

WEBINAR 1 DECEMBER 2020
15:00 – 16:00 GMT+1

Pest surveys following an outbreak: delimiting and buffer zone surveys

Stephen Parnell¹– Ignazio Graziosi²

¹University of Salford, UK; ²ALPHA unit, EFSA



Webinar guide for attendees

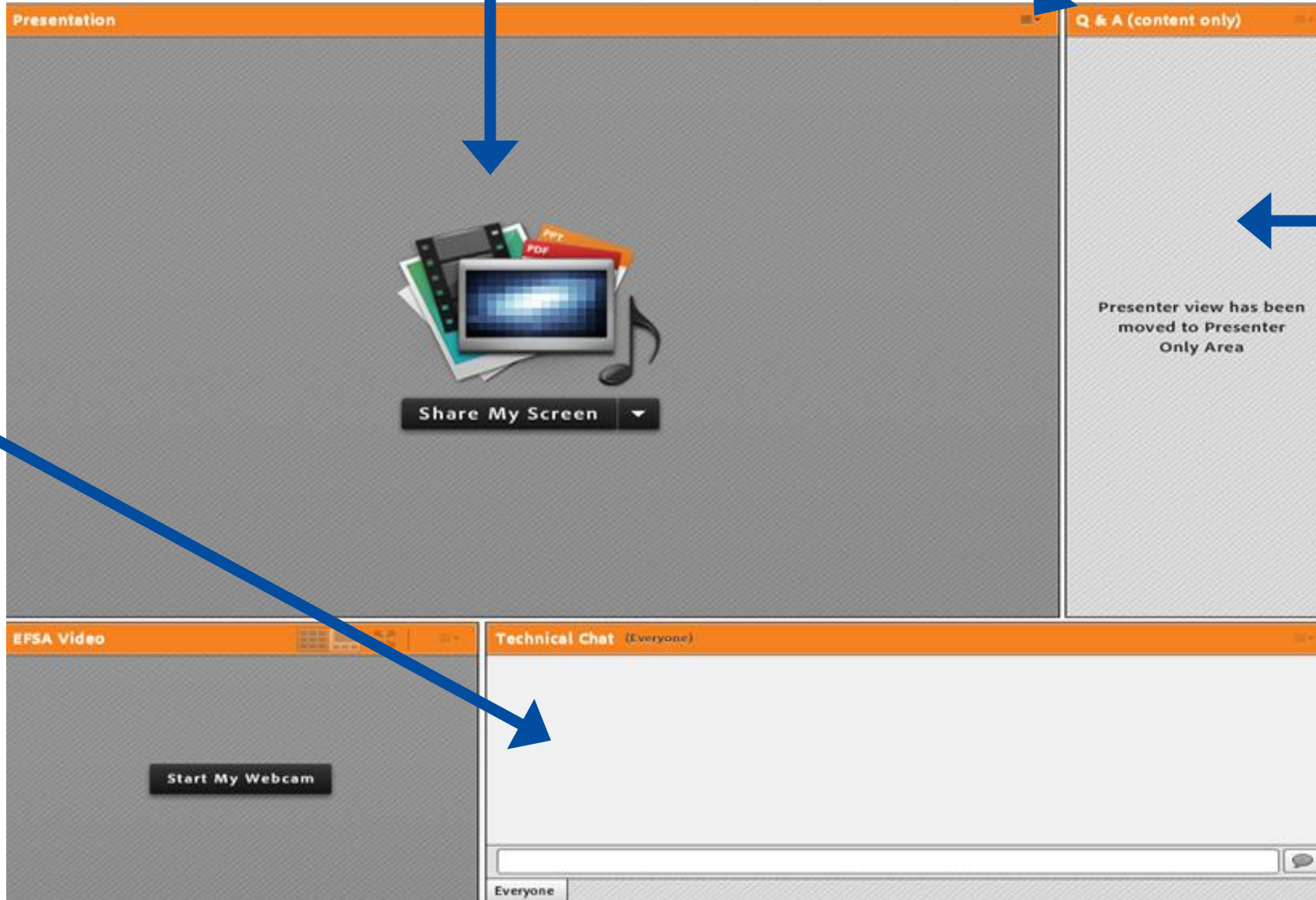
- This webinar is being recorded!
- The webinar is in English and questions should be submitted in English through the platform
- To communicate with us use the chat boxes



Presentation window



Technical box:
technical issues only



The screenshot shows a presentation window with an orange header bar. The main content area is grey and contains a central graphic of a laptop with documents and a 'Share My Screen' button. To the right is a 'Q & A (content only)' panel with a message: 'Presenter view has been moved to Presenter Only Area'. At the bottom, there is an 'EFSA Video' section with a 'Start My Webcam' button and a 'Technical Chat (Everyone)' section with a text input field and a 'Everyone' label.

