

5.1.1 Foodborne outbreaks

System in place for identification, epidemological investigations and reporting of foodborne outbreaks

LCAs and MAST shall inform the chief epidemiologist immediately if they become aware of a risk of infections. Laboratories detecting zonoosis in food should report to MAST. Phycisians and laboratories analysing human samples are required, according to article 3 in the Act on Health Security and Communicable Diseases no. 1997/19 to report notifiable diseases and diseases subject to registration to the the Chief epidemiologist. The Chief epidemiologist monitor the data and shall report suspected foodborne outbreaks to MAST and the relevant LCAs. MAST, chief epidemiologist and LCA if relevant work together in investigations of food borne outbreak. The chief epidemiologist is responsible for epidemiological investigation of humans and MAST is responsible investigation of animals and coordination of the LCAs. MAST and the LCSs are responsible for investigation of food and the FBOs .

Description of the types of outbreaks covered by the reporting:

All suspected foodborne outbreaks are notifiable. The definition of a foodborne outbreak is two or more human cases with the same infection where the cases are linked or are probably linked to the same foodsource, or when observed number of human cases exceeds the expected number of cases during the same time period and place, and food is a likely vehicle.

National evaluation of the reported outbreaks in the country:

Trends in numbers of outbreaks and numbers of human cases involved

Food borne outbreaks of domestic origin are very few (0-5) each year. The situation has been stable the past 10 years.

Additional information

Standarized reporting and harmonization not fully implemented between different authorities. Therefore, reporting of FBO's is not possible. Standardization to be implemented in 2017. Four different FBO registered in 2016; One VTEC, one Campylobacter and two Salmonella cases.

ANIMAL POPULATION TABLES

Table Susceptible animal population

			Рор	oulation	
Animal species	Category of animals	holding	animal	slaughter animal (heads)	herd/flock
Cattle (bovine animals)	Cattle (bovine animals) - calves (under 1 year) - dairy calves	709	11,626		709
	Cattle (bovine animals) - calves (under 1 year) - for slaughter	714	10,423		714
	Cattle (bovine animals) - dairy cows - adult	648	26,347		648
	Cattle (bovine animals) - dairy cows - young cattle (1-2 years)	636	6,546		636
	Cattle (bovine animals) - meat production animals - suckler cows	132	2,176		132
	Cattle (bovine animals) - unspecified			22,658	
	Cattle (bovine animals) - young cattle (1-2 years)	796	22,906		796
Gallus gallus (fowl)	Gallus gallus (fowl) - broilers	34	683,556	5,371,118	84
	Gallus gallus (fowl) - laying hens - adult	301	196,206		301
	Gallus gallus (fowl) - laying hens - during rearing period	38	51,294		38
	Gallus gallus (fowl) - parent breeding flocks for broiler production line - adult	4	50,590	24,687	19
	Gallus gallus (fowl) - parent breeding flocks for broiler production line - during rearing period	6	52,672		14
	Gallus gallus (fowl) - parent breeding flocks for egg production line - adult	4	7,984		6
	Gallus gallus (fowl) - parent breeding flocks for egg production line - during rearing period	1	5,092		1
Goats	Goats	106	1,188	194	106
Pigs	Pigs - breeding animals - raised under controlled housing conditions - boars	16	59	108	16
	Pigs - breeding animals - raised under controlled housing conditions - sows	19	3,451	1,492	19
	Pigs - fattening pigs - raised under controlled housing conditions	20	25,791	74,522	20
	Pigs - fattening pigs - raised under controlled housing conditions - piglets	15	8,715		15
Sheep	Sheep - animals over 1 year	2,470	389,800	42,356	2,470
	Sheep - animals under 1 year (lambs)	2,279	86,093	555,617	2,279
Solipeds, domestic	Solipeds, domestic - horses		67,186	7,889	
Turkeys	Turkeys - meat production flocks	8	17,005	44,665	9
	Turkeys - parent breeding flocks - adult	1	503		3
	Turkeys - parent breeding flocks - during rearing period	2	1,447		2

