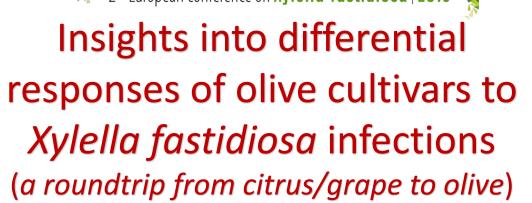






multidisciplinary-Oriented Research Strategy

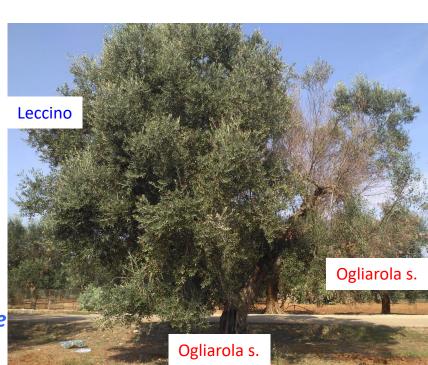




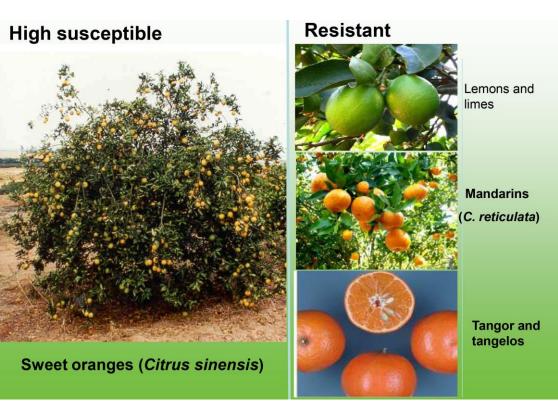
PASQUALE SALDARELLI

CNR, Istituto per la Protezione sostenibile delle piante Bari, Italy





Xylella resistance in citrus (citrus variegated chlorosis)



RESEARCH ARTICLE

Open Access

RNA-Seq analysis of *Citrus reticulata* in the early stages of *Xylella fastidiosa* infection reveals auxin-related genes as a defense response

Carolina M Rodrigues¹, Alessandra A de Souza¹, Marco A Takita¹, Luciano T Kishi² and Marcos A Machado^{1*}

Up-regulated Leucine-rich repeat receptor-like kinase (LRR-RLK) At4G08850, At3G47570

Differential colonization patterns of Xylella fastidiosa infecting citrus genotypes

B. Niza^{ab}, H. D. Coletta-Filho^a, M. V. Merfa^{ab}, M. A. Takita^a and A. A. de Souza^a*

- Colonization impaired in resistant
- No different xylem morphology
- Lignin deposition

Brasile

credits A. De Souza

Xylella resistance in Vitis, Walker, 2017

- V. arizonica/candicans b43-17 has single dominant gene for resistance to PD and it's homozygous
- All progeny from crosses to b43-17 are resistant to PD
- Genetically mapped PD resistance (*PdR1*), to chromosome 14. Linked markers have been used for marker-assisted selection (MAS)

Controlling Pierce's Disease with Molecular and Classical Breeding

M. Andrew Walker



Features of Xylella resistance in Vitis, 2019

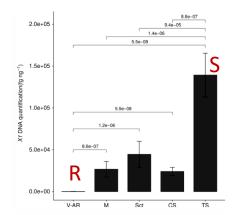
Assessment of Pierce's disease susceptibility in Vitis vinifera cultivars with different pedigrees

E. Deyett^{a†}, J. Pouzoulet^{b†}, J.-I. Yang^c, V. E. Ashworth^a, C. Castro^d, M. C. Roper^d and P. E. Rolshausen^a*



Disease severity

Pathogen abundance

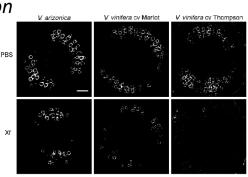


Genetic

In b43-17 V.arizonica/candicans PdR1 locus contains candidate Leucine-rich repeat receptor-like kinase receptors LRR-RLKs