Q&A box:
any question
related to the topic

Plant pests and diseases



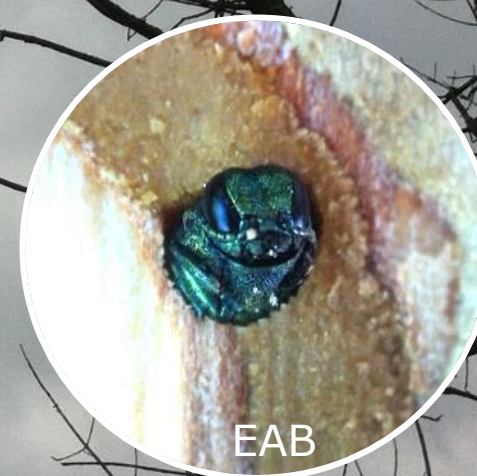
FAW



CBS



JB

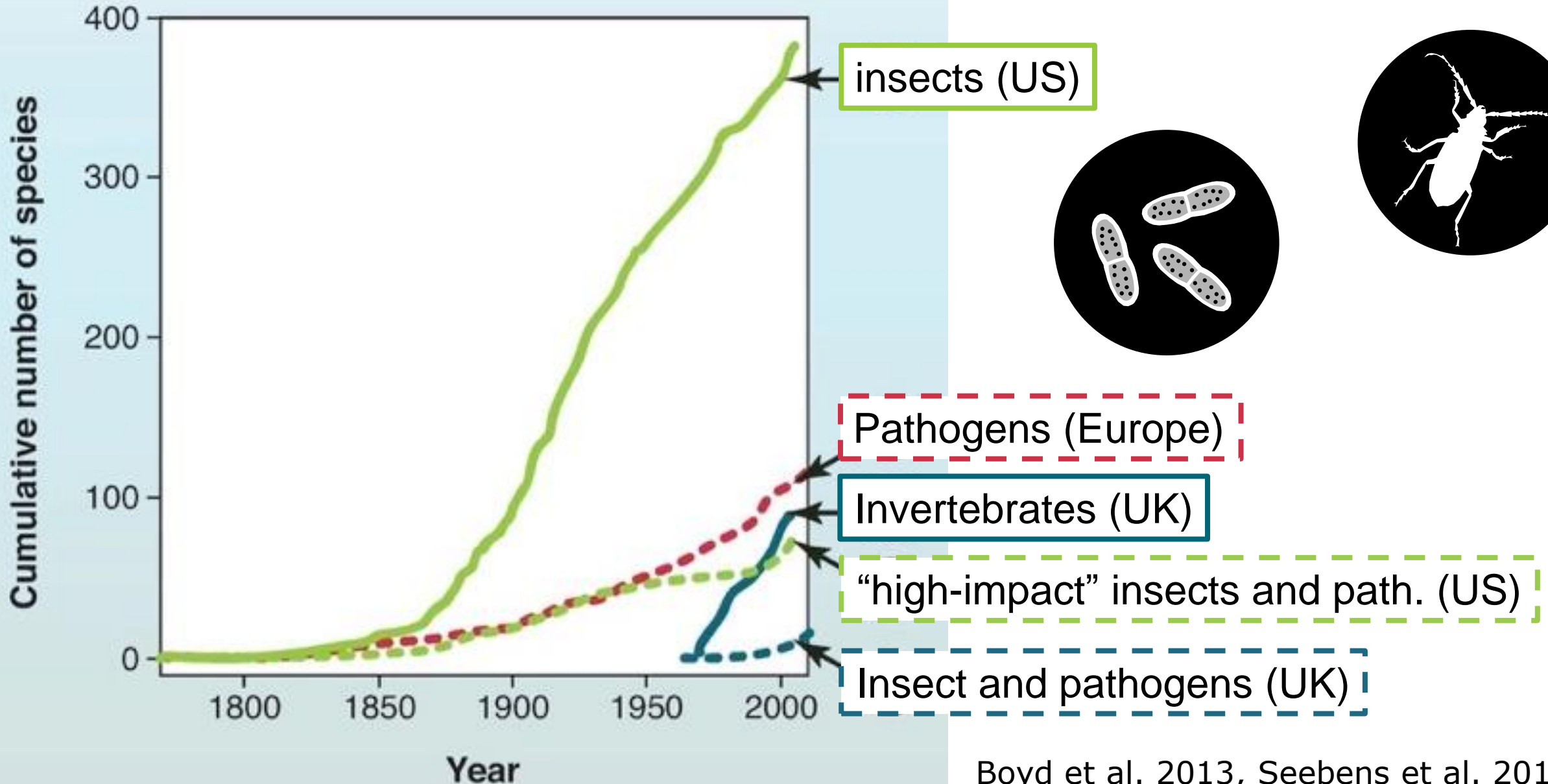


EAB

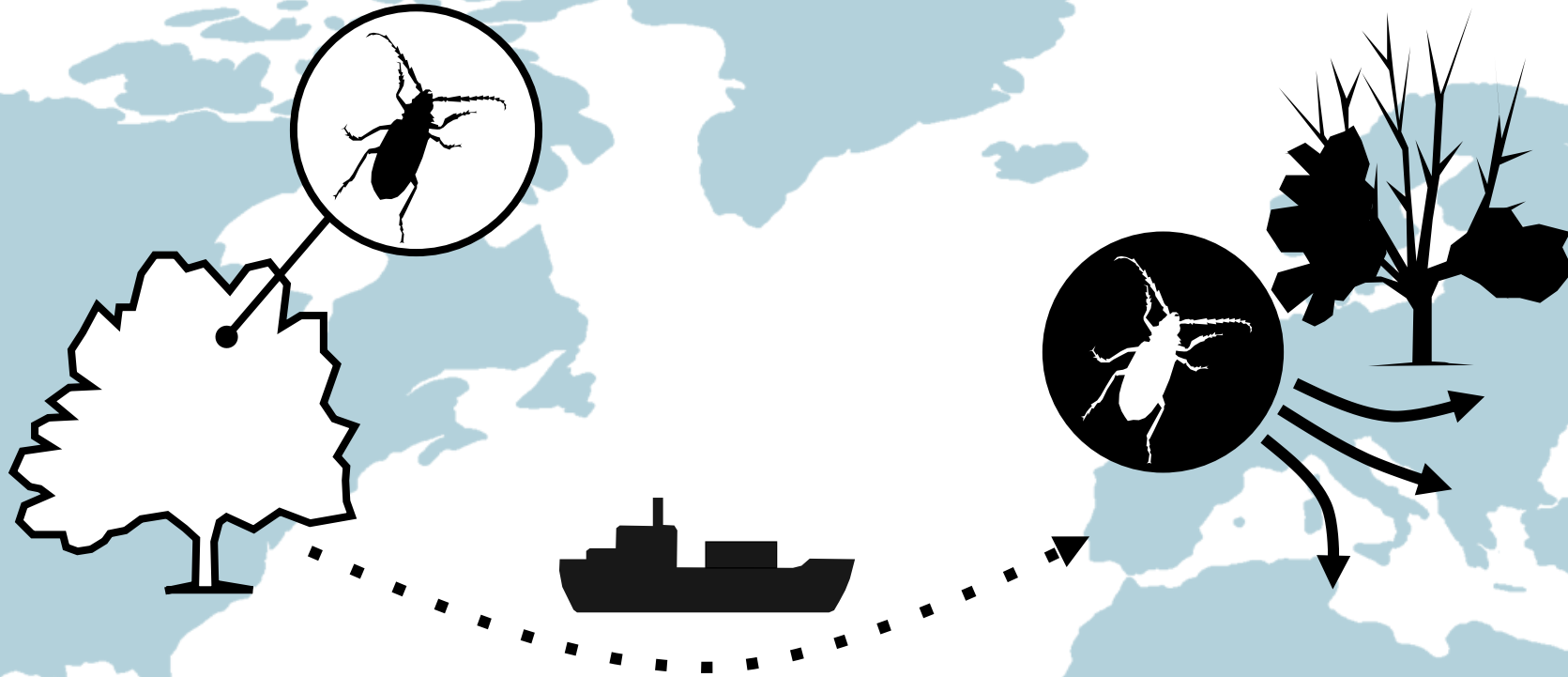


Xylella

Invasive species



Invasive species



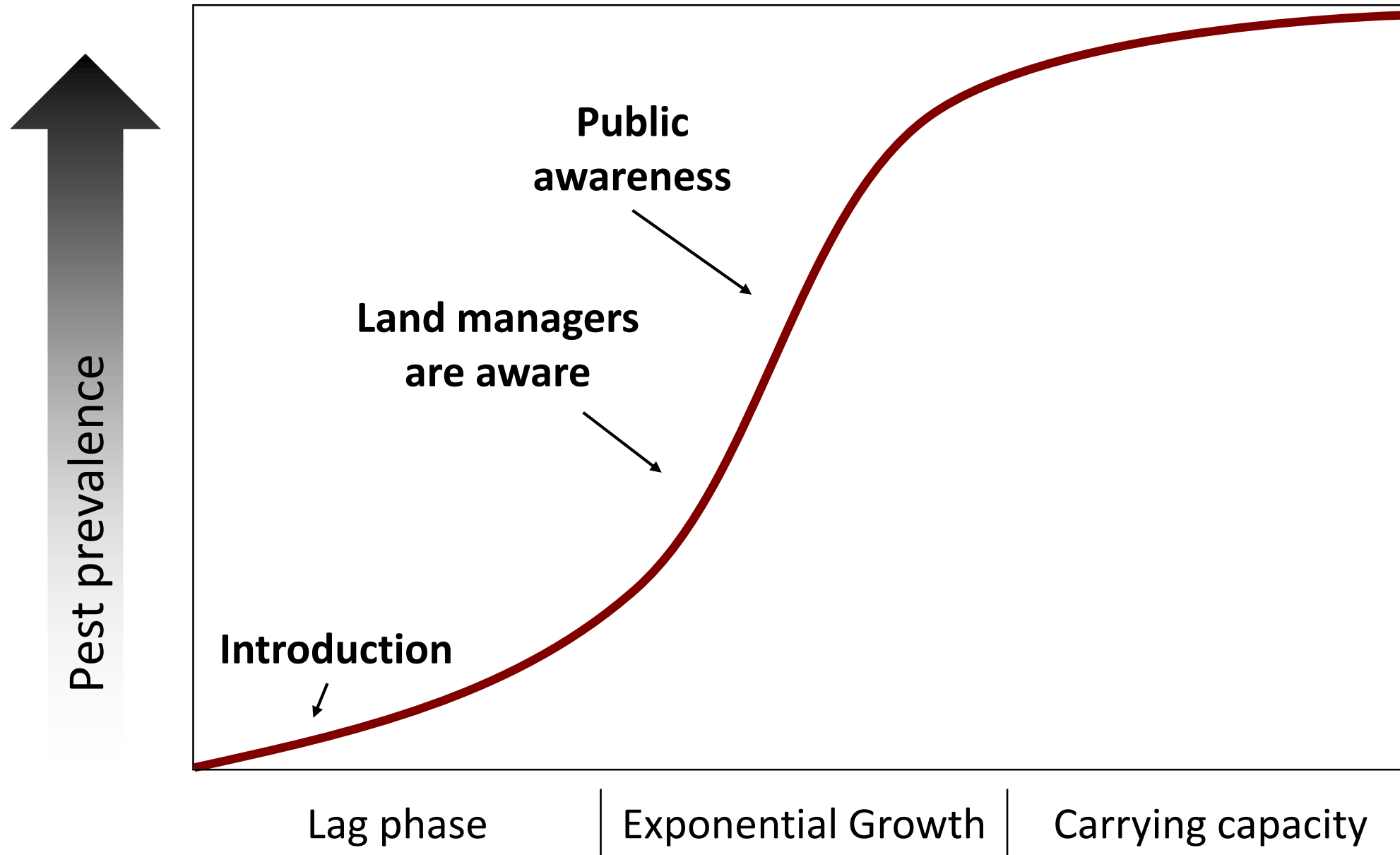
Entry

Establishment

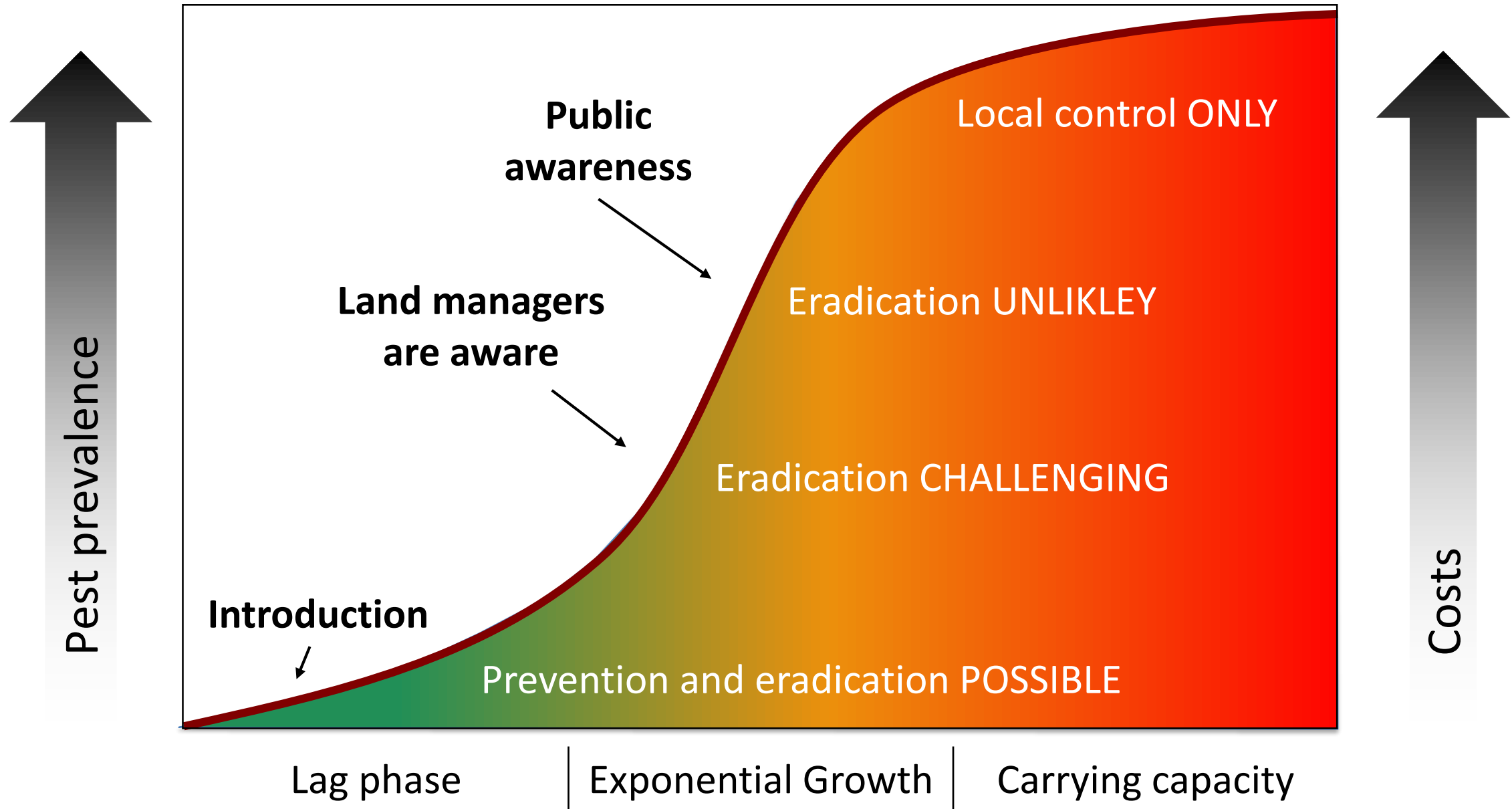
Spread

Impact

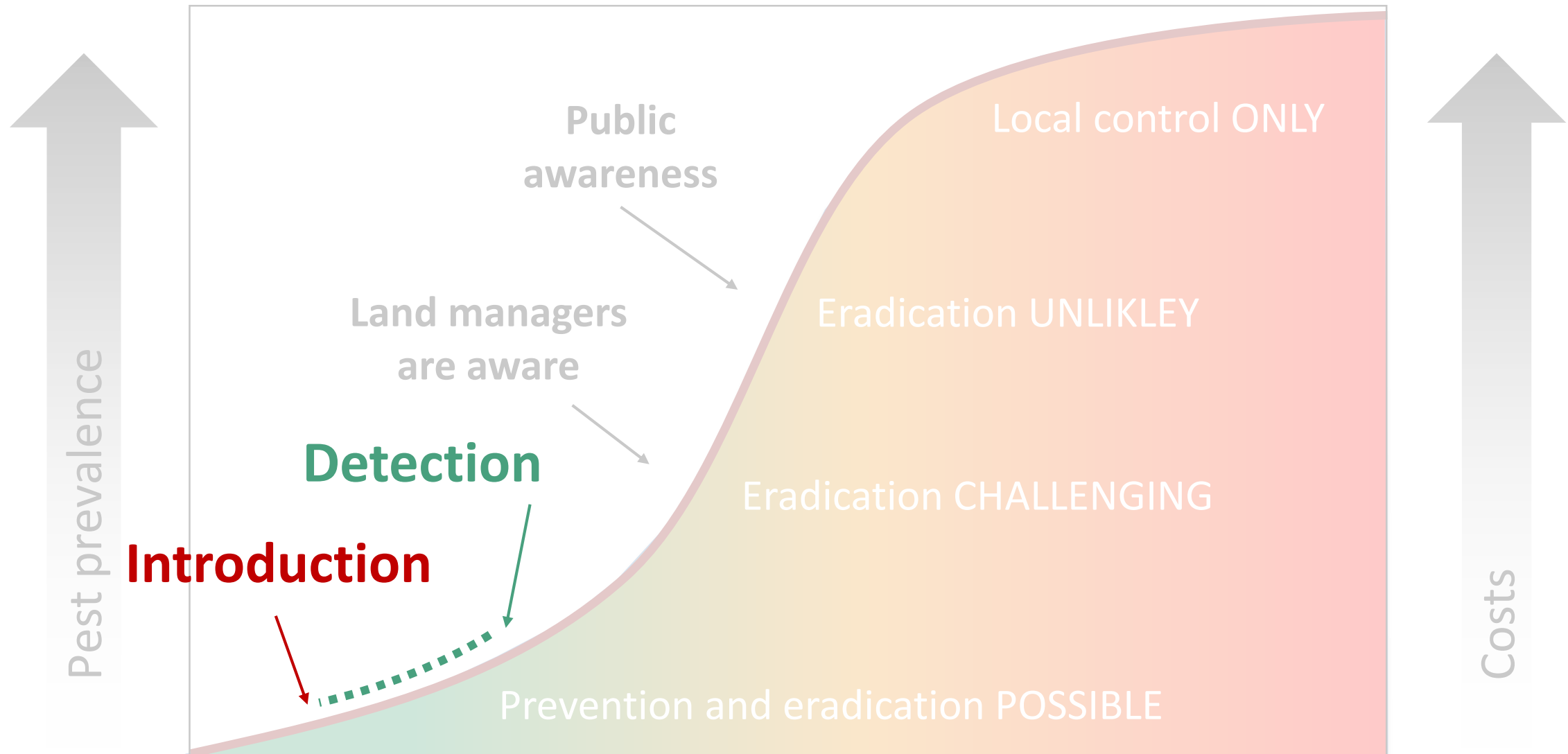
Invasive species



Invasive species



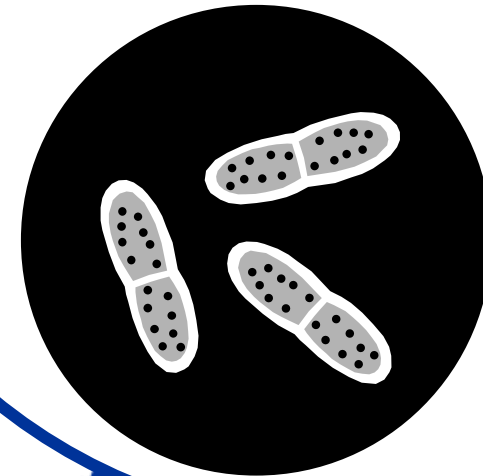
Invasive species



Aukema et al. 2010, Bebber et al. 2013, Bradshaw et al. 2016

Surveillance (IPPC ISPM 5)

EU/2016/2031 - EU/2019/2072 - EU/2019/1702



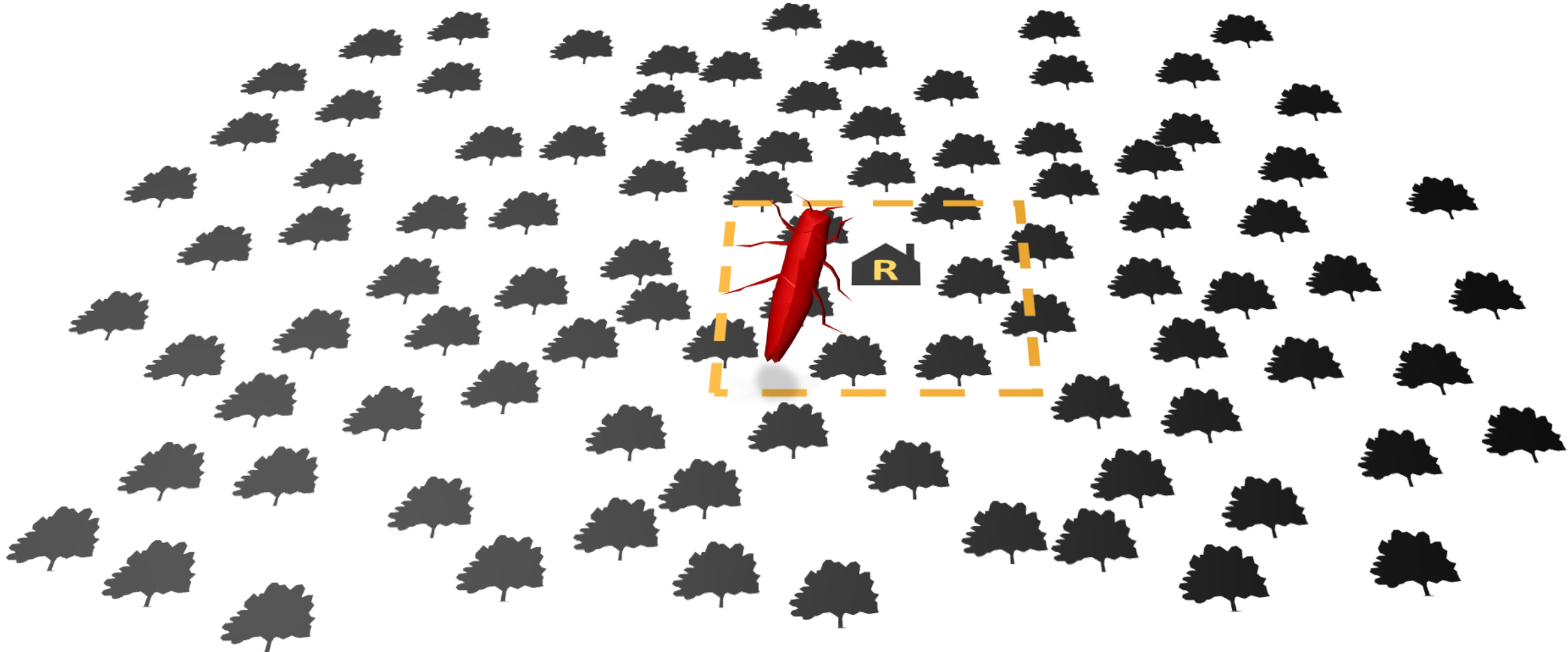
Types of surveys – Why?

Detection



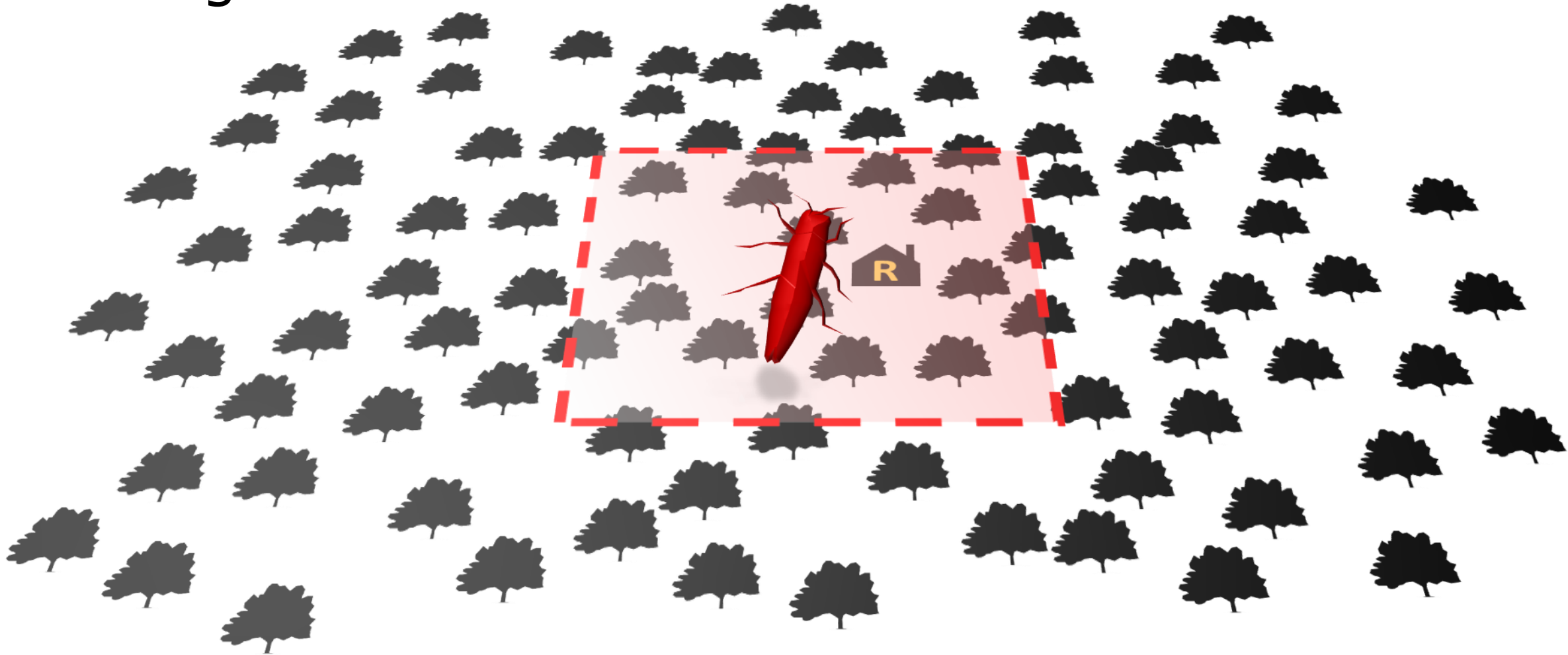
Types of surveys – Why?