DISEASE STATUS TABLES

Table Bovine brucellosis in countries and regions that do not receive Community co-financing for eradication programme

Region	Number of herds with status officially free	Number of infected herds	Total number of herds
ICELAND	846	0	846

Table Ovine or Caprine brucellosis in countries and regions that do not receive Community co-financing for eradication programme

Region	Number of herds with status officially free	Number of infected herds	Total number of herds
ICELAND	2,480	0	2,480

DISEASE STATUS TABLES

Table Bovine tuberculosis in countries and regions that do not receive Community co-financing for eradication programme

Region	Number of herds with status officially free	Number of infected herds	Total number of herds
ICELAND	846	0	846

PREVALENCE TABLES

Table CAMPYLOBACTER in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit		Total units positive	Zoonoses	N of units positive
Not Available	Gallus gallus (fowl) - broilers - before slaughter - Farm - Iceland - animal sample - faeces - Control and eradication programmes - Industry sampling - Census	herd/floc k	715	22	thermotolerant Campylobacter, unspecified	22
	Gallus gallus (fowl) - broilers - Slaughterhouse - Iceland - animal sample - caecum - Control and eradication programmes - Industry sampling - Selective sampling	slaughte r animal batch	449	11	thermotolerant Campylobacter, unspecified	11
	Turkeys - meat production flocks - before slaughter - Farm - Iceland - animal sample - faeces - Control and eradication programmes - Industry sampling - Census	herd/floc k	36	1	thermotolerant Campylobacter, unspecified	1
	Turkeys - meat production flocks - Slaughterhouse - Iceland - animal sample - caecum - Control and eradication programmes - Industry sampling - Selective sampling	slaughte r animal batch	25	0	Campylobacter	0

Table COXIELLA in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Total units tested	Total units positive	N of clinical affected herds	Zoonoses	N of units positive
Not Available	Cattle (bovine animals) - dairy cows - adult - Farm - Iceland - animal sample - milk - Monitoring - Official sampling - Objective sampling	herd/floc k	73	0	0	Coxiella	0

Table HISTAMINE in food

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	•		Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Fishery products, unspecified - ready-to-eat - chilled - Border inspection activities - Thailand - food sample - Surveillance - Official sampling - Selective sampling	batch (food/fee d)	5	Gram	27	0	<= 100	Histamine	0	0