Smaller vessel diameters=resistance % of vascular occlusion

Anatomic



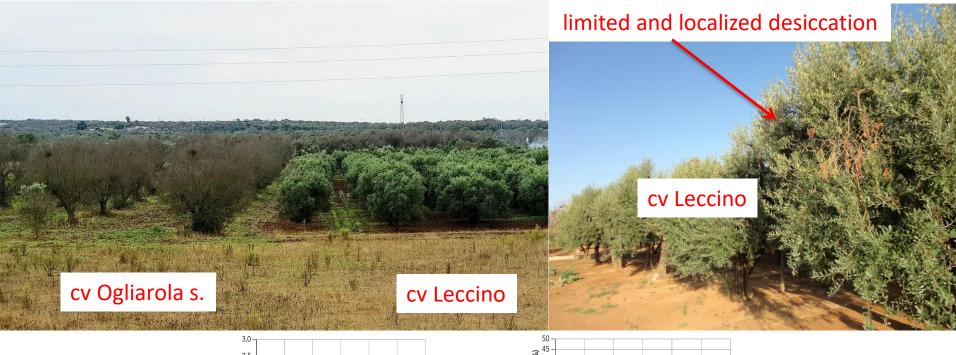
Features of Xylella resistance in Vitis, 2011

Assessment of the Process of Movement of *Xylella fastidiosa*Within Susceptible and Resistant Grape Cultivars

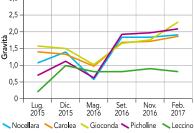
C. Baccari and S. E. Lindow

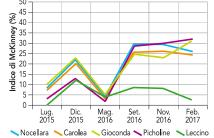
STEM

- ❖ Lower population sizes in resistant cv compared to susceptible
- ❖ Few vessel colonized in resistant cv compared to susceptible

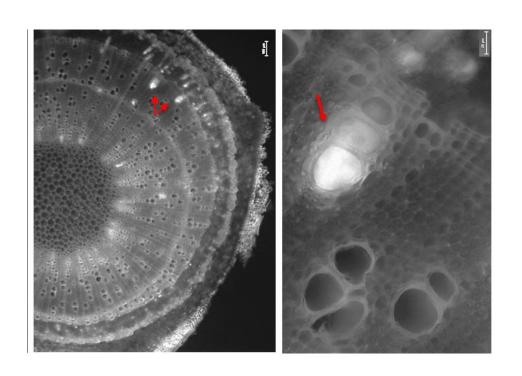


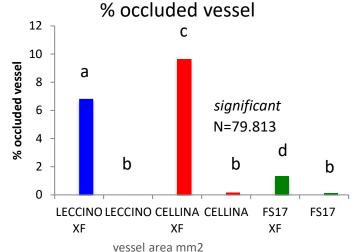
Disease severity

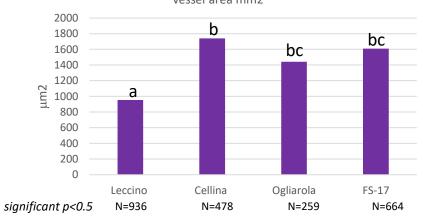




Anatomic







Pathogen abundance



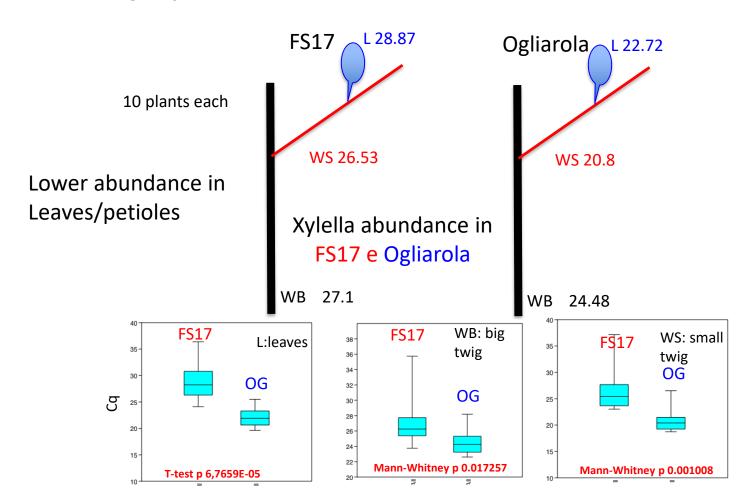
TABELLA 2 - Risultati dell'analisi molecolare su piante di FS-17°, Kalamata, Leccino e Ogliarola salentina				
Cultivar	Positivi/Totale	Cq media positiva	Assorbanza media positiv	
FS-17®	18/51	29,59 cicli	5,04·10⁴ UFC/mL	
Ka l amata	28/51	27,21 cicli	3,16·10⁵ UFC/mL	