Detection



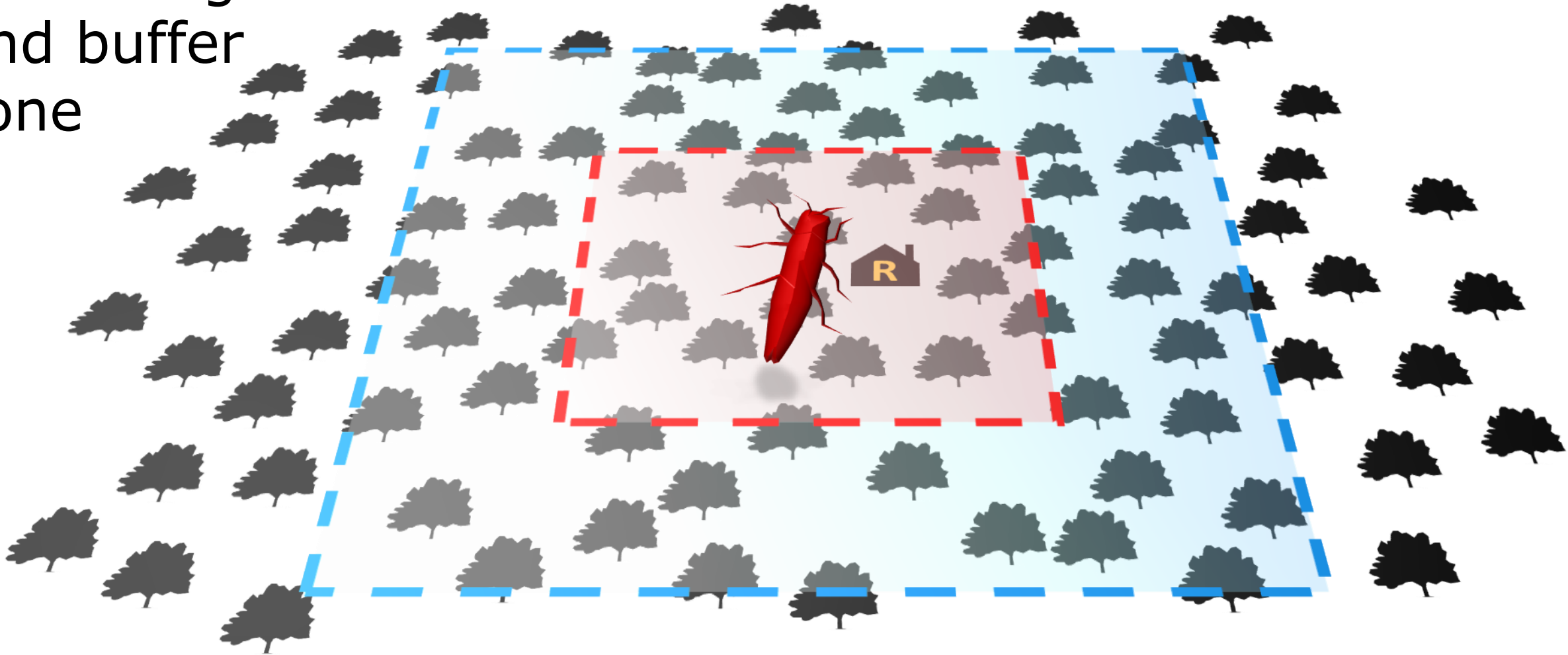
Types of surveys – Why?

Delimiting

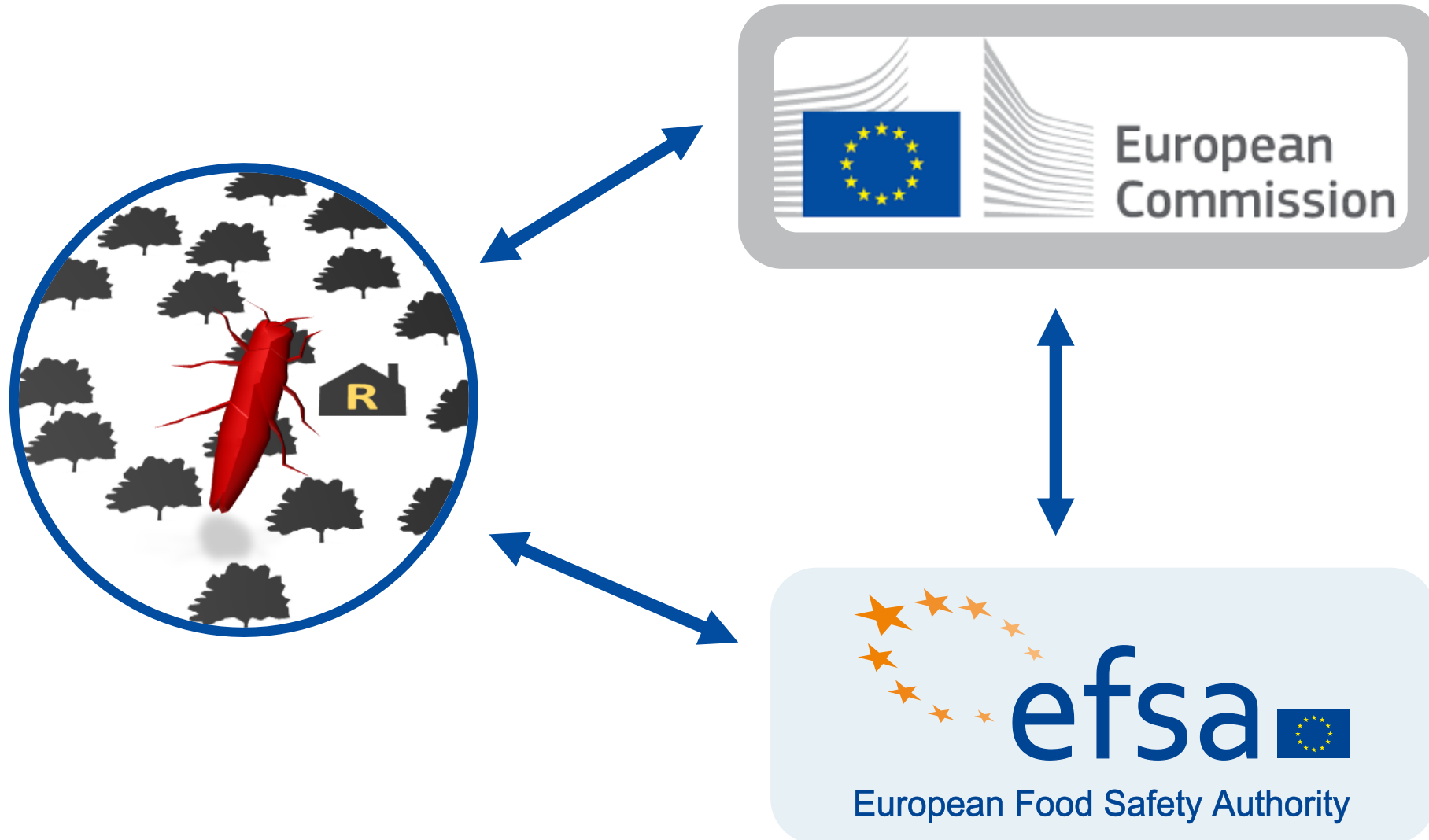


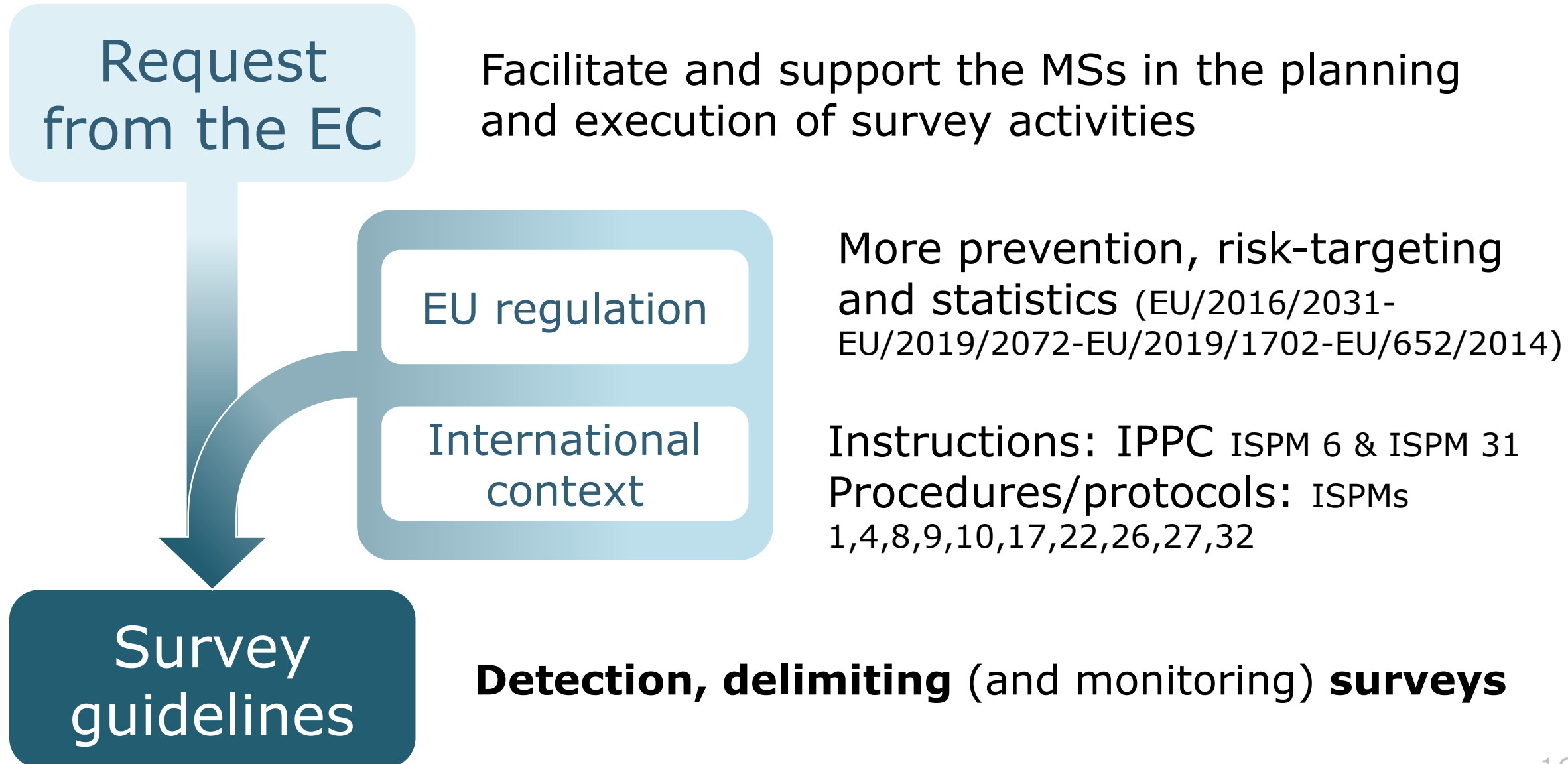
Types of surveys – Why?

Delimiting and buffer zone



EFSA mandate on pest surveys







WHAT

WHERE

WHEN

HOW

HOW MUCH

Survey preparation



PEST SURVEY CARD



APPROVED: 20 October 2020

doi:10.2903/sp.efsa.2020.EN-1945

Pest survey card on *Agrius planipennis*

European Food Safety Authority (EFSA),
Jan Schans, Gritta Schrader, Alice Delbianco, Ignazio Graziosi, Sybren Vos

ArcGIS StoryMaps

**Detection method
&
target population**

WHAT

WHERE

WHEN

HOW

Pest survey card

PEST SURVEY CARD

Abstract.....	1
Introduction.....	4
1. The pest and its biology	5
1.1. Taxonomy	5
1.2. EU pest regulatory status	6
1.3. Pest distribution	7
1.4. Life cycle	10
1.5. Host range and main hosts	13
1.6. Environmental suitability.....	18
1.7. Spread capacity	19
1.8. Risk factor identification	21
2. Detection and identification	25
2.1. Visual examination	25
2.2. Sampling	32
2.3. Pest detection and laboratory testing	34
3. Key elements for survey design	37
References.....	40
Glossary	48
Appendix A: Host plant selection tool.....	52

WHAT

WHERE

WHEN

HOW

Pest survey card

EAB

Trapping method	Effectiveness at low EAB densities	References
Dark green multifunnel traps with (3Z)-hexenol	87.5±12.5%	Francese et al., 2013; USDA APHIS PPQ, 2018; Poland et al., 2019
Double-decker traps with (3Z)-hexenol and manuka oil	100%	Poland and McCullough, 2014; McCullough and Poland, 2017
Green prism traps with (3Z)-hexenol and (3Z)-lactone	75–98%	Ryall et al., 2013; McCullough and Poland, 2017; Parker et al., 2020
Double-decker traps with manuka oil	56–95%	Marshall et al., 2010a, 2010b; McCullough et al., 2011
Green or purple prism traps with (3Z)-hexenol	37–82%	Ryall et al., 2013; Crook et al., 2014; Poland and McCullough,

Detection method

Target population

CBS

	Definition
Target population	Citrus plants growing in orchards, backyards and gardens in each Member State
Epidemiological unit	A single homogeneous area that contains at least one individual host plant (e.g. citrus orchard, backyard or garden)
Inspection unit	A host plant with mature fruits

Sample size & allocate samples to survey area

WHAT

WHERE

WHEN

HOW

HOW MUCH



TECHNICAL REPORT

APPROVED: 31 July 2020

doi:10.2903/sp.efsa.2020.EN-1919

General guidelines for statistically sound and risk-based surveys of plant pests

Guidelines for statistically sound and risk-based surveys of *Xylella fastidiosa*

RIBESS+



*Pest survey card
& story map*

Survey preparation

Characterise
the pest

Characterise host
plant population

Characterise
detection method





Survey guidelines

Set the aim of the survey
Detection – Delimiting – Buffer zone

Design prevalence and
confidence level

Population structure
and size

Method sensitivity

Sample size
(inspections, samples, tests)

Allocate samples to
survey area

Survey design

RiBESS+

*Pest survey card
& story map*

Survey preparation

Characterise
the pest

Characterise host
plant population

Characterise
detection method

Survey guidelines

Set the aim of the survey
Detection – Delimiting – Buffer zone

Design prevalence and
confidence level

Population structure
and size

Method sensitivity

Sample size
(inspections, samples, tests)

Allocate samples to
survey area

Survey design

RiBESS+



Survey cards available...