Table LISTERIA in food

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit		•	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Roe - frozen - Border inspection activities - Peru - food sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	5	0	detection	Listeria monocytogenes	5	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	N of flocks under contro programme		Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Cattle (bovine animals) - dairy cows - adult - Farm - Not Available - Not Available - Monitoring - Official sampling - Objective sampling	herd/floc k		N_A	73	0	Salmonella	0
	Gallus gallus (fowl) - broilers - before slaughter - Farm - Iceland - environmental sample - boot swabs -	herd/floc	713	Y	713	3	Salmonella Agona	1
	Control and eradication programmes - Official and industry sampling - Census	k					Salmonella Infantis	1
							Salmonella Worthington	1
	Gallus gallus (fowl) - laying hens - adult - Farm - Iceland - environmental sample - boot swabs - Control and eradication programmes - Official and industry sampling - Census	herd/floc k	53	Y	49	0	Salmonella	0
	Gallus gallus (fowl) - laying hens - day-old chicks - Farm - Iceland - environmental sample - delivery box liner - Control and eradication programmes - Industry sampling - Census	herd/floc k		N_A	33	0	Salmonella	0
	Gallus gallus (fowl) - laying hens - during rearing period - Farm - Iceland - animal sample - faeces - Control and eradication programmes - Industry sampling - Census	herd/floc k		N_A	7	0	Salmonella	0
	Gallus gallus (fowl) - parent breeding flocks for broiler production line - adult - Farm - Iceland - environmental sample - boot swabs - Control and eradication programmes - Official and industry sampling - Census	herd/floc k	39	Y	39	0	Salmonella	0
	Gallus gallus (fowl) - parent breeding flocks for broiler production line - day-old chicks - Farm - Iceland - animal sample - eggshells - Control and eradication programmes - Industry sampling - Census	herd/floc k		N_A	7	0	Salmonella	0
	Gallus gallus (fowl) - parent breeding flocks for broiler production line - during rearing period - Farm - Iceland - environmental sample - boot swabs - Control and eradication programmes - Industry sampling - Census	herd/floc k		N_A	15	0	Salmonella	0
	Gallus gallus (fowl) - parent breeding flocks for egg production line - adult - Farm - Iceland - environmental sample - boot swabs - Control and eradication programmes - Official and industry sampling - Census	herd/floc k	7	Y	7	0	Salmonella	0
	Gallus gallus (fowl) - parent breeding flocks for egg production line - day-old chicks - Farm - Iceland - environmental sample - boot swabs - Control and eradication programmes - Industry sampling - Census	herd/floc k		N_A	2	0	Salmonella	0
	Gallus gallus (fowl) - parent breeding flocks for egg production line - during rearing period - Farm - Iceland - environmental sample - boot swabs - Control and eradication programmes - Industry sampling - Census	herd/floc k		N_A	2	0	Salmonella	0
	Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Iceland - animal sample - meat juice - Control and eradication programmes - Official sampling - Objective sampling	slaughter animal batch		N_A	1056	275	Salmonella	275
	Turkeys - fattening flocks - before slaughter - Farm - Iceland - environmental sample - boot swabs - Control and eradication programmes - Official and industry sampling - Census	herd/floc k	25	Y	25	0	Salmonella	0
	Turkeys - parent breeding flocks - adult - Farm - Iceland - environmental sample - boot swabs and dust - Control and eradication programmes - Official and industry sampling - Census	herd/floc k	3	Y	3	0	Salmonella	0
	Turkeys - parent breeding flocks - day-old chicks - Farm - Iceland - environmental sample - boot swabs - Control and eradication programmes - Industry sampling - Census	herd/floc k		N_A	2	0	Salmonella	0
	Turkeys - parent breeding flocks - during rearing period - Farm - Iceland - environmental sample - boot swabs - Control and eradication programmes - Industry sampling - Census	herd/floc k		N_A	3	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Dairy products (excluding cheeses) - Border inspection activities - United States - food sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	5	0	Salmonella	0
	Dairy products (excluding cheeses) - milk powder and whey powder - Border inspection activities - Canada - food sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	5	0	Salmonella	0
	Dairy products (excluding cheeses) - milk powder and whey powder - Border inspection activities - United States - food sample - Surveillance - Official sampling - Objective sampling			Gram	10	0	Salmonella	0
	Meat from broilers (Gallus gallus) - carcase - Slaughterhouse - Iceland - food sample - neck skin - Control and eradication programmes - Industry sampling - Census	batch (food/fee d)	25	Gram	822	1	Salmonella Infantis	1
	Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - frozen - Border inspection activities - Thailand - food sample - meat - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	10	0	Salmonella	0
	Meat from pig - carcase - Slaughterhouse - Iceland - food sample - carcase swabs - Control and	slaughte		Not Available	2863	18	Salmonella Brandenburg	4
	eradication programmes - Official sampling - Census	r animal batch					Salmonella Infantis	1
		Datch					Salmonella Kedougou	1
							Salmonella London	1
							Salmonella spp., unspecified	2
							Salmonella Typhimurium	1
							Salmonella Typhimurium, monophasic	6
							Salmonella Worthington	2
	Meat from turkey - carcase - Slaughterhouse - Iceland - food sample - neck skin - Control and eradication programmes - Industry sampling - Census	batch (food/fee d)	25	Gram	64	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	All feedingstuffs - Feed mill - Iceland - environmental sample - dust - Surveillance - Industry	batch	25	Gram	341	5	Salmonella Enteritidis	2
	sampling - Selective sampling	(food/fee d)					Salmonella Infantis	1
							Salmonella Mikawasima	1
							Salmonella Stanley	1
	All feedingstuffs - Feed mill - Iceland - environmental sample - dust - Surveillance - Official sampling - Selective sampling	batch (food/fee d)	25	Gram	42	1	Salmonella Worthington	1
	Feed material of marine animal origin - fish meal - Feed mill - Iceland - environmental sample - dust - Surveillance - Industry sampling - Selective sampling	batch (food/fee d)	25	Gram	615	2	Salmonella Montevideo	2
	Pet food - final product - pelleted - Border inspection activities - United States - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	10	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit		Total units positive	Zoonoses	N of units positive
Not Available	Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Iceland - animal sample - organ/tissue - Monitoring - Official sampling - Census	animal	77603	0	Trichinella	0
	Solipeds, domestic - horses - Slaughterhouse - Iceland - animal sample - organ/tissue - Monitoring - Official sampling - Census	animal	7669	0	Trichinella	0