Kalamata, Leccino e Ogliarola salentina				
Cultivar	Positivi/Totale	Cq media positiva	Assorbanza media positivi	
FS-17®	18/51	29,59 cicli	5,04·10⁴ UFC/mL	
Kalamata	28/51	27,21 cicli	3,16·10⁵ UFC/mL	
Leccino	9/18	28,41 cic l i	9,93·10⁴ UFC/mL	
Ogliarola salentina	10/10	26,41 cic l i	4,51·10° UFC/mL	

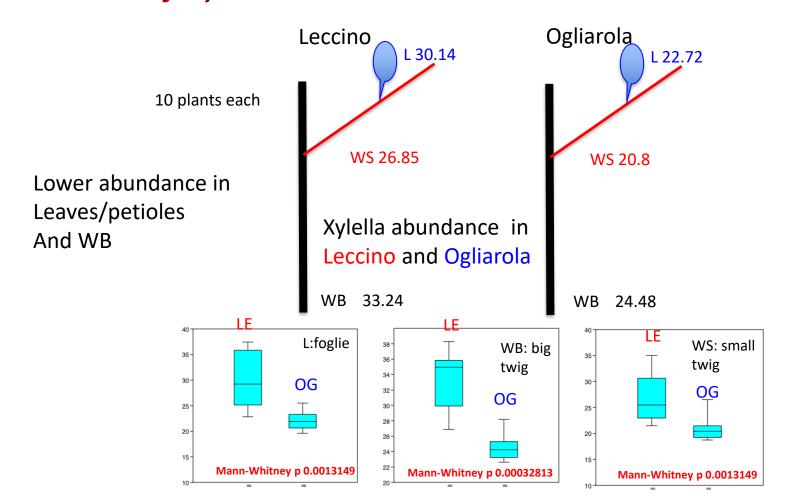




Pathogen abundance

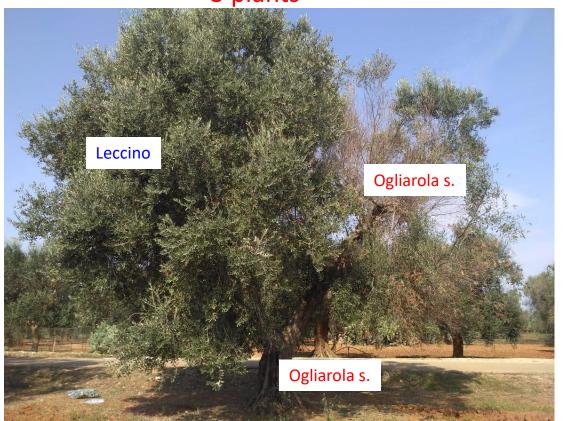


Pathogen abundance



Leccino resistance: further evidences

Leccino grafted on Ogliarola in the past 3 plants



Xf CFU/ml

Leccino1 1x10⁴ Ogliarola1 8x10⁵

Leccino2 5x10² Ogliarola2 1x10⁶

Leccino3 Ogliarola3 9x10⁵

N/D

Genetic



Ogliarola Xylella

Ogliarola, Leccino Healthy

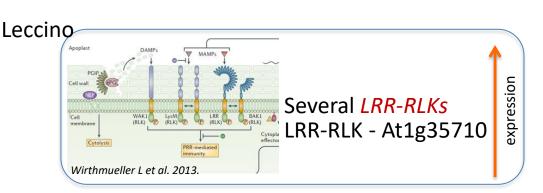
condition

htty

in its

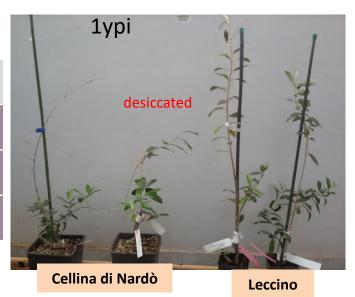
early light-induced proteins (ELIP)
Late embryogenesis abundant protein (LEA)
Expansin- like B1 (osmoprotectant)
ABA2 (abscisic acid)
deidrina (LEA protein)

Ogliarola suffers a drought stress upon Xf infection



Transcriptome studies of greenhouse trials

Xf strain	trial		
CO33	3x Cellina	3x Leccino	2x Mock
De Donno	3x Frantoio		2x Mock
De Donno	3x Fs17		2x Mock