44
cards

...64
pests

EFSA journal virtual issue

[https://efsa.onlinelibrary.wiley.com/doi/toc/10.1002/\(ISSN\)1831-4732.toolkit-plant-pest-surveillance](https://efsa.onlinelibrary.wiley.com/doi/toc/10.1002/(ISSN)1831-4732.toolkit-plant-pest-surveillance)

- **Pilot organisms**
- Citrus pests
- Forest pests
- Potato pests
- Miscellaneous pests





... and ESRI story maps

28
maps

Last
update


Online
&
mobile






Plant Pests Story Maps Gallery  

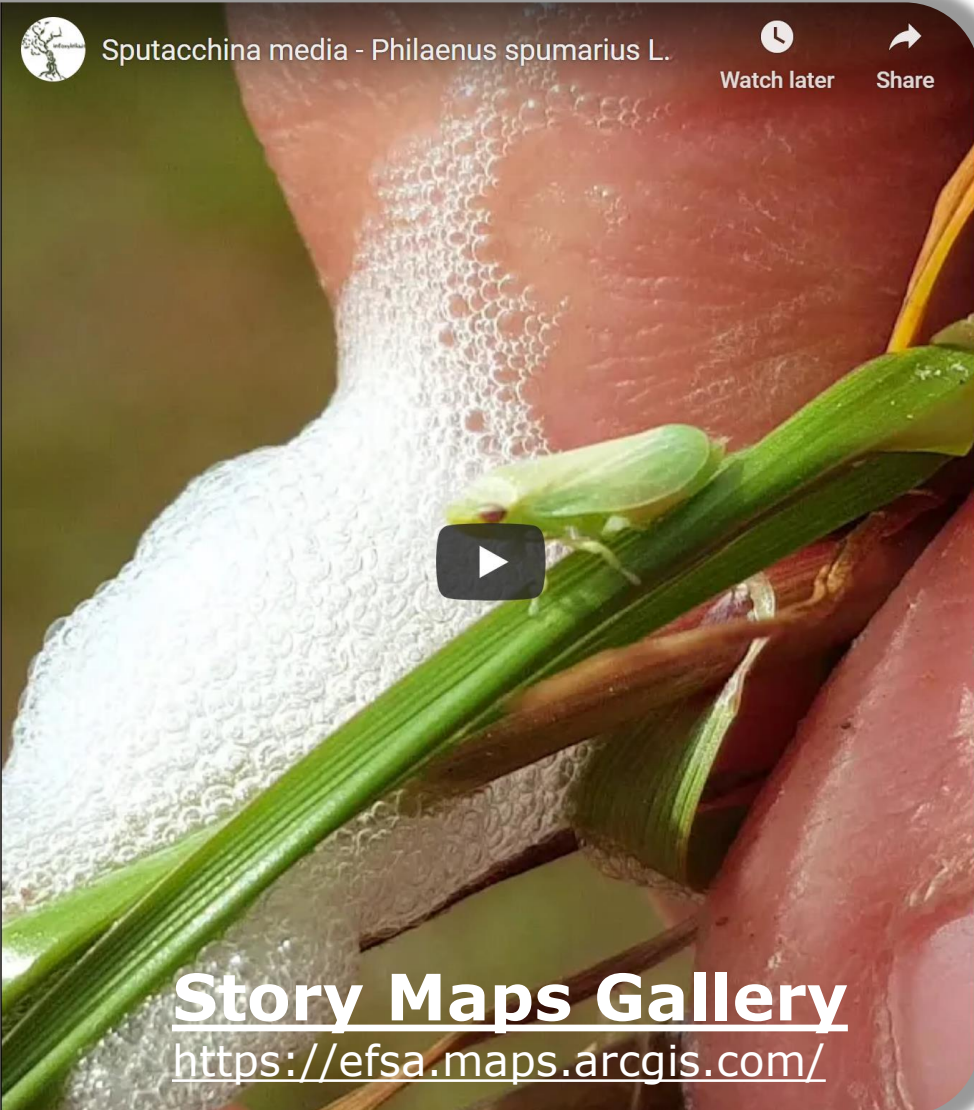
Story map for survey of *Xylella fastidiosa*

All Cicadoidea (cicadas) and Cercopoidea - such as the Aphrophoridae family, known as **frohoppers and spittlebugs** - are considered as xylem fluid feeders. Within the superfamily Membracoidea, only the insects belonging to the **subfamily Cicadellinae** (known as sharpshooters) are **xylem fluid feeders**. **Only these insects have been shown to be vectors of *X. fastidiosa*** (EFSA PLH Panel, 2015, 2018, 2019a).



Nymphs and spittle of *Philaenus spumarius*. Source: Tomasz Klejdysz, shutterstock.com

 Sputacchina media - *Philaenus spumarius* L.  Watch later  Share



Story Maps Gallery
<https://efsa.maps.arcgis.com/>



Q & A

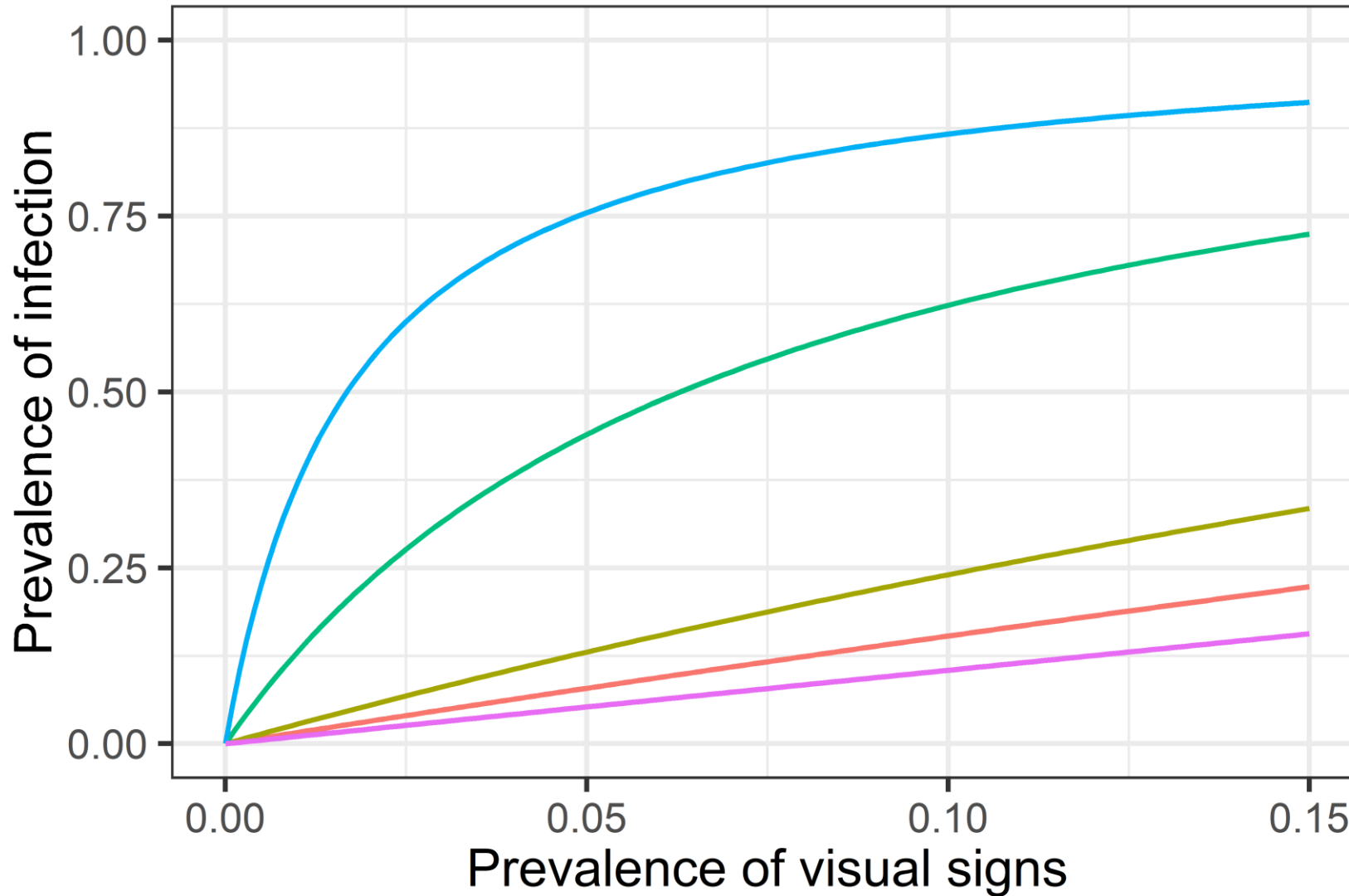
Trusted science for safe food

An aerial photograph of a rural landscape. In the foreground, there is a dense, lush green forest. To the left and slightly behind the forest, a large field is visible, containing several round hay bales. The field appears to be a mix of green grass and brown earth. In the background, there are rolling green hills with scattered white buildings, likely farmhouses or barns, and more dense forest. The overall scene is bright and clear, suggesting a sunny day.

Delimiting & buffer zone surveys

- **Key concepts for survey design**
- **Delimiting surveys**
- **Buffer zone surveys**

Detection method is key



Olive quick decline

(Xylella fastidiosa)

Huanglongbing

(Ca. Liberibacter asiaticus)

Citrus canker

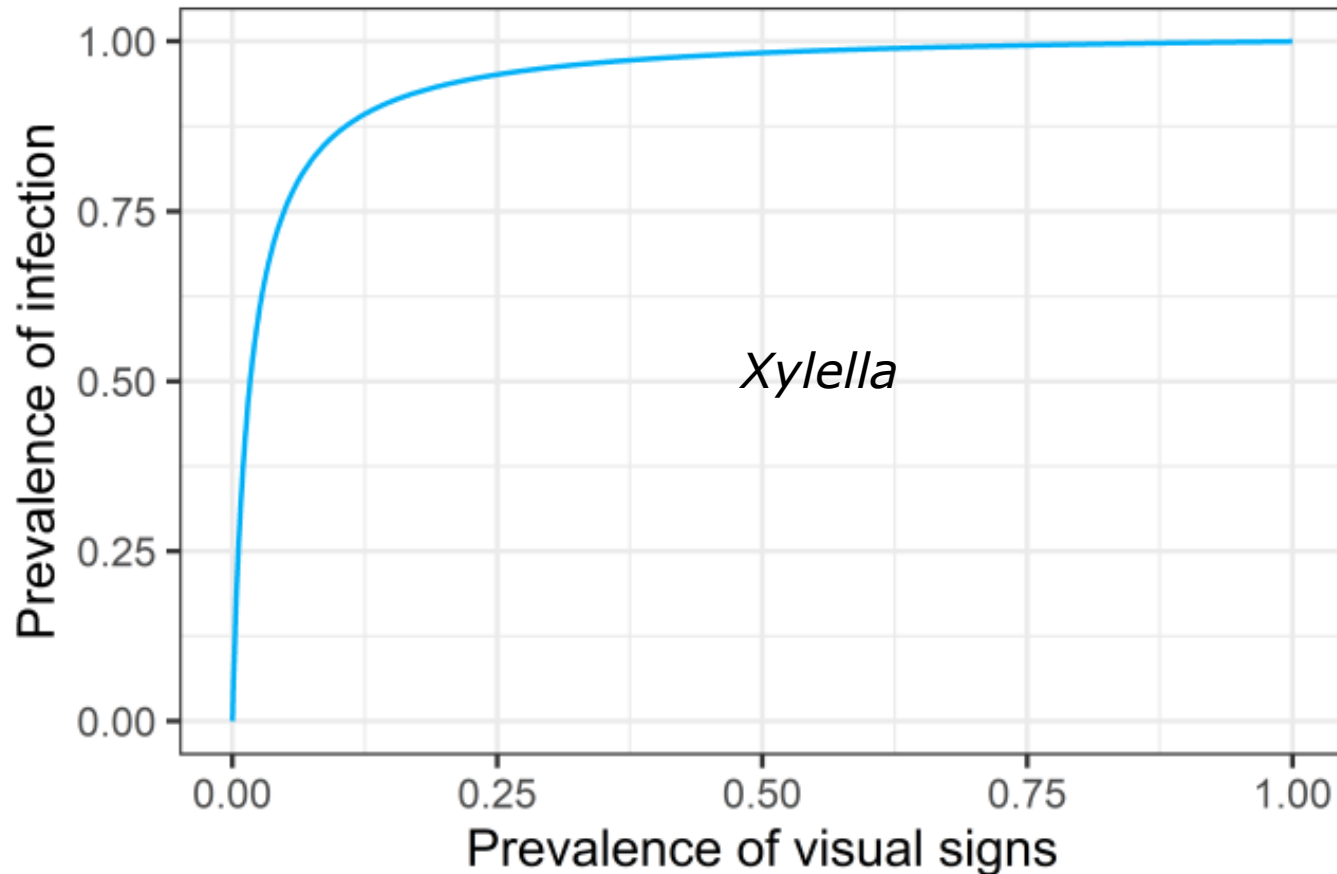
(Xanthomonas axonopodis)

Ash dieback

(Hymenoscyphus fraxineus)

Sudden oak death

(Phytophthora ramorum)



By the time a visual inspection survey first finds symptoms, the prevalence of infection can already be very high

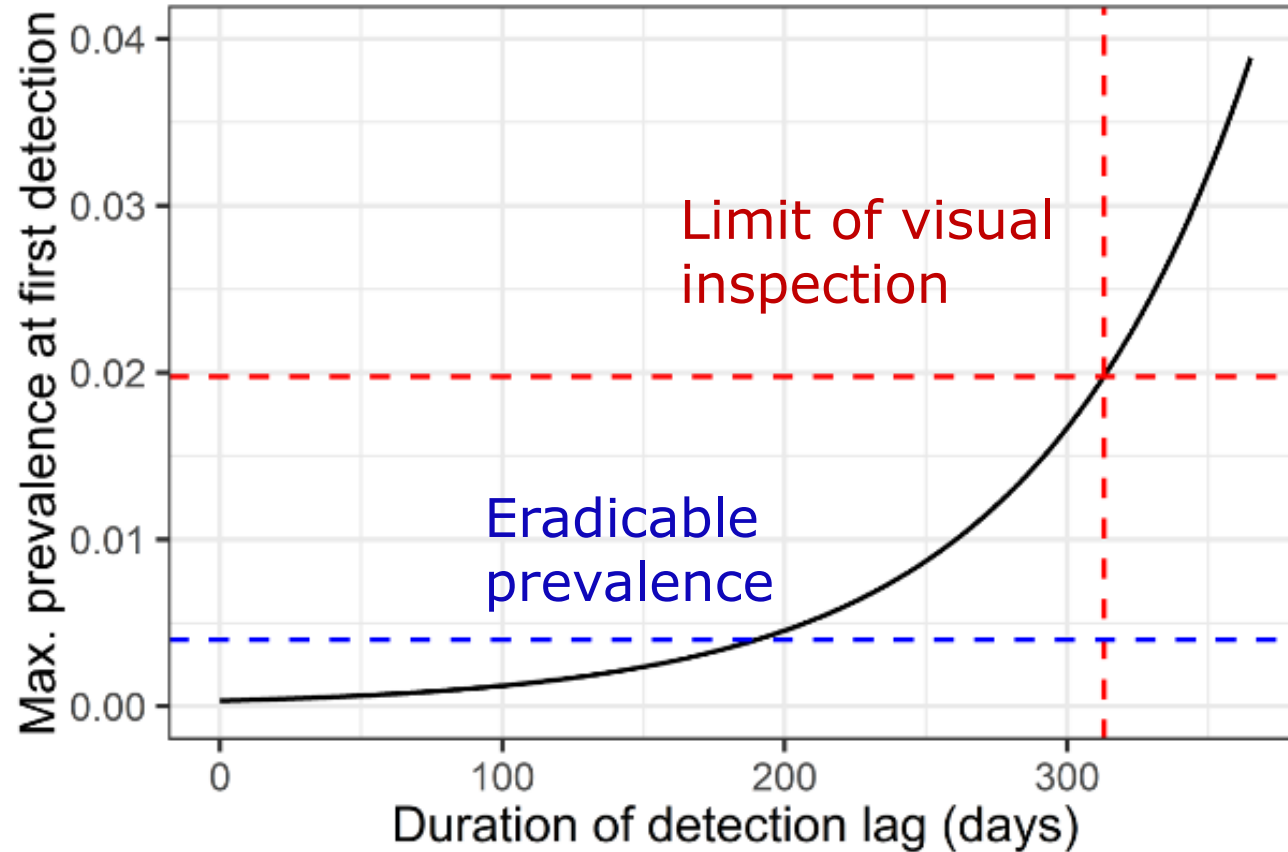
■ Method sensitivity

(efficacy of detection, ISPM 31)

- The probability to detect the pest in an individual inspection unit if it is present
- Sampling effectiveness × diagnostic sensitivity

- **Sampling effectiveness** → depends on the ability of the **inspector** to successfully choose the infected parts from a host plant.
- **Diagnostic sensitivity** → the probability that a sample tests positive when the sample is truly positive. (The lab method).