FOODBORNE OUTBREAKS TABLES

Foodborne Outbreaks: summarized data

Strong Foodborne Outbreaks: detailed data

Weak Foodborne Outbreaks: detailed data

ANTIMICROBIAL RESISTANCE TABLES FOR CAMPYLOBACTER

Table Antimicrobial susceptibility testing of Campylobacter jejuni in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm		Sampling Type: animal	sample - faeces	Sampling Conte	xt: Control and eradication pro	ogrammes	
Sampler: Industry samp	bling	Sampling Strategy: Cen	sus	Programme Coc	le: OTHER AMR MON		
Analytical Method: Dilut	tion - sensititre						
Country of Origin: Icelar							
	iu						
Sampling details: N_A							
	AM substance	Ciprofloxacin 9.0	 Erythromycin (Erythromycin A) 	Gentamicin 5	Nalidixic acid 16	5 Streptomycin	L Tetracycline
	Lowest limit	0.12	1	0.12	1	0.25	0.5
	Highest limit N of tested isolates	<u>16</u> 7	128 7	<u>16</u> 7	64 7	<u>16</u> 7	<u>64</u> 7
МІС	N of resistant isolates	2	0	0	2	0	0
<=0.12		5		4			
<=0.25						1	
0.25				3			
<=0.5							7
0.5						1	
<=1			7				
1						3	
2						2	
4					5		
8		2					
>64					2		

Table Antimicrobial susceptibility testing of Campylobacter jejuni in Gallus gallus (fowl) - broilers

Sampling Stage: Slaug	hterhouse	Sampling Type: anima	al sample - caecum	Sampling Conte	xt: Control and eradication p	rogrammes	
Sampler: Industry sam	ipiing	Sampling Strategy: Se	elective sampling	Programme Cod	e: Amr mon		
Analytical Method: Dilu	ution - sensititre						
Country of Origin: Icela	and						
Sampling details: Sampling o	only during high risk summer months						
	AM substance	Ciprofloxacin	Erythromycin (Erythromycin A)	Gentamicin	Nalidixic acid	Streptomycin	Tetracycline
	ECOFF	0.5	4	2	16	4	1
	Lowest limit	0.12	1	0.12	1	0.25	0.5
	Highest limit	16	128	16	64	16	64
	N of tested isolates	16	16	16	16	16	16
МІС	N of resistant isolates	2	•				1
			0	0	2	0	1
<=0.12		13	U	0 10	2		1
<=0.25		13	<u> </u>	10	2	0	1
<=0.25 0.25			<u> </u>	-	2		
<=0.25 0.25 <=0.5		13	<u> </u>	10 5	2	1	1
<=0.25 0.25 <=0.5 0.5		13		10			
<=0.25 0.25 <=0.5		13	0 16	10 5	2	6	
<=0.25 0.25 <=0.5 0.5 <=1 1		13		10 5		1	
<=0.25 0.25 <=0.5 0.5 <=1		13		10 5	1	6 8	
<=0.25 0.25 <=0.5 0.5 <=1 1 2 4 8		13		10 5	1	6 8	
<=0.25 0.25 <=0.5 0.5 <=1 1 2 4		13		10 5	1 6 5	6 8	