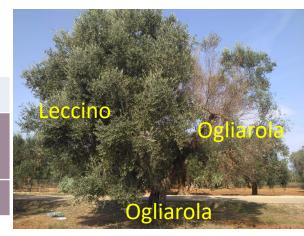
Late (cronic) infections

Genetic

Transcriptome studies of different field trials



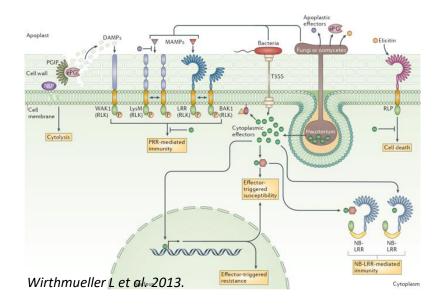
trial			
2.1	2 0-1:1-	A. Marali	
3x Leccino	3x Ogliarola	4x Mock	
3x Ogliarola grafted Leccino			
	3x Leccino 3x Og	3x Leccino 3x Ogliarola	





Late (cronic) infections

Summary of transcriptome studies



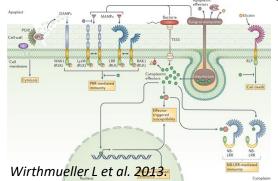
Common traits

Leucine-rich repeat receptor-like kinase receptors

LRR-RLKs UP-Regulated

- At1g35710
- At3g47570
- At4g08850

Summary of transcriptome studies

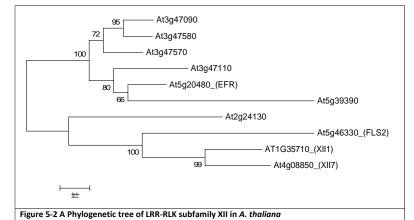


common traits

Leucine-rich repeat receptor-like kinase receptors

LRR-RLKs UP-Regulated
At1g35710, At3g47570, At4g08850

LRR XII

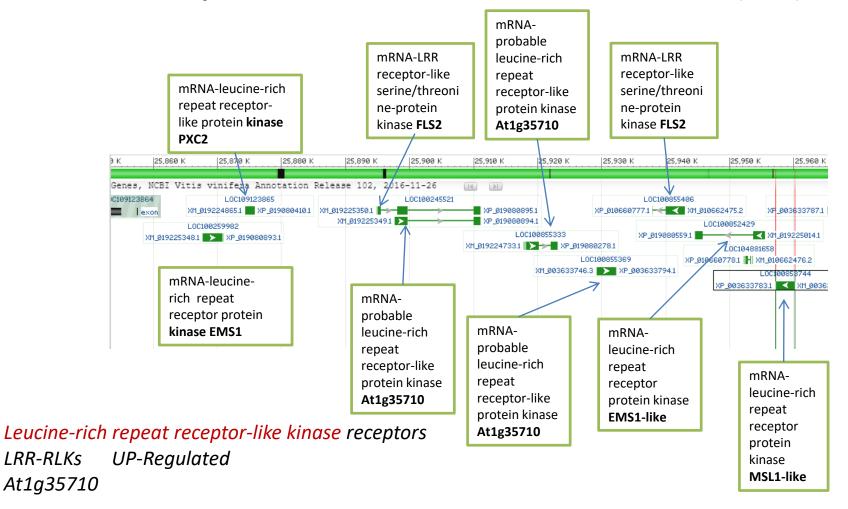


similar to well characterized LRR-RLKs recognizing bacteria components

FLS2: Flagellin sensing 2

EFR: EF-Tu Receptor

Vitis Vinifera NCBI Annotation, Pierce disease resistance locus (PdR1)



Citrus (early infection)

Rodrigues et al. BMC Genomics 2013, 14:676 http://www.biomedcentral.com/1471-2164/14/676 Page 4 of 13

Table 1 Differentially expressed genes in Ponkan mandarin in response to infection by X. fastidiosa

Gene symbol	gene id_Citrus clementina*	AGI**	Fold change***	P ≤ 0.001	Gene description
LRR-RLK	Cidev10004108m	AT4G08850	1.05611	0.000755101	leucine-rich repeat family protein
LRR-RLK	Cidev10014130m	AT3G47570	2.64522	3.84525e-09	leucine-rich repeat protein kinase family protein
RLP12	Cidev10003540m	AT1G71400	1.20144	2.69007e-06	receptor like protein 12
CC-NBS-LRR	Ciclev10007304m	AT4G27190	1.29361	4.28293e-06	nb-arc domain-containing disease resistance protein
	A	erecessors.			4.4