Method sensitivity



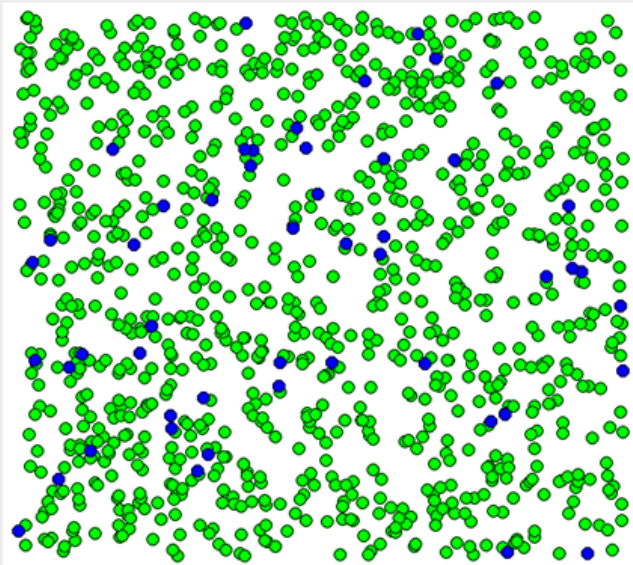
© CNR Bari

Consequently, the prevalence a visual inspection survey can detect *Xylella* (red line) is much higher than that which is considered “eradicable” (blue line)

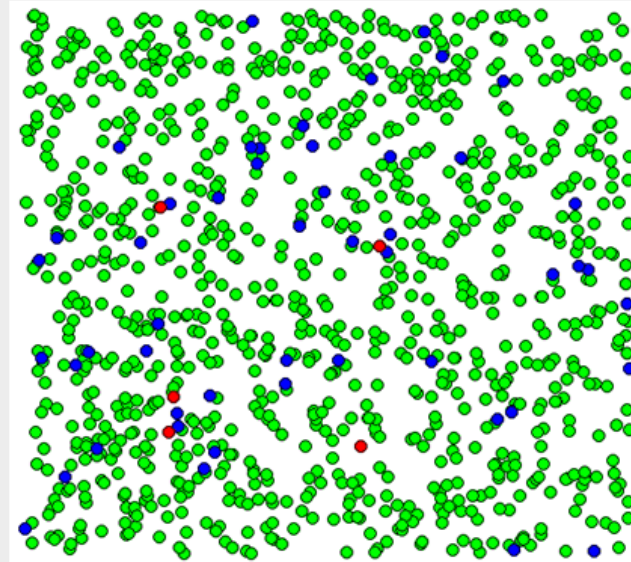
What is the evidence for pest freedom?

Suppose you conduct a survey and find no pest, what does that mean?
Is there really nothing there? How sure can we be?

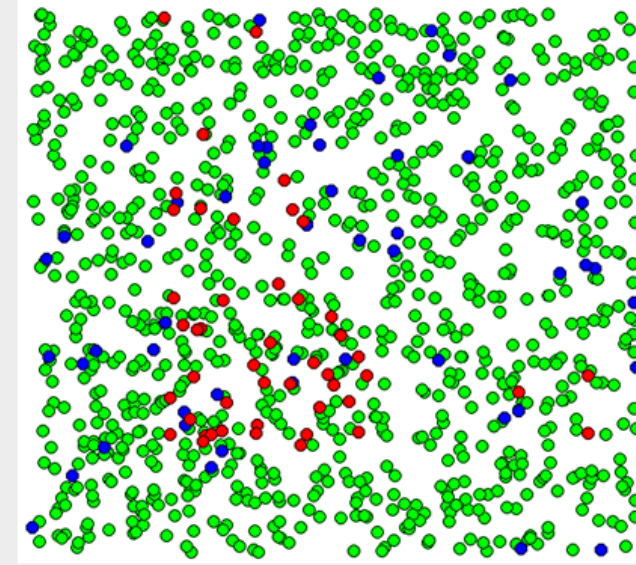
Scenario 1: **Disease-free**



Scenario 2: **Low incidence**



Scenario 3: **High incidence**



- Randomly sampled plants
- Healthy plants
- Infected plants

It is impossible to say with 100% certainty that the pest is not present.

So, what can we say?



We found no pest.
We can say with
90% confidence¹
that if the pest is
present its
**prevalence² is
below 1%**

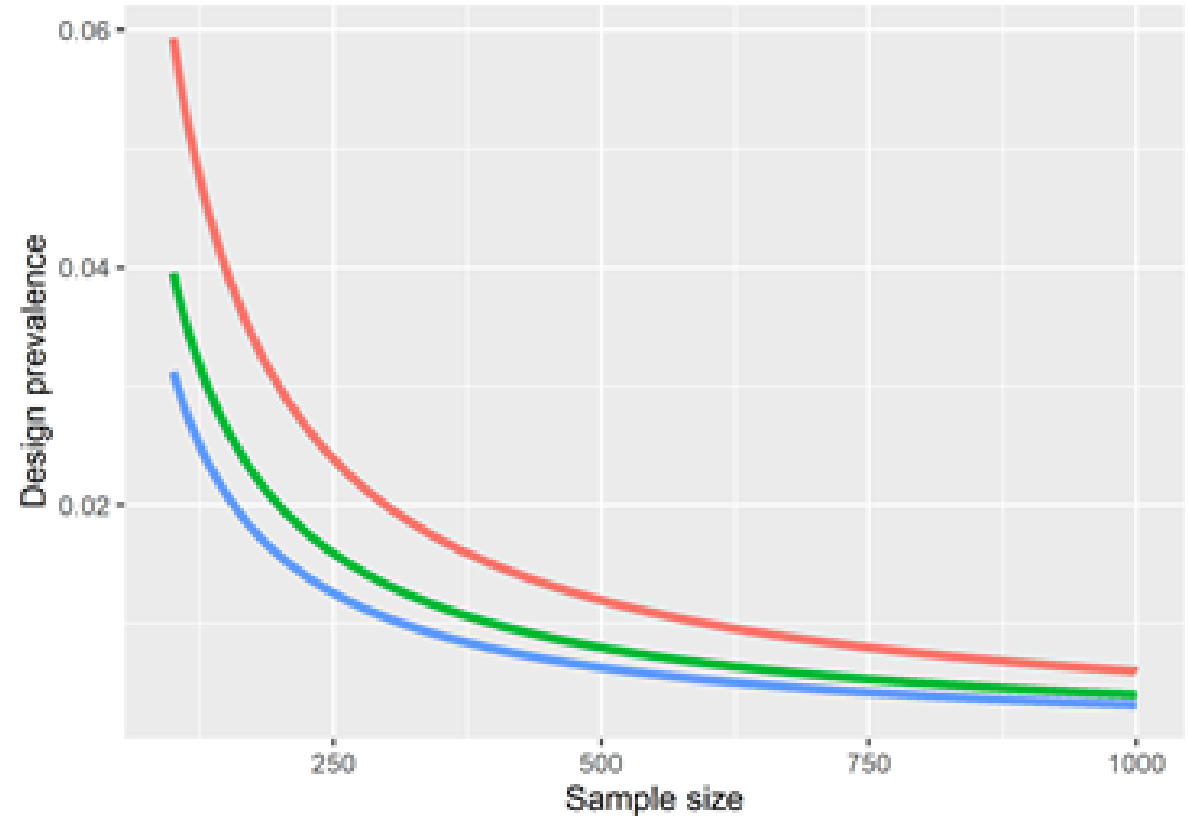
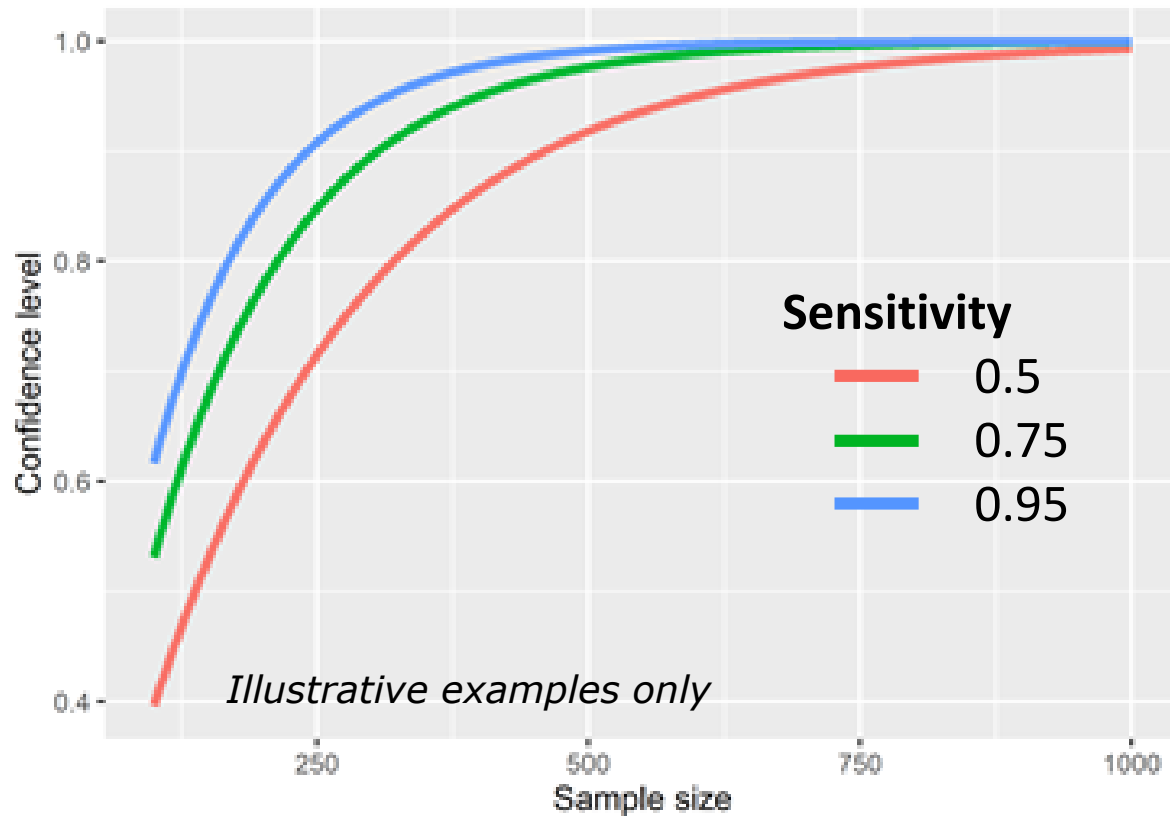
¹The **Confidence level CL** is the amount of confidence in finding the pest

²The **Design prevalence DP** (defined in ISPM 31) is the “maximum prevalence” of the pest allowed by the survey

- ➔ CL and DP depend on the aim of the survey: detection surveys and delimiting survey will have different CL and DP values
- ➔ CL and DP are set by risk managers: is a compromise between available resources and the level of risk acceptable for that specific pest

Interrelation of survey parameters

The lower the design prevalence and the higher the confidence level, the stronger the evidence for pest freedom.



Within an epidemiological unit the more inspection units that are sampled (sample size) and the higher the method sensitivity, the lower will be the design prevalence and the higher the confidence level. 35

RiBESS+: the statistical tool

A [video tutorial](#) is available via the EFSA YouTube Channel and will be played now.



How to use RiBESS+ Tutorial



REPUBLIC OF ESTONIA
AGRICULTURAL BOARD

Delimiting & buffer zone surveys

- **Key concepts for survey design**
- **Delimiting surveys**
- **Buffer zone surveys**

- I. Set survey parameters. They depend on:
 - Aims of the survey
 - Target population
 - Pest identification methods

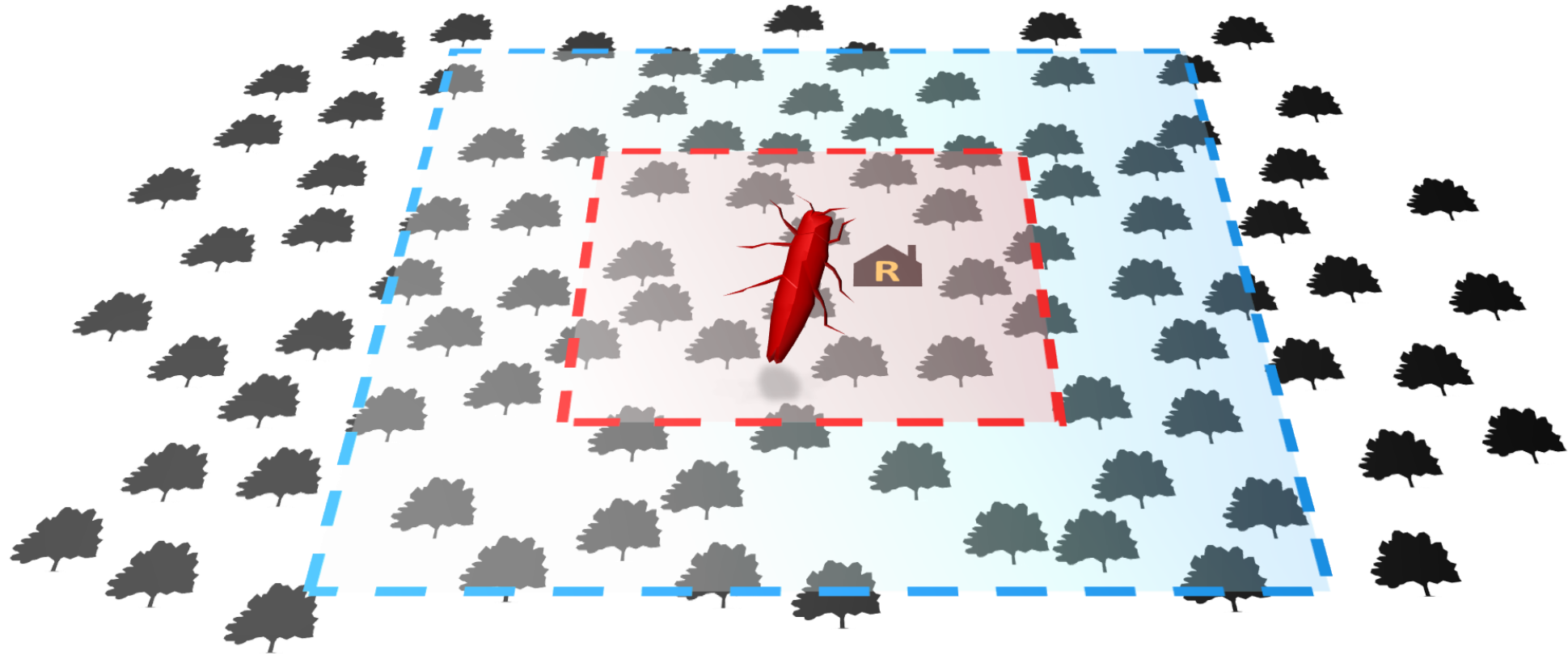
- II. Estimate the number of host plants (sample size) to be sampled (**RiBESS+**)

- III. Allocate the number of host plants to be sampled in the survey area

I. Quantify survey parameters

- Aims of the survey → **Confidence level (CL)** and **Design prevalence (DP)**
 - Detection surveys: pest freedom
 - Delimiting surveys: **infested zone boundaries**
 - Buffer zone surveys: **detection at low prevalence level**
- Host plant population → **Population size** and **Risk factors**
- Pest identification methods → **Method sensitivity (MeSe)**

The demarcated area should consist of an **infested zone** and **a buffer zone** (Regulation (EU) 2016/2031, Article 18)



- **Step 1: Identifying the source of the infestation or infection**
- Step 2: Construction of the potentially infested zone
- Step 3: Delimit the boundaries of the infested zone

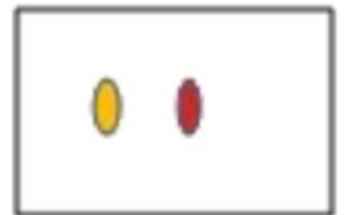
One infected host was found; no source locally identified. The infected host becomes the source of the infection.



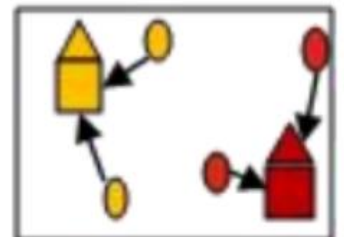
One infected host was found and a source identified.



More than one infected host was found; no sources locally identified. All infected hosts are considered as sources.