ANTIMICROBIAL RESISTANCE TABLES FOR SALMONELLA

Table Antimicrobial susceptibility testing of Salmonella Agona in Gallus gallus (fowl) - broilers

Sampling Stage: Farm

Sampler: Official and industry sampling

Analytical Method: Dilution - sensititre

Country of Origin: Iceland

Sampling Details: N_A

Sampling Type: environmental sample - boot swabs

Sampling Strategy: Census

Sampling Context: Control and eradication programmes Programme Code: AMR MON

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.06	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1											1
<=0.5					1				1						
0.5														1	
<=1		1						1							
4			1										1		
<=8						1									
8											1				
64												1			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from broilers (Gallus gallus) - carcase

Sam Ana	pling Stage: Slau pler: Industry sa lytical Method: D	mpling ilution - se				ling Type: fo	ood sample - y: Census	neck skin		proc	rammes	xt: Control ar e: AMR MON		in	
Cou	ntry of Origin: Ic	eland													
Samp	oling Details: N_A														
	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.06	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.03										1					
0.03							1								
<=0.25				1											
<=0.5					1				1						
0.5														1	1
<=1								1							
2		1									4				
<=4 4			1								1		1		
4			1			1							I		
64												1			
U T															

Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - broilers

Samp Analy	pling Stage: Farm pler: Official and ytical Method: Di ntry of Origin: Ice	l industry s Dilution - se				pling Type: e pling Strategy	environmental jy: Census	sample - b	oot swabs			xt: Control an le: AMR MON		'n	
	ling Details: N_A														
	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.06	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1											1
<=0.5					1				1						
0.5 <=1		1						1						1	
<=2			1												
<=4											1				
4													1		
<=8						1									
64												1			

 Table Antimicrobial susceptibility testing of Salmonella Worthington in Gallus gallus (fowl) - broilers

Analy	oler: Official and /tical Method: D .try of Origin: Ic	ilution - se				ling Strateg	environmental y: Census	sample - Di	JUL SWADS	prog	pling Conte rammes ramme Cod	e: AMR MON			
Sampl	ing Details: N_A														
	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.06	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1											1
<=0.5					1				1					4	
0.5 <=1		1						1						1	
<=2		l											1		
<=4											1		1		
4			1												
<=8						1									
16												1			

ANTIMICROBIAL RESISTANCE TABLES FOR INDICATOR ESCHERICHIA COLI

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Pigs - fattening pigs

Sampling Stage: SlaughterhouseSampling Type: animal sample - caecumSampling Context: MonitoringSampler: Official samplingSampling Strategy: Objective samplingProgramme Code: AMR MONAnalytical Method: Dilution - sensititreVertical SamplingVertical Sampling

Country of Origin: Iceland

Sampling Details: N_A

Sulfamethoxazole Chloramphenicol Trimethoprim Azithromycin Ciprofloxacin Nalidixic acid Tetracycline Meropenem Gentamicin Tigecycline AM Ceftazidim Cefotaxim Ampicillin substance Colistin ECOFF 0.25 0.5 0.064 0.125 0.25 0.5 Lowest limit 0.015 0.5 0.03 0.25 0.25 **Highest limit** N of tested isolates N of resistant MIC isolates <=0.015 <=0.03 <=0.25 <=0.5 0.5 <=1 <=2 <=4 <=8 >32 >64

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	21	21	21	21	21	21	21	21	21	21	21	21	21	21
MIC	N of resistant isolates	5	0	0	0	1	0	0	0	0	0	5	1	0	4
>1024												5			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Pigs - fattening pigs