Leucine-rich repeat receptor-like kinase receptors

LRR-RLKs UP-Regulated At3g47570, At4g08850



RESEARCH ARTICLE

The Arabidopsis leucine-rich repeat receptor kinase MIK2/LRR-KISS connects cell wall integrity sensing, root growth and response to abiotic and biotic stresses



Dieuwertje Van der Does¹, Freddy Boutrot¹, Timo Engelsdorf², Jack Rhodes¹, Joseph F. McKenna^{3na}, Samantha Vernhettes⁴, Iko Koevoets⁵, Nico Tintor⁶, Manikandan Veerabagu², Eva Miedes⁷, Cécile Segonzaca^{1nb}, Milena Roux^{1nc}, Alice S. Breda⁸, Christian S. Hardtke⁸, Antonio Molina⁷, Martijn Rep⁶, Christa Testerink⁵, Grégory Mouille⁴, Herman Höfte⁴, Thorsten Hamann^{2,3}, Cyril Zipfel¹*

MIK2/LRR-KISS is a component of the cell-wall integrity (CWI) sensing

MIK2/LRR-KISS is a LRR-RLK encoded by **At4g08850**

MIK2 belong to the XIIb sub-family of LRR-RLKs and is homologous (60%aa identity) to **At1g35710**

Working Hypotesis

MIK2 (At4g08850 similar to At1g35710): regulator of cell wall damage responses, is a CWI sensor

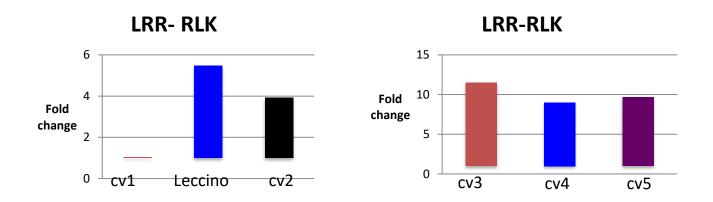
Xylella has a set of Cell Wall Degrading Enzymes (CWDE)

Disruption of Cell Wall Integrity >stress responses (ROS JA,SA, ET, lignin, callose, pect methyl-esterification)

Responses reminiscent of the Plant's Defence Reaction to pathogens and insects

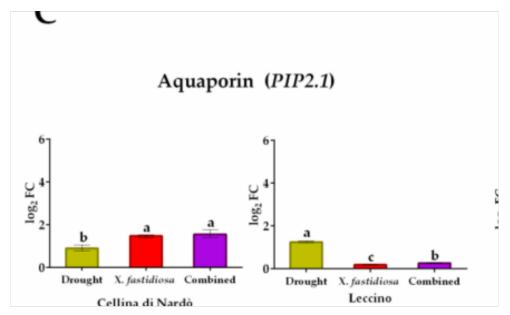
MIK2 is required for resistance to the fungal root pathogen Fusarium oxysporum, which multiplies in the xylem and produce CWDE

LRR-RLK detection in cultivars with diverse susceptibility/resistance to Xyella



greenhouse

ongoing







Article

Molecular Effects of *Xylella fastidiosa* and Drought Combined Stress in Olive Trees

Mariarosaria De Pascali ¹, Marzia Vergine ^{1,*}, Erika Sabella ¹, Alessio Aprile ¹, Eliana Nutricati ¹, Francesca Nicolì ¹, Ilaria Buja ², Carmine Negro ¹, Antonio Miceli ¹, Patrizia Rampino ¹, Luigi De Bellis ¹, and Andrea Luvisi ¹

Why do we say "resistance"

SUSCEPTIBLE: unable to resist to the infection of a specific pathogen

IMMUNE: does not allow infection by a specific pathogen

TOLERANT: able to contain the effects of an infection regardless of the pathogen replication

RESISTANT: able to limit the pathogen replication







Conclusions

Strong analogies with resistance in grape (citrus)

- Pathogen abundance
- Percentage of occluded vessels
- Area of xylem vessels
- LRR-RLK involvment

ongoing

- Acquiring information on olive gene locus involved
- > Xylella degrades cell-wall
- Degradation is sensed by specific LRR-RLKs (cell-wall integrity sensing)











Acknowledgments

Raied Abou Kubaa Annalisa Giampetruzzi Giuseppe Altamura Stefania Zicca Donato Boscia Maria Saponari