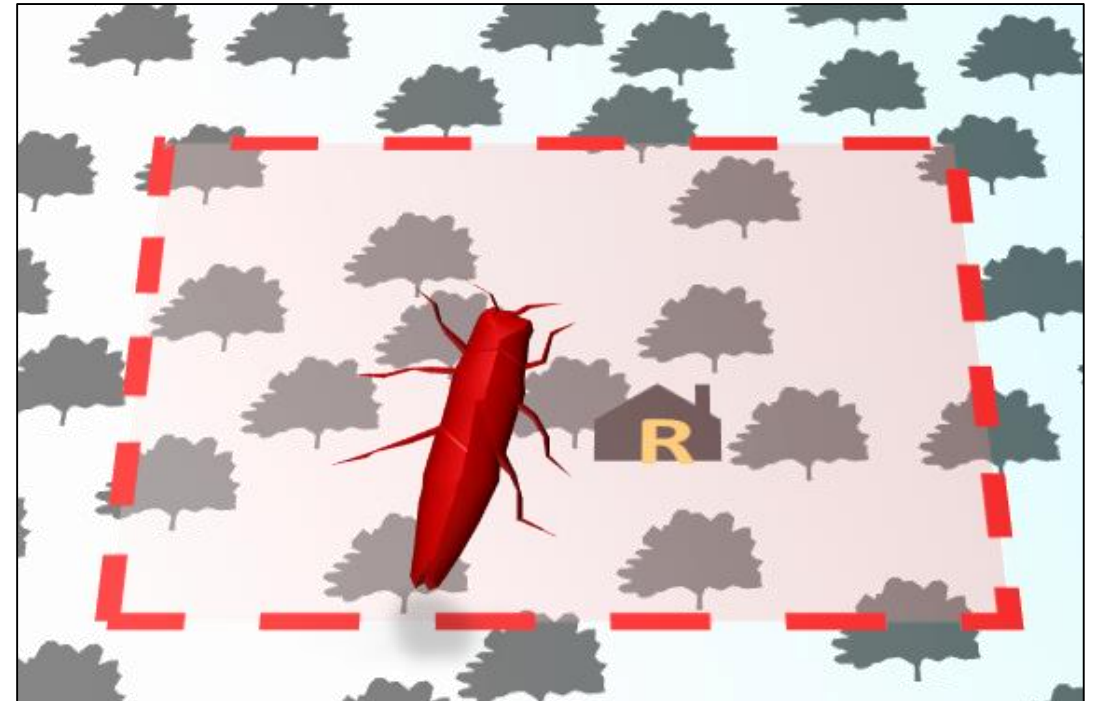


More than one source of infection was identified.



Delimiting surveys: a stepwise approach

- **Step 1:** Identifying the source of the infestation or infection
- **Step 2:** Construction of the potentially infested zone
- **Step 3:** Delimit the boundaries of the infested zone



Pest spread and survey design

Spread rates (m/yr) (99 percentile)



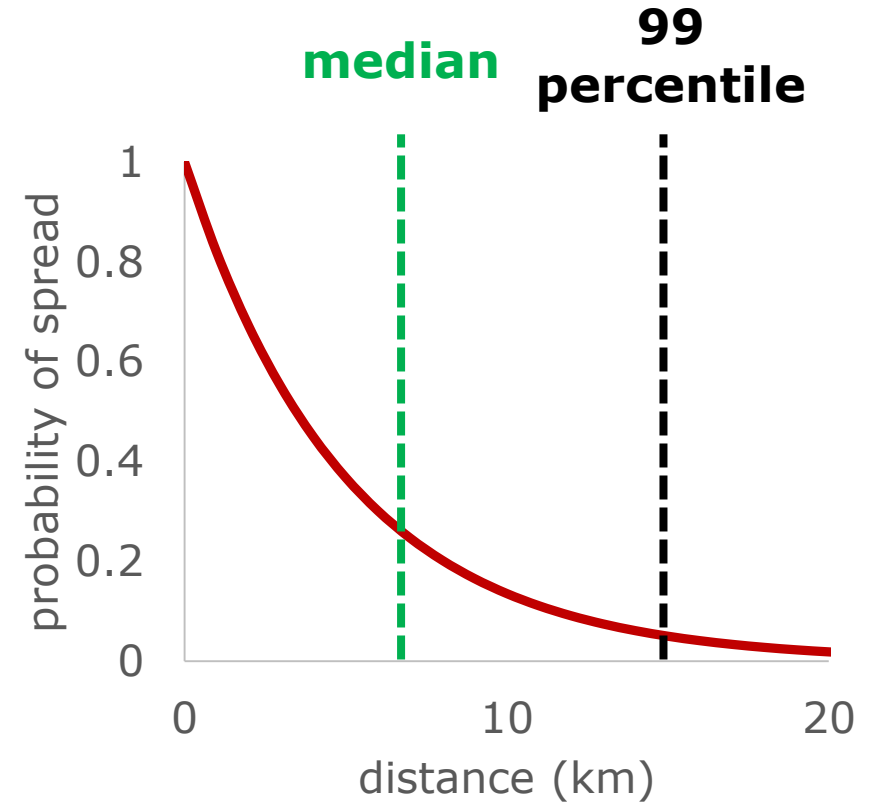
5000m



12,000m



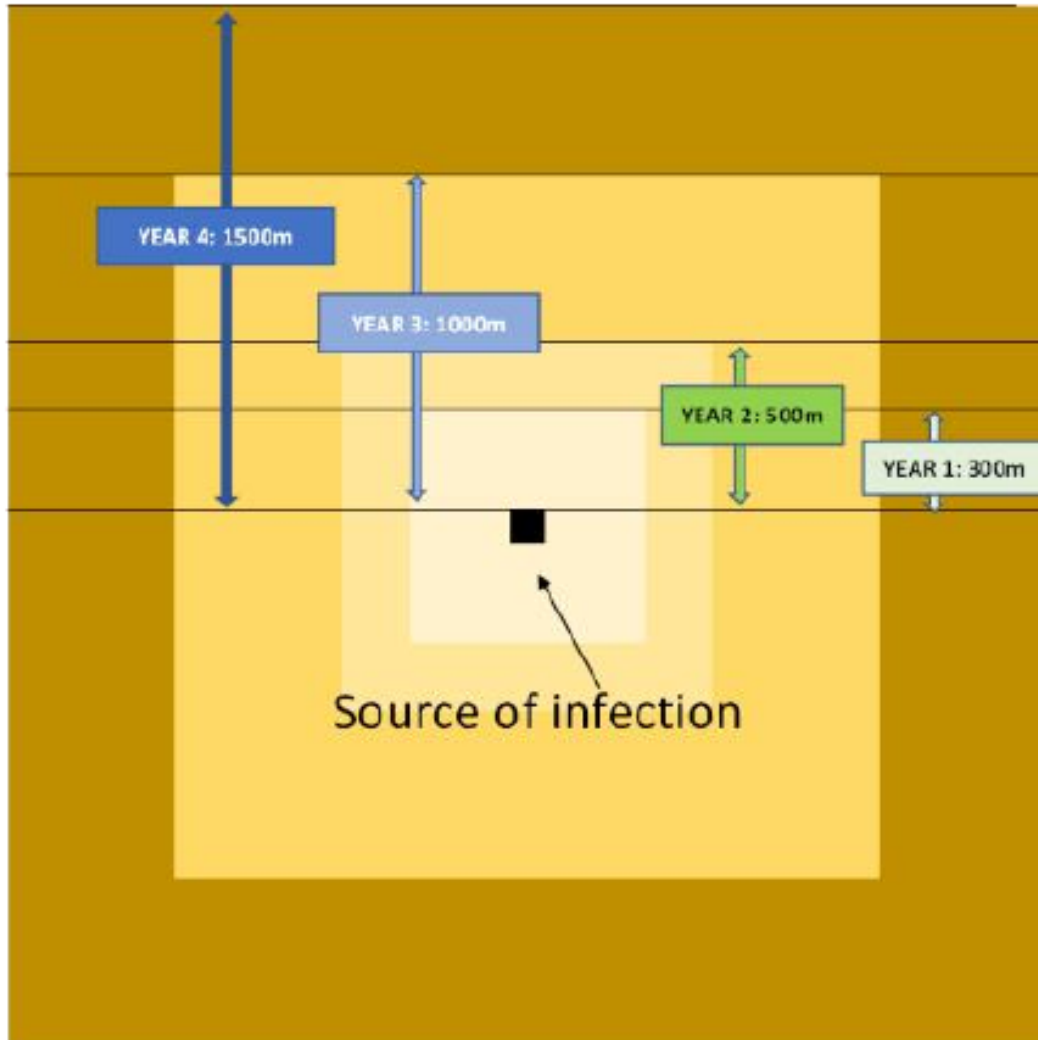
10,000m



Delimiting surveys: a stepwise approach



Step 2: Construction of the potentially infested zone



Years since last detection survey of the site	Estimated spread distance around the source of infection ^(a)
1	300 m ^(b)
2	500 m ^(c)
3 ^(c)	1000 m ^(c)
4	1500 m ^(c)

(a) The potential spread distance, from its introduction until the pest is n found, in the worse case corresponds to the years elapsed since the last detection survey was performed.

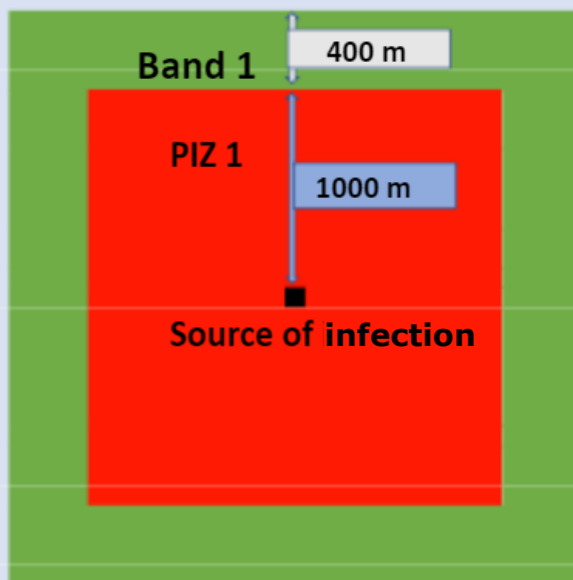
(b) Yearly median of short-distance dispersal 151 m (fitted to the spread rate in Apulia) (EFSA PLH Panel, 2019).

(c) Based on short-range spread model of the disease caused by *Xylella fastidiosa* (EFSA PLH Panel, 2019).

(d) This is the scenario chosen for the simulations.

Delimiting surveys: a stepwise approach

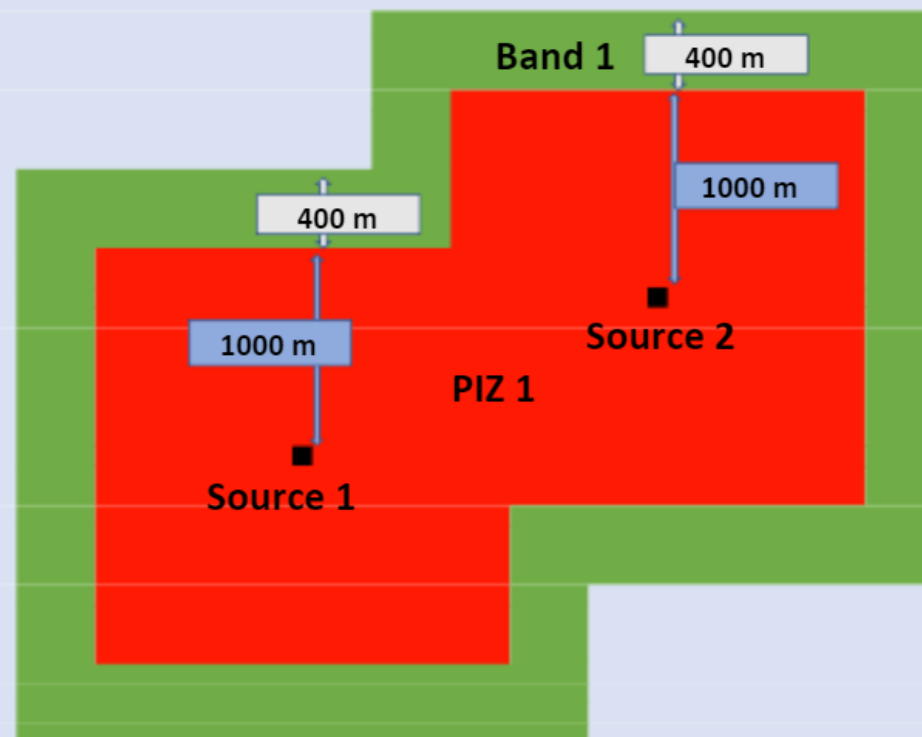
Single infection source



- Source of infection
- PIZ 1 : Potentially infested zone 1
- Band 1 : Survey band 1

Provisional demarcated area

Multiple infection sources

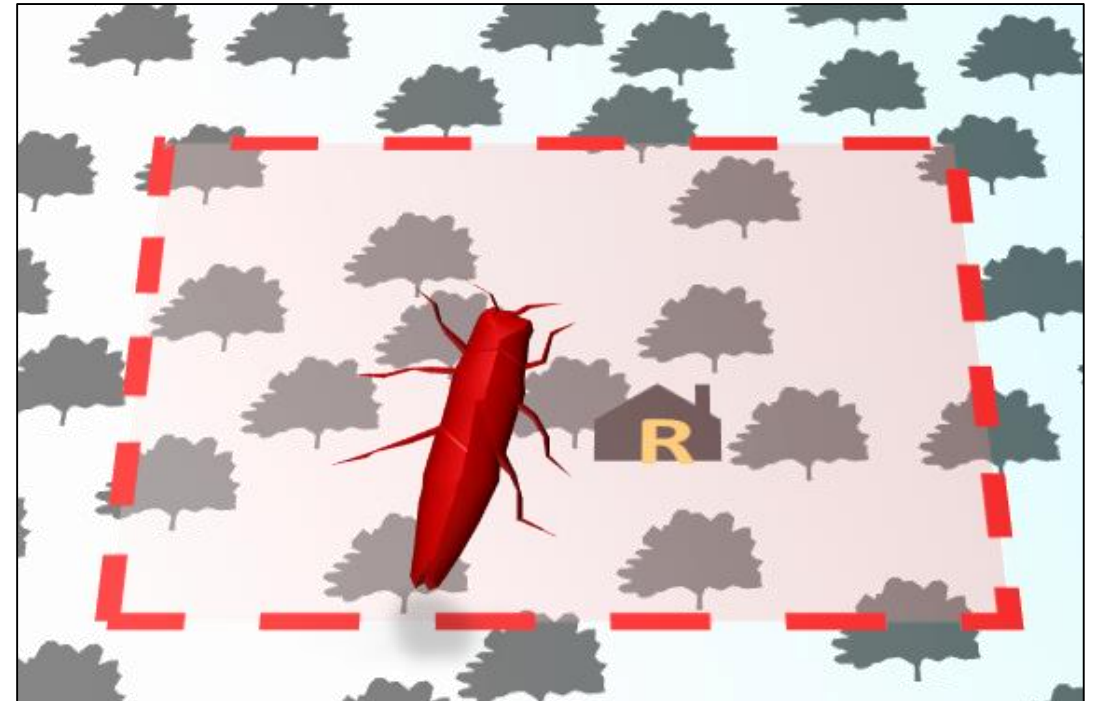


- Source of infection
- PIZ 1 : Potentially infested zone 1
- Band 1 : Survey band 1

Provisional demarcated area

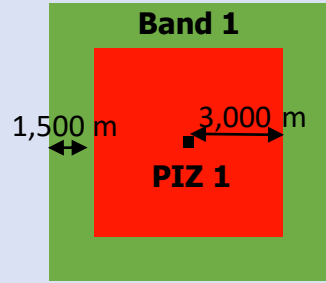
Delimiting surveys: a stepwise approach

- **Step 1:** Identifying the source of the infestation or infection
- **Step 2:** Construction of the potentially infested zone
- **Step 3:** Delimit the boundaries of the infested zone