Sam	pling Stage: Sla	aughterhous	se		Samp	oling Type: a	nimal sampl	le - caecum		Sam	pling Contex	t: Monitoring
Sam	pler: Official sa	mpling			Samp	oling Strategy	y: Objective	sampling		Prog	ramme Code	: ESBL MON pnl2
Anal	ytical Method:	Dilution - se	ensititre									
Cour	ntry of Origin:	Iceland										
Samp	ling Details: N_A											
	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	mipenem	Meropenem	Temocillin	
	Cefotaxime		-									
	synergy test	Not Available	Not Available	ent	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Negative/Abs ent	Not Available	Not Available	Not Available	Not Available	
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32	
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5	
	Highest limit	32	64	64	64	128	128	2	8	16	64	
	N of tested isolates	7	7	7	7	7	7	7	7	7	7	
МІС	N of resistant isolates	0	7	7	7	7	7	0	0	0	0	
<=0.015	isolales	0		1			/	1	0	0	0	
<=0.015								I		7		
0.03								1		I		
0.064								5				
<=0.12									1			
0.12		7							-			
0.25									5			
0.5									1			
2			2	5		1						
4			5	2			4					
8						5	3				4	
16						1					3	
64					7							

 Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Pigs - fattening pigs

Samp	oling Stage: Slau	ughterhous	se		Samp	oling Type: a	animal sample	e - caecum		Sam	pling Conte	xt: Monitoring	J		
Samp	oler: Official sam	npling			Samp	ling Strateg	y: Objective s	sampling		Prog	ramme Cod	le: ESBL MON	J		
	ytical Method: D		oncititra		-	-	· -			-					
			IISiuu e												I
Cour	ntry of Origin: Ic	eland													
Sampl	ling Details: N_A														
	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	7	7	7	7	7	7	7	7	7	7	7	7	7	7
МІС	N of resistant isolates	7	0	7	7	0	0	0	0	0	0	2	6	0	2
<=0.015							7								
<=0.03										7					
<=0.25 <=0.5									7					4	1
<=0.5 0.5									1					3	4
<=1								7							7
<=2			7												
2				2											
<=4											6				
4				4	1								1		
>4				1											
<=8						7						1			
8					5						1				
>8					1										
16												1			
32												3			2
>64		7											6		
>1024												2			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers

Samp	oling Stage: Slau	ughterhoug	se		Samr	pling Type: a	animal sample		Sam	pling Conte	ext: Monitoring	ıg			
															P
Samp	oler: Official sam	pling			Samp	Jing Strategy	y: Objective s	sampling		Prog	Jramme Cou	de: AMR MON	1		ŗ
Analy	tical Method: Di	ilution - se	ensititre												
Coun	try of Origin: Ice	eland													I
															P
Sampli	ing Details: N_A														
	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit N of tested	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	isolates	94	94	94	94	94	94	94	94	94	94	94	94	94	94
МІС	N of resistant isolates	6	0	0	0	0	1	0	0	0	1	3	7	0	1
<=0.015							91								
<=0.03										94					
0.064 <=0.25							2							37	F 2
<=0.25 0.25				94			1							31	52
<=0.5					94				86						
0.5					0-,									47	36
<=1		4						94							
1									6					10	5
<=2			84										26		
2		37							2						
<=4											85				
4		42	10										54		
<=8						94						19			
8		5									7		7		
16											1	40			
32 >32												25			4
>32 64												7			1
>64		6											2 5		
128		0										2			
120															

	AM substance	Ampicillin	Azithromyci	Cefotaxim	Ceftazidim	Chloramphen	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxa	Tetracycline	Tigecycline	Trimethoprim
E	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
L	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
Н	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	94	94	94	94	94	94	94	94	94	94	94	94	94	94
	N of resistant isolates	6	0	0	0	0	1	0	0	0	1	3	7	0	1
>128											1				
>1024								,				1			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers