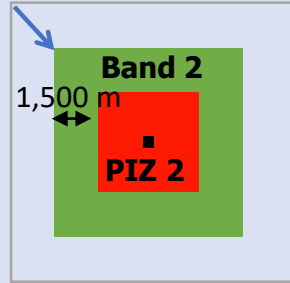


Narrowing down the provisional infested zone

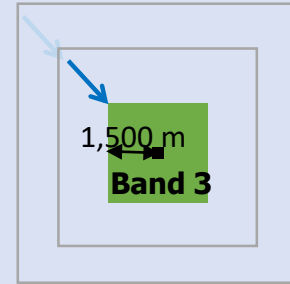
Provisional demarcated area



Band 1 cleared



Band 2 cleared

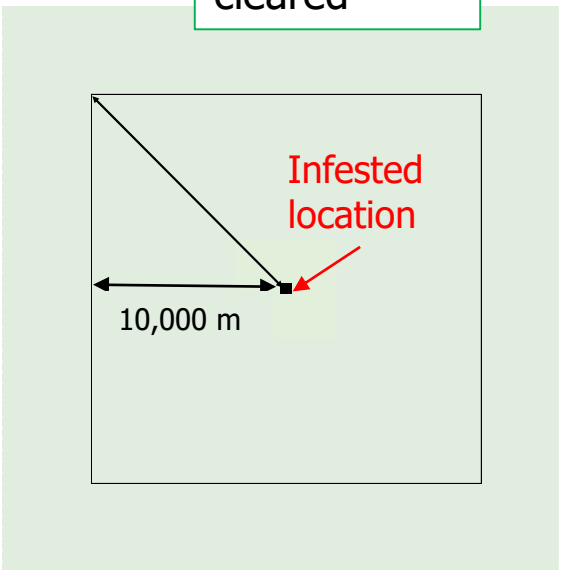
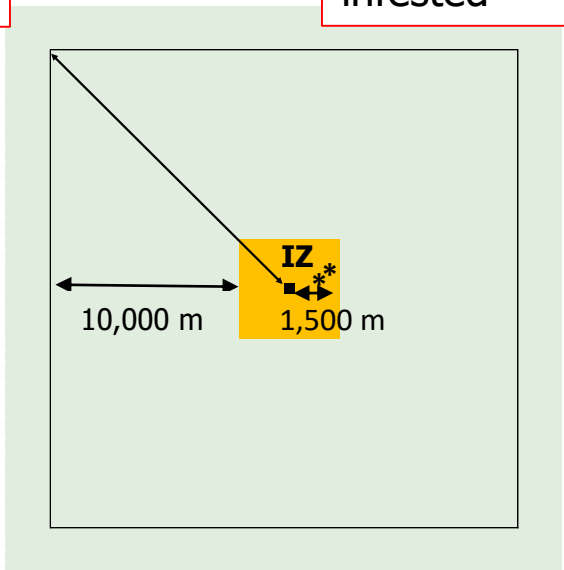
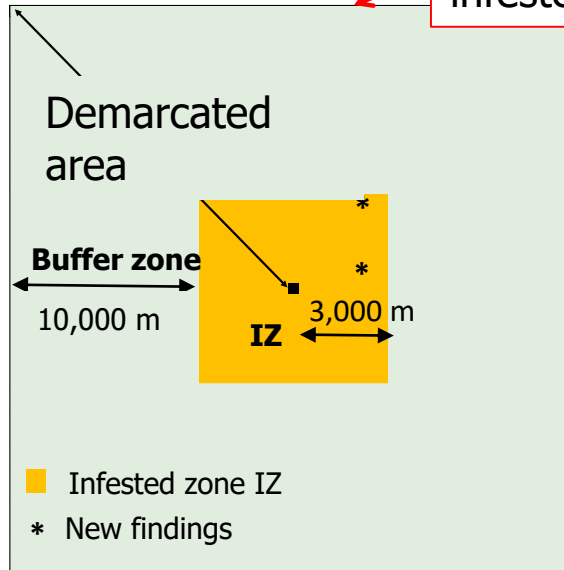


- Source of infestation
- Provisional infested zone = PIZ
- Survey band

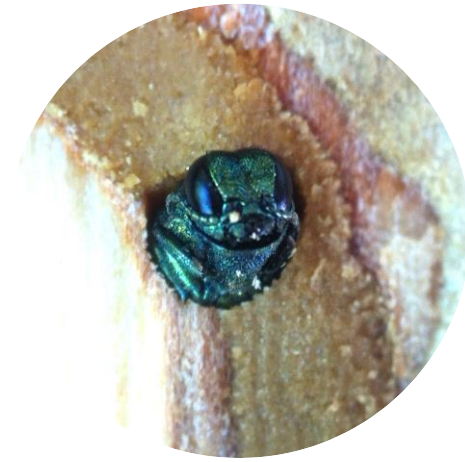
Band 2 is infested

Band 3 is infested

Band 3 is cleared

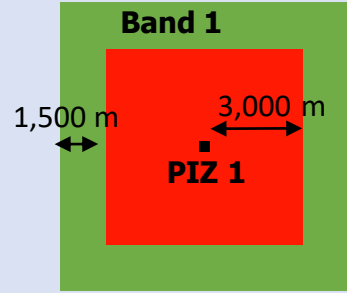


Boundaries of the infested zone confirmed and area demarcated



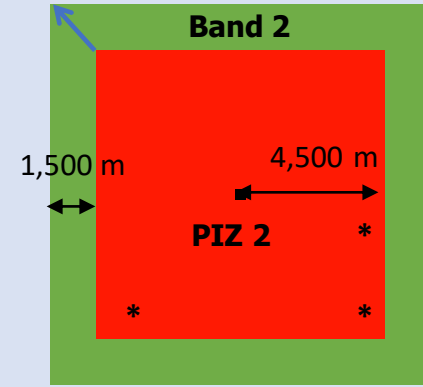
Enlarging the provisional infested zone

Provisional demarcated area



- Source of infestation
- Provisional infested zone = PIZ
- Survey band

Band 1 infested



* New findings

Band 2 infested

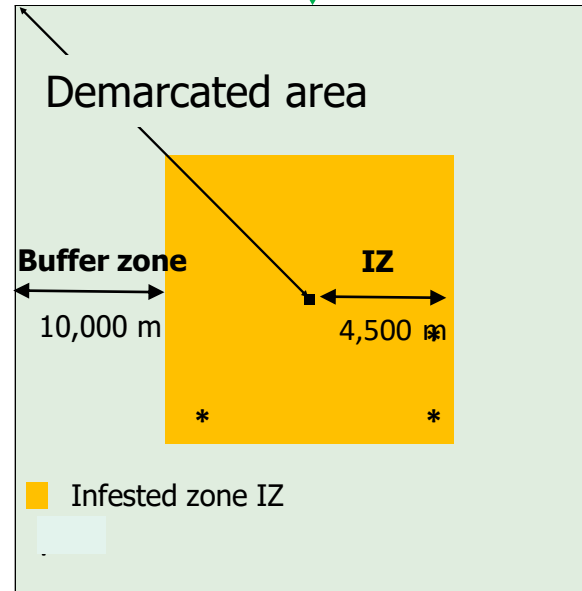
Iterative process following positive findings in the newly defined survey bands



Band 1 cleared

Band 2 cleared

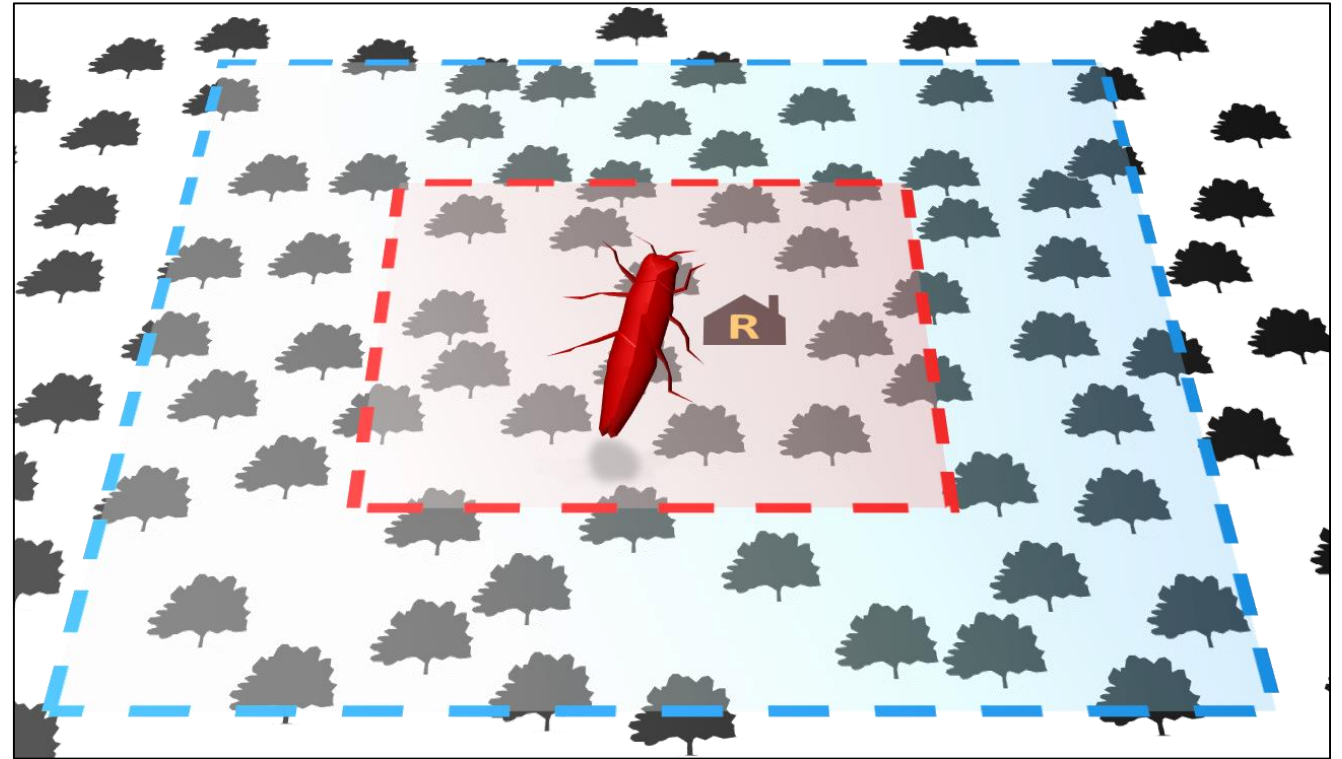
Narrowing down of the PIZ 1 following the process described in Step 3a



Boundaries of the infested zone confirmed and area demarcated

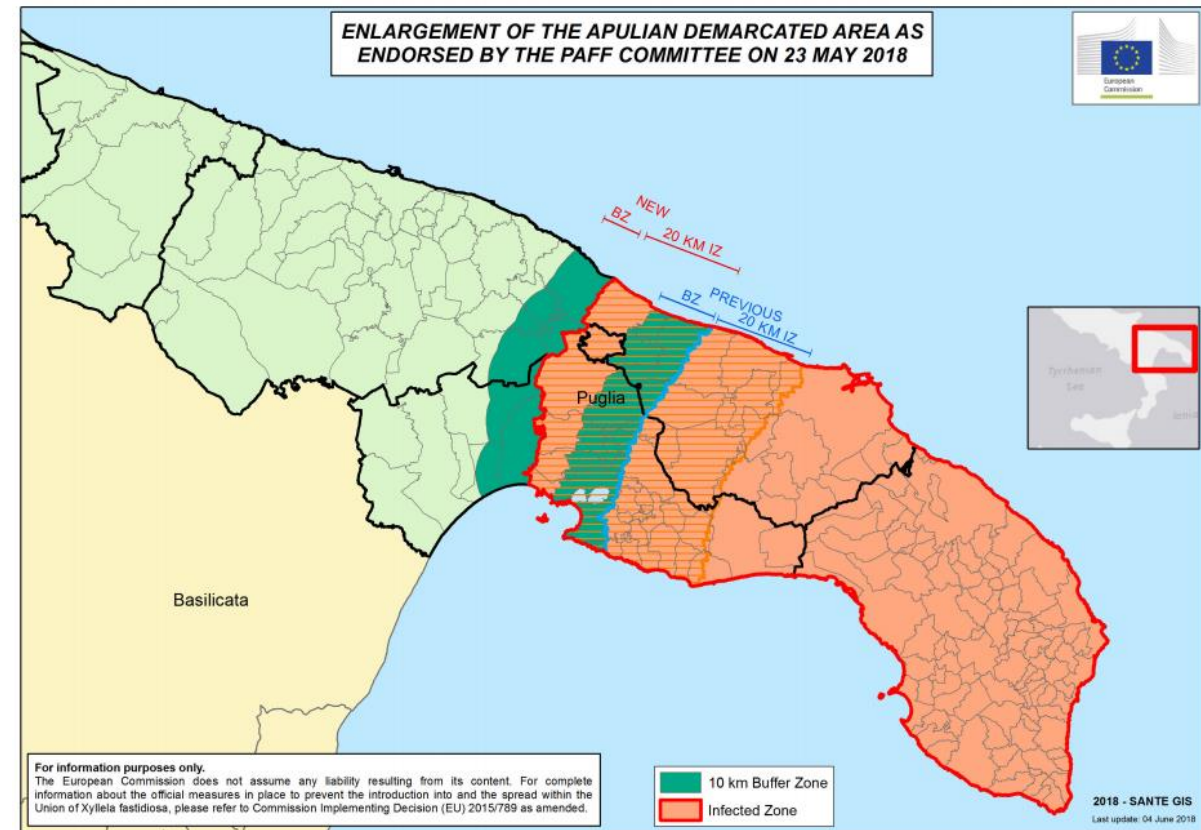
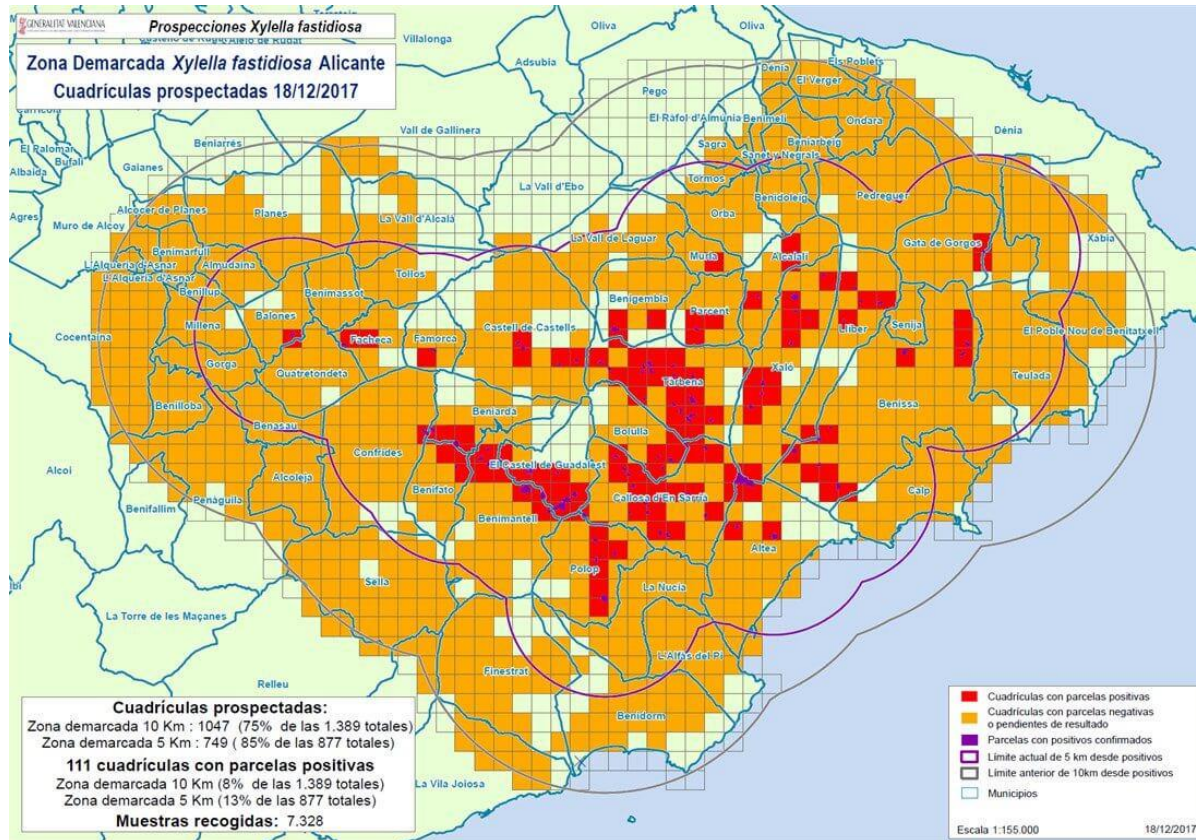
A buffer zone is *“an area surrounding or adjacent to an area officially delimited for phytosanitary purposes in order to minimize the probability of spread of the target pest into or out of the delimited area, and subject to phytosanitary or other control measures, if appropriate”*