Samp	ling Stage: S	laughterhous	se		Samj	pling Type: a	nimal sample	- caecum		Sam	pling Conte	xt: Monitorin	g
Samp	ler: Official s	ampling			Sam	pling Strategy	: Objective s	ampling		Prog	gramme Cod	e: ESBL MO	N pnl2
Analy	tical Method:	Dilution - se	ensititre										
Coun	try of Origin:	Iceland											
Sampli	ing Details: N_A												
	AM substance	Cefepime	Cefotaxim	-	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim		Ceftazidime + Clavulanic acid	Ertapenem	lmipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Positive/Pres ent	Negative/Abs ent	Not Available	Not Available	Not Av	vailable	Not Available	Not Available	Not Available	Not Available
	Coffeeidime	Not Available	Not Available					ositive/Pres	Negative/Ab ent	^S Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	0.25	8	0.5	ent 0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.064	0.5	0.25	0.12	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	64	128	128	128	2	8	16	64
	N of tested							-				-	
	isolates	5	5	5	5	5	5	5	5	5	5	5	5
	N of resistant		_	•	•	•	_		•	•		•	•
MIC	isolates	3	5	3	3	3	5	3	3	0	0	0	0
<=0.015 <=0.03										2		5	
0.03										2		5	
<=0.064				2						-			
0.064				_						1			
<=0.12								2					
0.12		2											
0.25		1									5		
1							1						
2		1					1						
4						2							1
8		1	3		3		2		3				4
16			1				1						
32						3							
aland 2016	-								70				

AM substance	Cefepime	Cefotaxim	janina (janina) janina (janina)		Cefoxitin	Ceftazidim		Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
Cefotaxime synergy test	Not Available	Not Available	Positive/Pres ent	Negative/Abs ent	Not Available	Not Available	Not A	vailable	Not Available	Not Available	Not Available	Not Available
Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	ositive/Pres	s Negative/Ab ent	^S Not Available	Not Available	Not Available	Not Available
ECOFF	0.125	0.25	0.25	0.25	8	0.5	0.5	0.5	0.06	0.5	0.125	32
Lowest limit	0.064	0.25	0.064	0.064	0.5	0.25	0.12	0.12	0.015	0.12	0.03	0.5
Highest limit	32	64	64	64	64	128	128	128	2	8	16	64
N of tested isolates	5	5	5	5	5	5	5	5	5	5	5	5
N of resistant isolates	3	5	3	3	3	5	3	3	0	0	0	0
		1										

Iceland - 2016

MIC 64 Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers

Sampling Stage: Slaughterhouse					Sampling Type: animal sample - caecum						Sampling Context: Monitoring					
Sami	Sampler: Official sampling					Sampling Strategy: Objective sampling					Programme Code: ESBL MON					
				Sampling Strategy: Objective Sampling												
Anal	ytical Method: D	ensititre														
Cour	ntry of Origin: Ic	eland														
Samp	ling Details: N_A															
	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim	
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2	
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25	
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32	
	N of tested isolates	5	5	5	5	5	5	5	5	5	5	5	5	5	5	
MIC	N of resistant isolates	5	0	5	5	0	0	0	0	0	0	2	0	0	0	
<=0.015							5									
<=0.03										5						
<=0.25														3	5	
<=0.5									5							
0.5														2		
<=1								5								
1					2											
<=2			5										1			
<=4											4					
4													3			
>4				5												
<=8						5										
8					1						1		1			
>8					2							2				
16 >64		5										3				
>64 >1024		5										2				
>1024												۷				

OTHER ANTIMICROBIAL RESISTANCE TABLES

Specific monitoring of ESBL-/AmpC-/carbapenemase-producing bacteria and specific monitoring of carbapenemase-producing bacteria, in the absence of isolate detected



Latest Transmission set

Table Name	Last submitted dataset transmission date
Antimicrobial Resistance	12-Oct-2017
Animal Population	27-Jun-2017
Disease Status	27-Jun-2017
Prevalence	29-Jun-2017
Text Forms	28-Jun-2017