ISPM 5 (FAO, 2019)

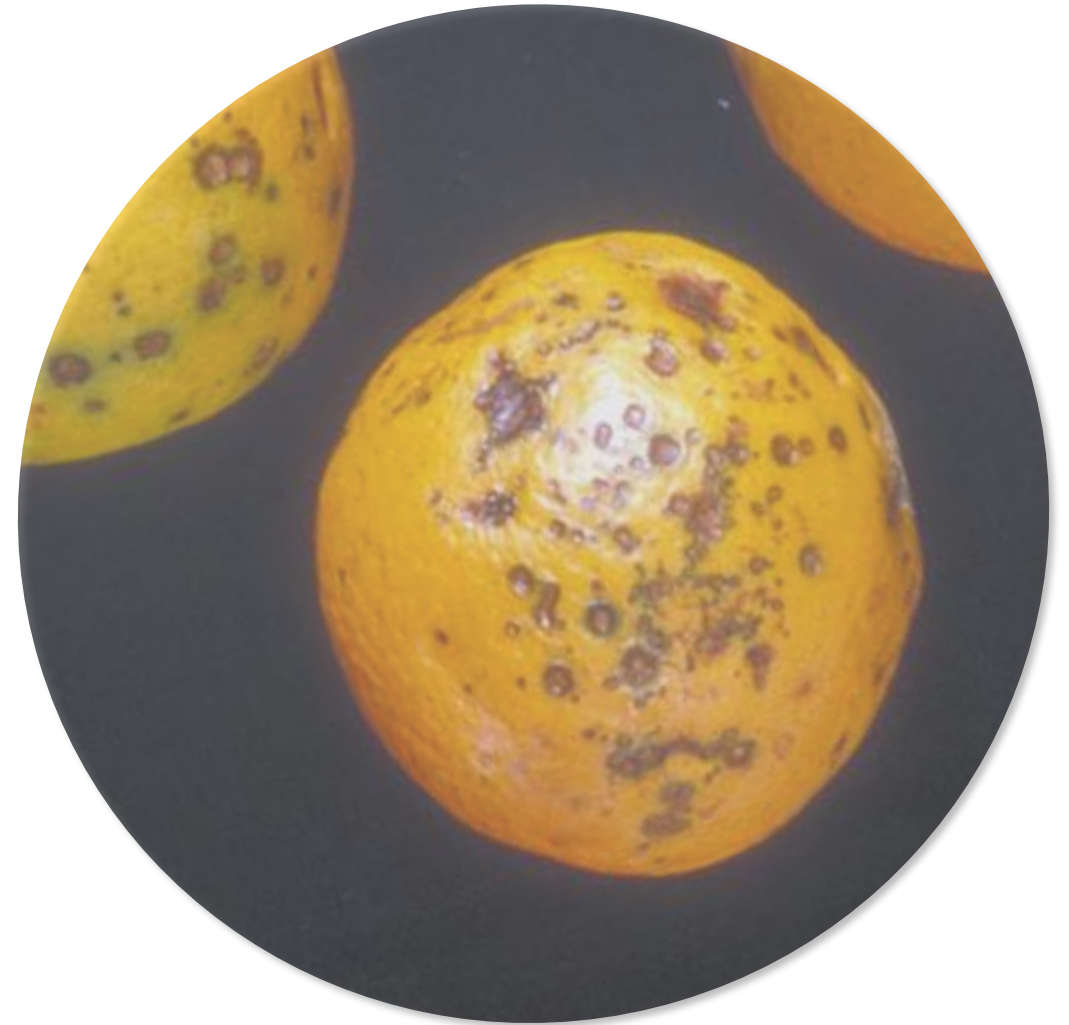
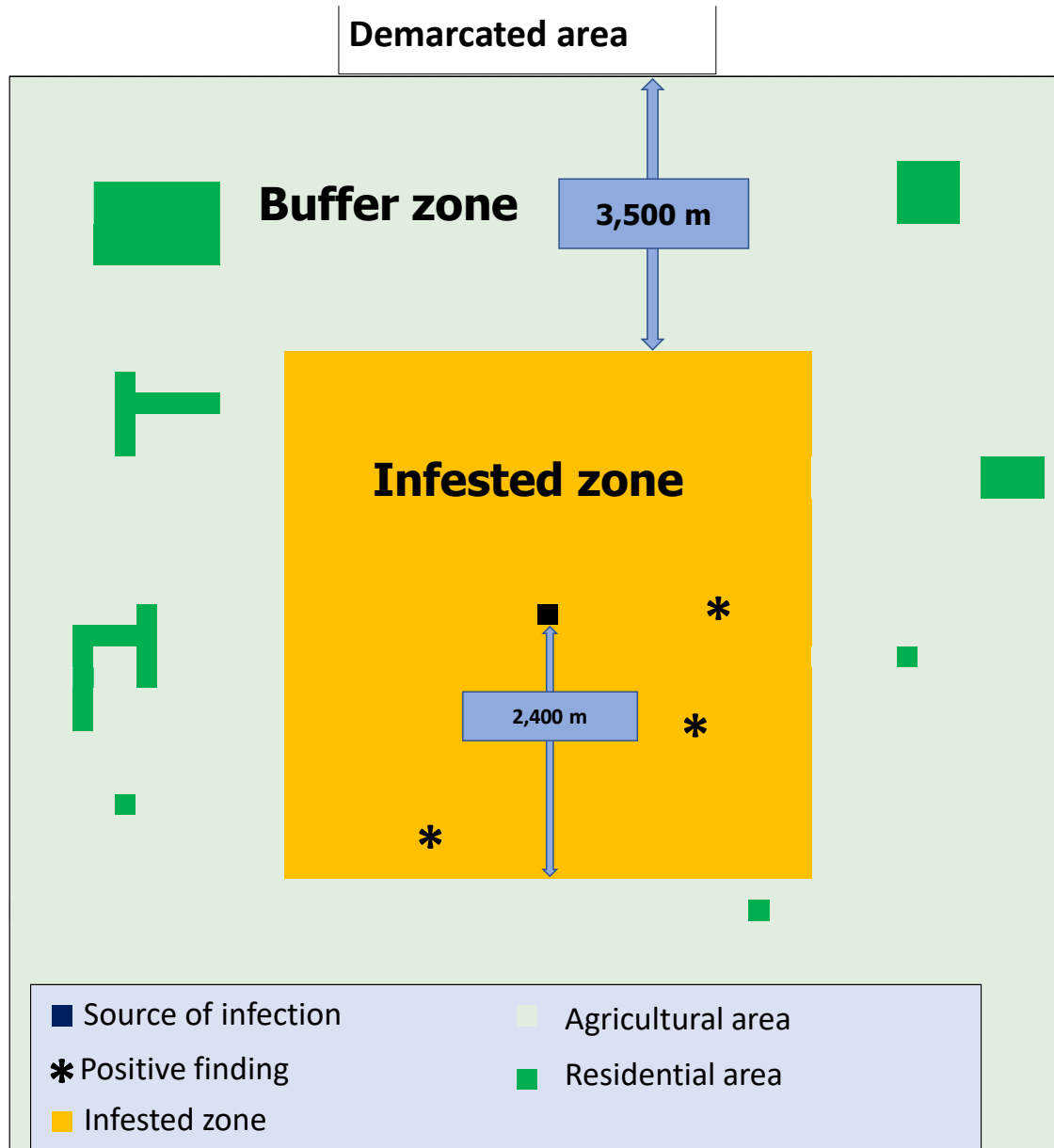


Buffer zone surveys

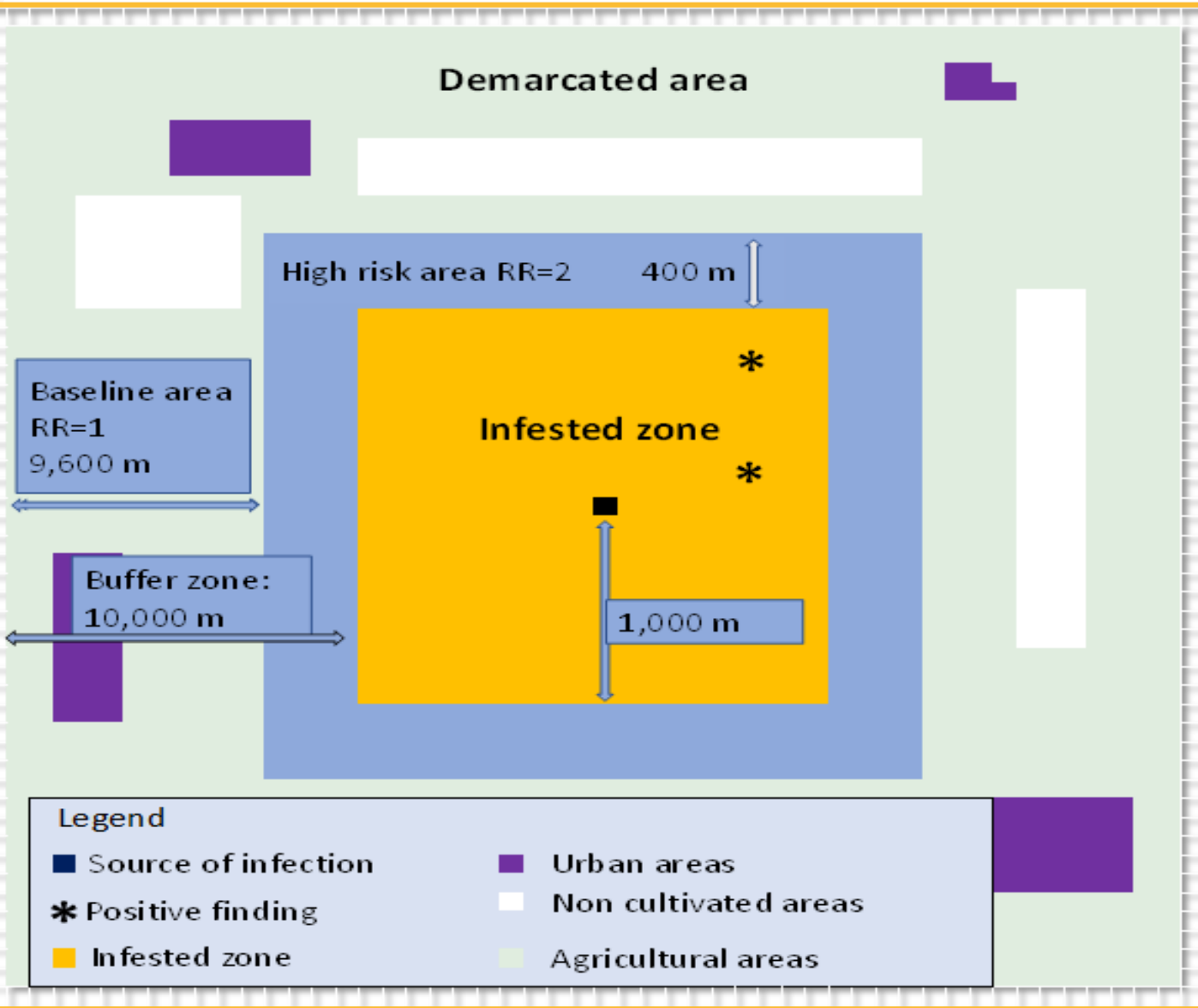
Examples of buffer zones as shown for *Xylella* for Valencia and Apulia



Citrus black spot



Xylella fastidiosa



Example: calculation

SURVEY PARAMETERS		AGRI AREAS	URBAN AREAS	
Goal of the survey	Confidence level (CL)	0.78	0.78	
	Design prevalence (DP)	0.04%	0.1%	
Target population	Host plants	<i>Prunus sp.</i> <i>Vitis sp.</i>	<i>Lavandula sp.</i>	
	Size	7.5 million host plants	1.2 million host plants	
	Extent	25,000 ha	12,000 ha	
	Epidemiological units	Whole extent	Whole extent	
	Risk factor	High risk (24,600 ha) RR=2	120,000 host plants (0.016)	-
		Baseline (400 ha) RR=1	7,380,000 host plants (0.984)	-
Identification methods	Method Sensitivity (MeSe)	0.55	0.55	

Result: N of samples needed

LAND USE	DESIGN PREVALENCE (%)	CONFIDENCE LEVEL (%)	RISK LEVEL	RR	CONVENIENCE SAMPLING	SAMPLES	
Agri area	0.04	78	High risk	2	2	2,784	
			Baseline	1	1	1,392	
Urban area	0.04	45	N/A	N/A	N/A	2,751	
UNIQUE DP		$OCL = 1 - (1 - CL_{AA}) \cdot (1 - CL_{UA})$					
Total	0.04	$1 - (1 - 0.78) \cdot (1 - 0.45) = 0.8799 \rightarrow 87.99\%$					6,927



In conclusion



Q & A

Trusted science for safe food

What is next...

Survey cards for >200 pests in 6yr

- Quarantine, protected zone, and emerging pests
- From pest-based to crop-based survey
- Plant health specific stats tool

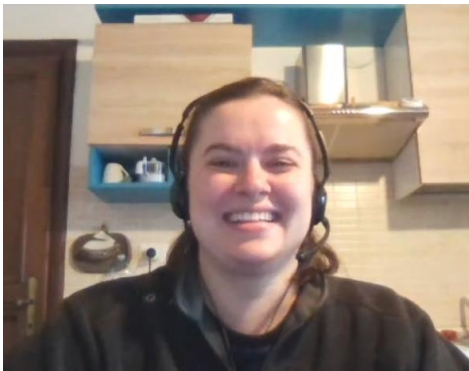


- **6, 21 October and today's webinar available online**

Check for:

- **New survey cards and guidelines in the EFSA journal**
- **New story maps in the gallery**

Thanks for attending!



EFSA surveillance

- **Staff:** Sybren Vos, Giulia Mattion, Alice Delbianco, Ignazio Graziosi, Jose Cortiñas Abrahantes, Gabriele Zancanaro
- **Experts:** Elena Lazaro, Antonio Vicent Civera, Stephen Parnell
- **Partners:** Netherlands Food and Consumer Product Safety (NVWA); Julius Kühn-Institut Federal Research Centre for Cultivated Plants (JKI); HORT@: Maria Chiara Rosace (story maps)
- **Thanks** to NPPOs of MSs for suggestions in the development of survey cards and guidelines

We also thank Alzbeta Mikulova, Sara Tramontini, the EFSA Plant Health team, and EFSA Comms

Thanks for attending!




Please feel free to reach out at:
alpha@efsa.europa.eu

Please take 5 more minutes to [fill out the evaluation form](#) that you will receive shortly. Your feedback will help us improve our work!

Thanks for attending!

New PLH website
<https://bit.ly/3dtyyqm>



Pest Surveillance

EU Member States must be prepared to meet the increasing threat posed by plant pests. To help them plan their pest surveys, EFSA provides a toolkit that includes pest survey cards, survey guidelines, and statistical tools. The aim is to assist national authorities in carrying out plant pest surveys in their territories and to harmonise surveillance methods across the EU.

EFSA is also developing pest story maps updating the pest survey cards, an easy-to-use digital tool that makes it easier to navigate through the documents.

[Explore the toolkit](#)

[Pest story maps](#)



INTERNATIONAL YEAR OF
PLANT HEALTH
2020







Plants EFSA
@Plants_EFSA

[Follow](#)

The EU hub for information on assessment and emerging risks on [#plants](#). Account managed by the [@EFSA_EU](#) [#PlantHealth](#) team.

On Twitter:
#PlantHealth
#IYPH2020
@Plants_EFSA